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Sliding further into scientific reality. 40 new species and 14 new subspecies within the Australian skink assemblage known as *Lerista* Bell, 1833 *sensu lato*.

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ABSTRACT

A review of the Australian skink genus *Lerista* Bell, 1833 *sensu lato* by Hoser published in 2024 resulted in the following changes. Division into 24 genera was formally proposed. Eight, including the most divergent clade were formally named for the first time.

The arrangement of the definitive work of Wells (2012) was largely, but not entirely, correct and broadly adopted as the correct taxonomy.

Hoser in 2024 made appropriate changes, including the merging of three Wells (2012) recognized genera into others and reassigned some species from the genus-level placements of Wells (2012).

19 species and 5 subspecies were also formally named for the first time.

"*Lygosoma walkeri* Boulenger, 1891", had a Lectotype designated in accordance with the rules of the *International Code of Zoological Nomenclature* (Ride *et al.* 1999) ("the Code").

Hoser in 2024 also appropriately dealt with acts of taxonomic vandalism and egregious misconduct by Glenn Shea.

The overall effect of the Hoser 2024c paper was to unambiguously stabilize the taxonomy and nomenclature of this large group of well over 100 species by bringing the genus-level classification and divisions in line with that of other Australian lizard groups.

Clades generally 10 MYA or more divergent were placed in separate genera making 24 in all.

This paper adds to the work of that of Hoser in 2024 by naming 40 additional species and 14 subspecies within this group.

At the species level, it is important that all Australian species of lizard are formally identified and named as soon as practicable. Furthermore, by naming an obvious overhang of previously unnamed species and divergent subspecies, this objective is being completed in a timely manner.

In the absence of discovery and identification, governments, NGO's and other stakeholders cannot possibly conserve or manage the relevant entities.

Keywords: Taxonomy; nomenclature; lizard; skink; Australia; *Aaah*; *Acdc*; *Ah; Aphroditia*; *Go*; *Gaia*; *Miculia*; *Lerista*; *Oh*; *Rhodona*; *Telchinoscincus*; *Tism*; *aericeps*; *asgicondi*; *bipes*; *greeri*; *griffini*; *ips; oomph*; *taeniata*; *timida*; *vermicularis*; new species; *thunderstruck*; *tnt*; *yess*; *pfuk*; *sheat*; *harriganorum*; *amazing*; *anangu*; *aviyemenii*; *dezifreeman*i; *liesarenotscience*; *nomenclaturematters*; *nottobemissed*; *otit*; *rate*; *ruleofpriority*; *taxonomymatters*; *narcissismawareness*; *owe*; *questionthenarrative*; *robwatsoni*; *truthisscience*; *vellai*; *wow*; *forit*; *chrismartini*; *coldplayorum*; *puseyi*; *vacopoulosae*; *abscondeetus*; *dorsei*; *faaarkinelle*; *gottabekidding*; *greatfind*; *laylawangae*; *lopezi*; *overlookedit*; *oxyi*; *wholeeecowe*; *calistaclementsae*; new subspecies; *gotit*; *hellyeah*; *badimaya*; *nhanta*; *howzat*; *nanda*; *yinggarda*; *kalbarriensis*; *ah*; *har*; *faaark*; *sheeta*; *bestoff*; *whatareya*;

INTRODUCTION

The genus *Lerista* Bell, 1833 as originally conceived was a genus of Australian skinks.

All are fossorial in nature, small in size, have limbs that are reduced in some form and an elongate body form as befits species that burrow in loose ground.

By end 2024, the genus as recognized by most Australian herpetologists had just under or just over 100 recognized species occurring across Australia.

They vary greatly in form and even when convergent in key characteristics, repeated molecular studies have commonly shown taxa to be quite distantly related.

Phylogenies published have all shown *Lerista* Bell, 1833 as a "monophyletic group", but within this assemblage, divergence of many major stems happened between 10 and 15 million years ago.

With this in mind and relying mainly on morphological traits rather than genetic data, Wells (2012) produced a massive 361-page monograph on the genus *Lerista sensu lato*, in which he erected 11 new genera. This was in addition to the three others created by Wells and Wellington (1984 and 1988) and another 5 resurrected from synonymy, meaning 19 in total for the group, meaning an average of just over 5 species per genus and noting the existence of further undescribed forms.

This gave the following generic arrangement. *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells *2012,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005, *Wondjinia* Wells 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) with various morphologically alike species being placed in the relevant groups.

The response to the meticulous and detailed 361-page work of Wells (2012) was swift.

The Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang of thieves, published a diatribe and circulated it globally by way of SPAM email.

This document was originally known as Wüster (2012) but later rebadged and published in hard copy.

This became known as Kaiser *et al*. (2013) as cited herein, although the real authorship was Wolfgang Wüster.

We know this because when shopping the same document for authors in 2012, Kaiser in his SPAM email pointed out that he played no part in authoring the document.

Without a shred of evidence or any direct knowledge of the relevant species (versus a lifetime's work on them by Richard Wells and his collaborators), Wüster (2012) advocated 1/ Non-use of any Wells (2012) names and 2/ Further urged readers inclined to agree with any of the scientific findings of Wells to simply steal his work, rename the very same entities in breach of Copyright laws and the *International Code of Zoological Nomenclature* (Ride *et al.* 1999) and to then 3/ Claim "name authority" for the same discoveries.

Kaiser *et al.* (2013) later mutated into a number of updated publications, ultimately settling on Rhodin *et al*. (2016), which was a petition to the International Commission on Zoological Nomenclature (ICZN) to formally squash the names of myself, Raymond Hoser in all my scientific publications post-dating year 2000 and also scooping up the Wells papers postdating year 2000 at the same time.

This clearly included Wells (2012) and all the names proposed therein.

Now if the science of Wells (2012) was rubbish, his proposed names could easily by synonymised under the ICZN Code and that would have ended the matter.

The problem for the Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang of thieves was that the science of Wells (2012) was robust and as a result the bulk of his names were valid and needed to be used. They also had priority over anything Wüster and has gang may choose to rename, making their later names "invalid" and merely synonyms.

The Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang of thieves through the website they control, Peter Uetz's "The Reptile Database" made it known that the Wells (2012) names should not be used by their followers and if the taxonomy was actually ever agreed to, then the names were to be over-written with the Wüster gang's names.

Ultimately the attempt of the Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang of thieves to rewrite history failed and in terms of *Lerista*, the taxonomy and nomenclature of the somewhat oversized putative genus had in effect languished in effective suspended animation since.

On the Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang of thieves-controlled website, Peter Uetz's "The reptile database" at: https://reptile-database.reptarium.cz/species?genus=Lerista&species=lineata&search_param=%28%28search%3D%27Lerista%27%29%29

Is written:

"*Synonymy: Kaiser et al. 2013 considered the generic names Alcisius Wells 2012, Aphroditia Wells 2012, Cybelia Wells 2012, Gaia Wells 2012, Goldneyia Wells 2012, Krishna Wells 2012, Lokisaurus Wells 2012, Marrunisauria Wells 2012, Spectrascincus Wells 2012, Tychismia Wells 2012, Wondjinia Wells 2012 invalid and rejected their use instead of Lerista*."

Of course, there was not a shred of scientific evidence to counter the 361 pages of hard evidence published by Wells (2012). This is not really out of the ordinary as neither Uetz, Kaiser or Wüster or anyone else in their gang of thieves are actually scientists or people who use the scientific method.

They are better described as conmen and thieves.

Two of them live in the USA and Wüster rarely ventures beyond foggy, wet Wales, where he spends most of his time being a keyboard warrior trolling others on platforms like Facebook, TikTok and Twitter.

In other words the trio are about as far removed from continental Australia and skinks in *Lerista* Bell, 1833 *sensu lato* as could be possible.

To get a good idea as to what this gang of thieves do in their spare time on the internet see Mackay (2024) or Hoser (2024b) as cited in the Hoser 2024c paper relevant to this species, herein cited as Hoser (2024c).

Wells (2012) did also sum up all that is known about each and every recognized species of putative *Lerista* in Australia as of year 2012, including diagnoses, distributions, synonymies and obvious species groups.

Based on the evidence before him at the time, Wells (2012) grouped like species and placed them in what he thought were appropriate genera.

He resurrected from synonymy old and available names as is the correct ICZN rules and in the majority of cases, where no names were available, he erected new genera, which is also in accordance with the legally binding rules of scientific zoological nomenclature.

As part of an audit of Australia's herpetofauna, the genus *Lerista sensu lato* came under scrutiny by myself in the period post-dating Wells (2012).

This included a review of the relevant literature, including all that preceded and post-dated Wells (2012) as well as inspection of living and dead specimens of the relevant taxa.

As part of this review, Hoser in 2023 published a review of the *Nodorha bougainvillii* (AKA *Lerista bougainvillii*) *sensu lato* species complex (Hoser 2023c), including formal descriptions of five new species and a new subspecies.

That is an essentially south-east Australian assemblage.

Other potentially unnamed species and genera within *Lerista sensu lato* had came to light previously and in the absence of any indications of anyone else seeking to inspect or name these taxa, they became the subject of a major study published in Hoser (2024c) as cited in full at the end of this paper.

There were other potential candidate species in the assemblage, but rather than unreasonably delay the final result indefinitely, Hoser (2024c) was published in which 8 new genera were erected as well as the formal identification and naming of 19 new species and 5 new subspecies.

This paper of 2026 formally names 40 further species and 14 subspecies in the greater "*Lerista*" assemblage as a further contribution to completing the inventory of Australia's native vertebrates.
It appears that there remain further unnamed species-level taxa in the assemblage, especially in the species rich state of Western Australia.

MATERIALS AND METHODS

Essentially as for Hoser (2024c) unless otherwise stated.

Preceding this paper and as a methodology, all the relevant published literature, including as cited in detail in Wells (2012), Hoser (2024c) and herein was reviewed to:
1/ Confirm that the previously named taxa in *Lerista sensu lato* were valid species.

2/ Confirm the correct genus-level assignment of each form, and by validating that they:
A/ Conformed to an obvious species group and that,
B/ Based on publicly available sequence data they had diverged less than 10 MYA from their other nearest common ancestor and,
3/ In light of the preceding things to flag any potentially unnamed forms at genus or species levels, including subgenera or subspecies.

This was backed up by way of inspection of specimens, alive, dead, in photos and preserved in museums.

Inspection of specimens from the relevant areas was done to confirm what was mooted by way of literature review and the unnamed taxa flagged in any available relatively recent molecular studies.

Publicly available sequences available at Genbank were downloaded and analysed using Mega11 (Tamura *et al*. 2021) and checked against previously published phylogenies using the same data.

Similar to the methodology employed by Colgan *et al.* (2009) and Sadlier e*t al*. (2019), MEGA version 11 (Tamura *et al.* 2021) was downloaded from the web and used to calculate Kimura 2-parameter (K2P) genetic distances with pairwise deletion of missing data and assuming a discrete approximation

to the gamma distribution for modelling rate variation between sites (a shape parameter set to 1.0). Standard errors of the distances were estimated by a bootstrap analysis with 500 replicates.

The upper limit of the divergence rate generally assumed for substitution in cytochrome b in reptiles

is 2.5% per million years (Crochet *et al.* 2004).

Assuming this rate as correct, as was done by Colgan *et al.* (2009), it was decided that where *Lerista* clades had apparently diverged beyond 1.5 MYA, the relevant lizard groups were assessed to see whether or not they were:
1/ Morphologically diagnosable as separate species and
2/ If a likely or logical biogeographic factor causing isolation of populations could be identified.

The latter may be:
A/ Habitat barrier/s (e.g. soil type, vegetation type, rock type, etc),
B/ Competing or predatory species in an area or
C/ Presumed past climatic events that may have facilitated either of the prior types of factor.

Where applicable, if a case for separation of populations was made out, due to morphological divergence, but date of divergence was not believed to be great, then I opted to make the relevant taxa subspecies.

Alternatively, where no sequence data was available, but other factors such as significant morphological divergence had occurred, especially if connected to a well-known or obvious biogeographical barrier of known antiquity, then the decision would be made to determine the relevant form as a separate species (or if appropriate as subspecies).

Most publications relevant to the taxonomy and nomenclature of the species within *Lerista sensu lato* and the final decisions made herein are listed in Hoser (2024c).

Wells (2012), included a vast number of *"Lerista"* related references not included in Hoser (2024c) or herein or relied upon in terms of the taxonomic or nomenclatural decisions herein.

Those relate to other aspects of the relevant species not required for taxonomic assessment or are in the genre of field surveys and reports of taxa in which "*Lerista*" species were mentioned as caught in a given place, but with little if any meaningful information beyond that.

Alternatively, a lot of the references cited by Wells (2012) not cited herein merely duplicated the material and or findings of the publications cited herein.

As much as possible, I have tried to rely on the original sources of information rather than secondary materials (refer also to the etymology for the species *Gaia questionthenarrative sp. nov*. as well).

To that end, I read every single formal description of every putative species within the *Lerista sensu lato* group and all are cited in Hoser (2024c) itself.

In the intervening period between Hoser (2024c) and the present publication, two more species have been formally described from northeast Australia in the Gulf of Carpentaria region, being

"*Lerista karichigara* Zozaya, Vanderduys, Macor, Read and Amey, 2025"

and

"*Lerista munuwajarlu* Zimny, Vanderduys, Kemp and Zozoya, 2025"

Both species are herein transferred to the genus *Gavisus* Wells and Wellington, 1984, based on the information in the original papers that were published in the non-peer reviewed online journal *Zootaxa*, associating them with *Gavisus wilkinsi* (Parker, 1926).

In terms of genus-level assignments, relevant published phylogenies were reviewed, including Skinner *et al*. (2008) and Pyron *et al.* (2013), the latter of which I note post-dated Wells (2012).

Publications relevant to the taxonomic conclusions made within this paper in terms of species level classifications, including the identification and use of biogeographical barriers for co-distributed, similarly affected species and the like included all the material cited in Hoser (2024c) and in addition including the following papers: Hoser (2024c-e, 2025a-e), Singhal *et al*. (2025), Torkkola *et al*. (2026), Zimny *et al*. (2025), Zozaya *et al*. (2025) and sources cited therein.

Hoser (2024d) and Hoser (2025e) both relate to putative species being split into large numbers across similar and sometimes the same biogeographical barriers and in the case of Hoser (2025e) a group of lizards that have speciated extensively in arid areas.

Websites and online references cited anywhere in this paper, were active, available and as cited as of 8 June 2026.

The relevant papers in total were: Ahl (1935), Amey and Couper (2009), Amey *et al.* (2005, 2019a, 2019b), Anonymous (1969), Aplin and Smith (2001), Bamford (1988), Bauer et al. (2003), Bell (1833a, 1833b), Blackburn (1999), Boehm (1943), Boulenger (1887, 1891, 1896), Bradshaw (1988),

Brandle (2010), Brandley *et al.* (2008), Brown (2014), Burbidge and McKenzie (1983), Ceriaco *et al*. (2023), Chapman and Dell (1985), Cogger (1979, 2014), Cogger *et al.* (1983), Colgan et al. (2009), Cope (1892), Cotton (2014), Crochet *et al.* (2004), Couper and Amey (2009, 2017), Couper and Ingram (1992), Couper *et al*. (2006, 2016), Covacevich (1971), Covacevich *et al.* (1997), Daudin (1802), De Vis (1888, 1889), Dubois *et al.* (2019), Duméril and Bibron (1839), Dunn and Dunn (1940), Ehmann (1992), Fairbairn *et al.* (1988), Farquhar *et al*. (2014), Fischer (1882), Fitzinger (1843), Ford (1963, 1965), Fry (1914), Fuhn (1969), Gaikhorst (2002), Glauert (1960, 1961, 1962), Gray (1830, 1839, 1841, 1845, 1864, 1865, 1867), Greer (1967, 1970, 1974, 1979a, 1979b, 1980, 1983, 1986, 1987, 1989, 1990a, 1990b, 1991), Greer *et al*. (1983, 1991), Grimm-Seyfarth *et al.* (2019), Günther (1867, 1876), Hawkeswood (2021), Horner (1991), Hoser (1989, 2007, 2014a, 2013b, 2016, 2018a, 2018b, 2018c, 2018d, 2019a, 2019c, 2019d, 2019e, 2019f, 2020a, 2020b, 2021, 2022a, 2022b, 2022c, 2023a, 2023b, 2023c, 2023d, 2023e, 2023f, 2024a, 2024b, 2024c, 2024d, 2024e, 2025a, 2025b, 2025c, 2025d, 2025e), Hutchinson (2008), Hutchinson and Donnellan (1993), ICZN (1991, 2001, 2012, 2021), James and Shine (2000), Jörger and Schrödl (2013), Kay and Keogh (2012), Kohlsdorf and Wagner (2006), Lande (1978), Longman (1937), Loveridge (1933, 1934), Lucas and Frost (1895, 1896), Marx and Hosmer (1959), Maryan (1993, 1996, 1998, 2006), Maryan and Reinhold (2010), Maryan and Robinson (1997), McCoy (1881), Mitchell (1955), Mittleman (1952), Mosyakin (2022), Müller (1880), Parker (1926a, 1926b), Pianka (1972), Prates *et al.* (2021, 2022a, 2022b, 2023), Pyron *et al.* (2013), Rabosky *et al.* (2007), Ride *et al.* (1999), Schluter and Hallermann (1997), Shea (1991a, 1991b, 1993, 2021), Shea and Sadlier (1999), Singhal *et al*. (2025), Skinner (2007a, 2007b, 2010), Skinner and Lee (2009), Skinner *et al.* (2008), Smith (1849), Smith (1996), Smith and Adams (2007), Smith (1937), Sternfeld (1919), Stirling and Zietz (1893), Storr (1964a, 1964b, 1971, 1976, 1978, 1979, 1980, 1982, 1984a, 1984b, 1985a, 1985b, 1986a, 1986b, 1990a, 1990b, 1991a, 1991b, 1991c, 1991d, 1991e, 1991f), Storr and Harold (1978, 1984, 1985), Storr *et al*. (1981, 1983, 1999), Swan *et al.* (2022), Torkkola *et al*. (2026), Waite (1900), Wellington (2016), Wells (2012), Wells and Wellington (1984, 1985, 1988), Werner (1903), White (1976), Wiens (2009), Wiens and Slingluff (2001), Wiens *et al.* (2006), Wilson (2022), Wilson and Knowles (1988), Wilson and Swan (2010, 2021), Zietz (1920) Zimny *et al*. (2025), Zozaya *et al*. (2025) and sources cited therein.

RESULTS

Following both literature review, including reviewing the available published molecular data as outlined above and/or in Hoser (2024c), and direct inspection of relevant specimens, it was deemed appropriate to recognize in this paper a total of 40 new species and 14 new subspecies.

All are named in accordance with the rules of the *International Code of Zoological Nomenclature* (Ride *et al*. 1999) as amended (ICZN 2012), as part of the permanent published scientific record.

This is in line with all other papers published by myself in *Australasian Journal of Herpetology* since and including year 2009.

Most new taxa named in this paper are from Western Australia which for many years has been recognised as an important centre for speciation within *Lerista sensu lato*.

A SUMMARY OF THE RESULTS, INCLUDING TAXONOMIC AND NOMENCLATURAL ACTS IN FURTHER DETAIL

AAH EDWARDSAE (STORR, 1982) FURTHER DIVIDED.

Hoser (2024c) formally named *Aaah edwardsae skink* Hoser, 2024 which had previously been treated as the eastern population of *Aaah edwardsae* (Storr, 1982), better known as "*Lerista edwardsae* Storr, 1982" with a type locality of Streaky Bay. South Australia, Australia (Latitude -32.50 S., Longitude 134.15 E.), being on the western edge of the Eyre Peninsula.

Aaah edwardsae skink Hoser, 2024 is a taxon of the Spencer Gulf and the lower Eyre Peninsula in South Australia.

Nominate *Aaah edwardsae* (Storr, 1982) is a taxon found roughly from Streaky Bay in the east to near Penong in the West in South Australia.

Two other subspecies are formally named for the first time within this paper.

These are *Aaah edwardsae gotit subsp. nov.* from the arid region west of Lake Torrens, South Australia and *Aaah edwardsae hellyeah subsp. nov.* from a 20 km radius around Yalata, South Australia.

PUTATIVE ACDC FLAMMICAUDA (STORR, 1985) IS SPLIT THREE WAYS

Until now, both *Acdc tnt sp. nov*. and *Acdc thunderstruck sp. nov*. were treated as outlier populations of the Pilbara (Australia) taxon *Acdc flammicauda* (Storr, 1985) (type locality Marandoo, Western Australia, Australia, Latitude 22.37 S., Longitude 118.08E.), which was originally placed by Storr and most subsequent authors in the genus *Lerista* Bell, 1833.

The three species (as defined herein) are distributionally disjunct, being separated by major known biogeographical barriers, have clearly been

separated for millions of years (certainly in excess of 1.5 MYA based on the age of the barriers in their present forms and available molecular data that is publicly available) and are morphologically divergent.

Therefore, I have no hesitation in formally describing and naming the outlier populations as new species.

Acdc tnt sp. nov. occupies the north-west Pilbara District of Western Australia, generally north of the Fortescue River.

Acdc flammicauda (Storr, 1985) occupies the main Pilbara District south of the Fortescue River and north of the Ashburton River.

Acdc thunderstruck sp. nov. appears to be restricted to the Barlee Range area, south of the Ashburton River.

GENUS *AH* HOSER, 2024 IS REDEFINED AND THREE NEW SPECIES FORMALLY NAMED

The genus *Ah* Hoser, 2024 now has the following five species: *Ah ha* Hoser, 2024 (type species); *Ah christinae* (Storr, 1979), *Ah yess sp. nov.* (this paper); *Ah pfuk sp. nov.* (this paper) and *Ah sheat sp. nov.* (this paper).

The three newly named species are most closely related to the species *Ah christinae* (Storr, 1979) and *Ah ha* Hoser, 2024.

They are:

Ah yess sp. nov. is found along the west coast and nearby hinterland of Western Australia from just north of Kalbarri, Western Australia south to about Jurian Bay (encompassing an area known as the lower Coral Coast).

Ah pfuk sp. nov. is found in the region from Nerren Nerren in the south, along the coast and nearby hinterland north to Exmouth, Onslow, Barrow Island and the Montebello Islands.

Ah sheat sp. nov. is apparently restricted to the Kennedy Range area of western Western Australia

It should also be noted that the morphologically convergent species *Miculia elegans* Gray, 1845 is in effect confined to the region south of Jurian Bay, along the West Australian coastline and nearby hinterland south through Perth to Augusta.

***APHRODITIA GERRARDII* (GRAY, 1864) IS SPLIT INTO THREE SUBSPECIES**

Until now, no subspecies have been recognised in the putative species *Aphroditia gerrardii* (Gray, 1864), with a type locality of Swan River, north-east of Perth, Western Australia, Australia.

The nominate form occupies most of the range, with outlier populations in the north-west (on the coast) and north-east herein formally recognised and named as new morphologically divergent subspecies, being *A. gerrardii nhanta subsp. nov.* and *A. gerrardii badimaya subsp. nov.* respectively.

A WESTERN POPULATION OF PUTATIVE *APHRODITIA DESERTORUM* (STERNFELD, 1919) IS FORMALLY NAMED AS A NEW SPECIES

Aphroditia harriganorum sp. nov. has until now been treated as a western population (from west-central western Australia) of the relatively widespread species *A. desertorum* (Sternfeld, 1919), type locality of Hermannsburg Mission, upper Finke River, N. T., designated by Mertens (1967).

A. desertorum occurs in most of the southern half of the Northern Territory, most of South Australia, except for the far south and south-east and into inland southern Western Australia extending west to north of the Goldfields (Kalgoorlie).

The species *Aphroditia harriganorum sp. nov*. appears to be restricted to a relatively small area bound by Leinster Downs Station, Latitude -27.975556 S., Longitude 120.299444 E., Carnarvon Range, Latitude -25.317778 S., Longitude 120.870556 E. in the north, broadly extending about 150 km to the west from these two points.

PUTATIVE *GAIA IPS* (STORR, 1980) IS SPLIT THREE WAYS

Until now, each of *Gaia dezifreemani sp. nov.* and *Gaia aviyemenii sp. nov*. have been treated as outlier populations of *Gaia ips* (Storr, 1980) originally placed in the greater genus *Lerista* (Bell, 1833), with a type species of *Lerista lineata* Bell, 1833 by monotypy.

The type locality of *Gaia ips* is Lake Auld, Western Australia, Australia, Latitude 22°07' S., Longitude 123° 52 E., which is within the Great Sandy Desert bioregion, being where that species as recognised herein occurs.

Where *Gaia dezifreemani sp. nov.* occurs to the east in far east Western Australia and the adjacent southwest parts of the Northern Territory is in the western part of the Tanami region and separated by a zone of unsuitable habitat dividing the two taxa.

Gaia aviyemenii sp. nov. is a species of the Little Sandy Desert, separated by hills extending from the Pilbara from the Great Sandy Desert to the north and northeast of the Pilbara.

The three populations of each species are physically isolated from one another, genetically divergent from one another, as shown in Singhal *et al.* (2025) (see also their supplementary data) and so morphologically divergent from one another that it is entirely appropriate that they are recognised as three full species rather than as one highly variable one.

The three species are most readily separated from one another by dorsal colouration in adults.

TWO MORE SPECIES PREVIOUSLY ASSOCIATED WITH *GAIA GREERI* (STORR, 1982) ARE FORMALLY NAMED

Hoser (2024c) split the species *Gaia greeri* (Storr, 1982) as recognized then (being that of Storr, 1982)

into two species based on very significant and obvious divergences, distributional disjuncture and known antiquated biogeographical boundaries.

Singhal *et al.* (2025) in their supplementary Fig S5 showed that Storr's concept of *G. greeri* included four putative species implying that Hoser (2024c) had not gone far enough.

This view was clarified somewhat by Torkkola *et al.* (2026) at Fig 1., where they showed that the Western type form was in fact split into two closely related species (one north form from Storr Island, West Kimberley and the south Type form of *G. greeri*, from 8 km south southeast of Derby, Western Australia, Latitude 17.22 S., Longitude 123.40 E.).

Additionally, Torkkola *et al.* (2026) found that an eastern sample, being West Australian Museum specimen number WAM R137387 collected from Fowlhouse Junction, Osmond Range, East Kimberley district of Western Australia, Latitude -17.266667 S., Longitude 128.483333 E. was a very divergent taxon.

This was essentially in line with the concept of Hoser (2024c) for *G. oomph* Hoser, 2024, supporting the erection of that species in the strongest possible terms.

It should be noted that the type form of *G. oomph* from the Tanami Desert, while similar in most respects to other East Kimberley animals previously referred to *G. greeri,* did differ from some.

Were it not for my inherently conservative nature, I would have recognised at least one further species or subspecies on the basis of the differences. However, me being ever (over) sensitive to ongoing unsupported claims by detractors that I have over split reptile species, I refrained from doing so.

In hindsight, taking notice of unscientific keyboard warriors, working as flying monkeys for the Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang of thieves, generally from fake accounts, was a serious mistake on my part.

To confirm my mistake in listening to trolls, Singhal *et al.* (2025) in their supplementary Fig S5 also indicated that there were in fact two species occupying the same general region, this being the inland south and east Kimberley districts.

In other words, only two of the relevant four species had been formally named until now, although this did include the two main lineages.

Rather than having the less divergent, but still unnamed species-divergent lineages expire through general indifference I formally name them both herein. This is so that proper conservation measures can be implemented by governments, non-governmental organizations and concerned individuals.

The south and southeast Kimberley form is herein named *Gaia rate sp. nov.*, while the Storr Island form is herein named *Gaia otit sp. nov.*.

This means that the diagnoses and concepts for *Gaia greeri* and *Gaia oomph* as given in Hoser (2024c) must be amended slightly herein to accommodate the two other species formally named within this species complex.

PUTATIVE *GAIA VERMICULARIS* (STORR, 1982) IS SPLIT FOUR WAYS

As originally described and conceived "*Lerista vermicularis* Storr, 1982", herein transferred to the genus *Gaia* Wells, 2012 in line with Hoser (2024c) was a species only known from the Great Sandy Desert, in north-west Western Australia (see Storr, 1982 including "Figure 3").

More recently specimens that are morphologically similar from the northern extension of the Great Sandy Desert, Tanami Desert Region and the Gibson Desert Region have been collected, lodged in the Western Australian Museum and assigned to the same species.

The same applies to colour images of these lizards posted on photo sharing sites such as "Inaturalist" and "Flickr".

However, the lizards themselves (as seen in the images posted) are quite different in form to one another. They are separated by barriers of unsuitable non-dune habitat of significant antiquity, and the molecular results of Torkkola *et al.* (2026) have confirmed the existence of three very different species divergent forms.

The northern-most form was not included in the study of Torkkola *et al.* (2026) but is morphologically divergent from the others and also separated by areas of unsuitable (non-dune) habitat, implying lengthy isolation.

Therefore, the three hitherto unnamed forms are formally named herein for the first time.

Gaia nomenclaturematters sp. nov. is the form previously assigned to *G. vermicularis* from the Tanami Desert and North Gibson Desert in east Western Australia.

Gaia taxonomymatters sp. nov. is the form previously assigned to *G. vermicularis* from the middle Gibson Desert in east Western Australia.

Gaia ruleofpriority sp. nov. occurs in a confined area east of Baines Hills and Elsey Hills in the area between the Great Sandy Desert (to the south-west), southeast Kimberley District (to the north), the open grass plains to the east and the Gardiner Range to the southeast

PUTATIVE *GAIA GRIFFINI* (STORR, 1983) IS REDEFINED AS FOUR FULL SPECIES

Predating the publication of Hoser (2024c), *Gaia griffini* (Storr, 1983), with a type locality of Kununurra, Western Australia, Australia, Latitude -15.47 S., Longitude 128.44 E., had been treated as a species

occurring across the Kimberley District of Western Australia and nearby areas, across a wide zone in the lower top end of the Northern Territory.

This distribution is also disjunct.

Relying primarily on morphological evidence, Hoser (2024c), split off the eastern population as *Gaia griffini asgicondi* Hoser, 2024.

Gaia griffini asgicondi Hoser, 2024 has a centre of distribution on the Sturt Plateau bioregion of the north central Northern Territory including nearby hilly areas, such as the uplands near the south-west Gulf of Carpentaria.

There is a distributional gap of over 100 km straight line between these populations.

Populations of putative *Gaia griffini* from all parts of the Kimberley District were erroneously treated by Hoser (2024c) as being a single taxon, when it was stated "*The Kimberley population (both east and west Kimberley) are of the type subspecies*."

This error was shown in the paper of Singhal *et al.* (2025) in which they found three molecular divergent forms inhabiting the Kimberley District.

This included the type form from Kununurra and environs in the east Kimberley, a form from the west Kimberley and a form from the Dampier Peninsula and south along the nearby coast in Western Australia.

In a second error, Hoser (2024c) recognised the species originally described as "*Lerista praefrontalis* Greer, 1986" and mistakenly followed Wells (2012) by placing it in the genus *Rhodona* Gray, 1839, when it should have correctly been placed in the genus *Gaia* Wells, 2012.

"*Lerista praefrontalis* Greer, 1986" corresponds to the west Kimberley population and so there is no need to engage in Wüster / Britton / Kaiser style taxonomic vandalism by giving it a junior synonym name.

On the basis of distinct gaps in the distributions of all four forms of putative *Gaia griffini* (Storr, 1983), morphological and molecular divergences, all are herein treated as full species.

In other words they are referred to as *Gaia griffini* (Storr, 1983), *Gaia asgicondi* (Hoser, 2024), *Gaia praefrontalis* (Greer, 1986) and *Gaia nottobemissed sp. nov.*.

Gaia nottobemissed sp. nov. is herein formally described.

The diagnosis for *Gaia asgicondi* Hoser, 2024 is modified slightly from that publication (being Hoser 2024c) to accommodate the three taxa formerly all treated as Kimberley *G. griffini.*

THE *GAIA BIPES* (FISCHER, 1882) SPECIES GROUP IS SPLIT FROM 3 TO 13 SPECIES, WITH TEN FORMALLY NAMED FOR THE FIRST TIME

Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

The thirteen species in the *G. bipes* complex as identified herein are as follows:

G. bipes is a taxon of the Pilbara region, found only north of the Fortescue River in Western Australia in the region of Nickol Bay, the Burrup Peninsula and near the coast.

G. kunja is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin.

G. arrernte is herein restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

G. amazing sp. nov. is a coastal taxon restricted to a small area from around Broome to about 100 km north of Broome, in Western Australia.

G. wow sp. nov. is a taxon found from Port Hedland to around Sandfire Flat/Mandora in northwest Western Australia.

G. robwatsoni sp. nov. is known only from around the Derby area in northwestern Western Australia.

G. vellai sp. nov. is a species of the west coastal south Pilbara, south of the Fortescue River and including nearby dune country to the south.

G. questionthenarrative sp. nov. is a species of the eastern Great Sandy Desert in Western Australia.

G. narcissismawareness sp. nov. is a species with a centre of distribution being the Little Sandy Desert and nearby parts of central Western Australia.

G. truthisscience sp. nov. is a species from the Goldfields area and nearby parts of south-east Western Australia, well north of the coast.

G. liesarenotscience sp. nov. is confined to the central west of South Australia, north and east of the Nullarbor (but not near the coast) and south of the floodplain areas in north-west South Australia. The Lake Eyre system forms a hard barrier to the east.

G. anangu sp. nov. is restricted to central Australia south of the Amadeus basin and north of the flooding areas in far north South Australia.

G. owe sp. nov. is confined to the Simpson Desert in the south-east of the Northern Territory.

G. anangu sp. nov. and *G. owe sp. nov.* are the only two relevant species named in the absence of molecular data confirming or denying their status.

However, both occur in well-defined areas with established barriers on all sides and appear to be among the most divergent of the relevant forms.

_GO_ HOSER, 2024 GETS A NEW SPECIES … _GO FORIT SP. NOV._

For most of the past 160 years, the species originally described as "*Rhodona punctato-vittata* Günther, 1867" with a type locality of "Queensland" has been treated as a single species occupying the major part of inland eastern Australia.

This remained the case until relatively recently when Hoser (2024c) created the new genus *Go* Hoser, 2024 to accommodate this divergent lineage.

At the relevant time the three recognised species were:

Go punctatovittata (Günther, 1867) of the Murray Darling System, *Go emmotti* (Ingram, Couper and Donnellan, 1993) of the Cooper's Creek drainage and *Go colliveri* (Couper and Ingram, 1992) of the the Burdekin drainage in north-east Queensland), the genus being mainly west of the Great Dividing Range in Eastern Australia (except for the eastern distribution of *G. colliveri*).

Singhal *et al*. (2025) found three species within putative *Go punctatovittata.* These were the type form from inland southern Queensland (and also most of western New South Wales, except for the far south-west), another from north-west Victoria and nearby South Australia as well as far southwest New South Wales and a third lineage from the northern Flinders Ranges, in the Lake Torrens drainage basin.

The north-west Victorian population has an available name in the form of "*Rhodona officeri* McCoy, 1881" with a type locality of Swan Hill in Victoria.

Therefore, there is no need for me to engage in Wolfgang Wüster / Hinrich Kaiser / Adam Britton style taxonomic vandalism in renaming this taxon and pretending that McCoy's work never existed (see Hoser 2024c for details and relevant citations).

Cogger *et al.* (1983) synonymised "*Rhodona officeri* McCoy, 1881" with their *Lerista punctatovittata* (Günther, 1867) and in this paper I formally reverse this by resurrecting it from synonymy.

The northern Flinders Ranges population in South Australia is genetically and morphology divergent and so I have no hesitation in naming it as a new species herein, being *Go forit sp. nov*..

PUTATIVE _MICULIA DISTINGUENDUS_ (WERNER, 1910) IS FURTHER SPLIT WITH TWO NEW SPECIES BEING FORMALLY NAMED

Until 2024, the putative species *Ablepharus distinguendus* Werner, 1910, with a type locality of Freemantle (Perth), Western Australia, mostly placed in the genus *Lerista* Bell, 1833 was treated as a single species found from the Eyre Peninsula, South Australia west across southern Australia to the west coast and north to just past Geraldton in Western Australia.

In line with Wells and Wellington (1984, 1985) and Wells (2012), Hoser (2024c) placed the relevant species in the genus *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845.

The phylogeny of Torkkola *et al.* (2026) again confirmed a genus-level divergence of more than 10 MYA for the relevant group of species.

Hoser (2024c) also described the population from the northern two thirds of the Eyre Peninsula of South Australia as a separate and divergent species, namely *M. ruficauda* Hoser, 2024.

The published phylogeny of Torkkola *et al.* (2026) implied two other undescribed species in this complex, the first being the putative localised outlier population from the Geraldton district of the central west coast of Western Australia.

Because it is morphologically divergent, it is herein formally named as *Micula chrismartini sp. nov.*.

Another geographically separated and morphologically divergent population is found from about the Stokes National Park, which is roughly 70 km straight line west of Esperance, Western Australia, eastwards to about Madura on the western edge of the Nullarbor Plain proper, also in Western Australia, with outlier populations stretching inland and west to at least as far west as Hyden/Kondinin Cells (roughly 280 km in a straight line east of Perth).

That species is herein formally named as *Micula coldplayorum sp. nov.*.

MICULIA AERICEPS (STORR, 1986) IS FORMALLY SPLIT

The type form of *Miculia aericeps* (Storr, 1986), type locality of 25 km south of Alice Springs, Northern Territory, Australia is a taxon found from northwest New South Wales, across northern South Australia, the Northern Territory and across a large swathe of Western Australia to the eastern edge of the Pilbara District. It is a distinctive lizard with an orange or orangish brown head, yellowish orange body, mainly yellow tail and immaculately white upper labials, that are sometimes barred or partially barred along the suture lines from the top.

Putative *M. aericeps* from the Barkly Tableland in the Northern Territory are so morphologically divergent that they appear to be of a different species. Were it not for the fact that I have seen specimens of apparently intermediate form (but more like *M. aericeps* in appearance) from Barrow Creek in the Northern Territory, I wouldn't have hesitated in naming the Barkly Tableland specimens as a separate species.

Instead, they are conservatively named herein as the subspecies *Micula aericeps howzat subsp. nov.*.

PUTATIVE _MICULIA TAENIATA_ (STORR, 1986) IS SPLIT TWO WAYS.

Until now, *Miculia puseyi sp. nov*. has been treated as a southern population of putative *Miculia taeniata* (Storr, 1986), a species that was originally placed by Storr in the genus *Lerista* Bell, 1833 and with a type locality of 20 km east of The Granites, Northern Territory, Australia, Latitude -20.34 S., Longitude 130.38 E.

Singhal *et al.* (2025) showed a molecular basis to separate the two morphologically divergent forms and so the unnamed one has been named herein.

Miculia taeniata (Storr, 1986) occupies much of central and southern Australia north of the Great Australian Bight and well away from the coast. At the southern extremity of this range in southern Australia, *Miculia puseyi sp. nov*. occurs.

MICULIA VACOPOULOSAE SP. NOV. A NEW SPECIES FROM INLAND SOUTHERN AUSTRALIA.

Until now *Miculia vacopoulosae sp. nov.* has been variously treated as a southeastern population of either *Miculia xanthura* (Storr, 1976), type locality of 7 km southwest of the Pollock Hills, eastern Western Australia, Australia, Latitude -22.54 S., Longitude 127.30 E. or *Miculia aericeps* (Storr, 1986) with a type locality of 25 km south of Alice Springs, Northern Territory, Australia.

Specimens of *M. vacopoulosae sp. nov.* lodged in Australian museums and posted online have been lodged and posted as both other species in an obvious case of misidentification of a hitherto unnamed species.

PUTATIVE _PHANEROPIS CLARA_ (SMITH AND ADAMS, 2007) IS SPLIT TWO WAYS.

Since 2007, the putative species "*Lerista clara* Smith and Adams, 2007", herein placed in the genus *Phaneropis* Fischer, 1881 (type species *Phaneropis muelleri* Fischer, 1881) has been treated as a taxon occupying a distribution encompassing the coastal west Pilbara in a zone from just north of Carnarvon in the south to just north of Sandfire Flat in the north. However recent phylogenies published by Singhal *et al*. (2025) and Torkkola *et al.* (2026) showed two very divergent species level lineages within this population.

As they are morphologically divergent as well, the unnamed northern population is herein formally named as *P. abscondeetus sp. nov.*.

PUTATIVE _PHANEROPIS TIMIDA_ (DE VIS, 1888) IS SPLIT 11 WAYS WITH 8 SPECIES NAMED FOR THE FIRST TIME

Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Two other taxa, "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia have been generally treated as synonyms of "*Lerista timida*" by publishing herpetologists in recent years (postdating Cogger *et al.* 1983 or Cogger 2014).

Contrary to this position, Hoser (2024c), placed the relevant species into the genus *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 and recognised the east Australian species with the priority name *Phaneropis rhodonoides* (Lucas and Frost, 1896).

Molecular results of both Singhal *et al*. (2025) and Torkkola *et al.* (2026) have shown the Hoser (2024c) taxonomy and nomenclature to be correct.

However, their results confirmed that the previously synonymised ""*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935"" was divergent at the species level from all other named species-level taxa.

In addition to that, the phylogeny of Torkkola *et al.* (2026), also showed three other unnamed species within their putative "*Lerista timida*" including populations from south of Lake Eyre in South

Australia, Northwest of Kalgoorlie, Western Australia, both as part of a larger mainly east Australian clade as well as another population from south-east of the Pilbara district in Western Australia.

A further sweep by myself of the various known populations of putative "*Lerista timida*" yielded five further unnamed populations.

Each are isolated by biogeographical barriers, usually in the form of extensive areas of periodically flooding black soil plains, or alternatively extensive mobile or stationary sand dunes.

The isolation of these populations is reflected in morphological divergence.

To make it clear, while species in this complex occupy a range of habitat types, they appear to be excluded from large areas of sand dunes and also periodically flooded black soil areas. This means for example that none occur in the lower Cooper's Creek drainage system (of eastern inland Australia) or the Simpson Desert proper.

Therefore, I have no hesitation in naming all 8 relevant populations as new species and caution also that there may yet be further unnamed species within this complex.

These 8 newly named species are as follows:

Phaneropis dorsei sp. nov. is the taxon of south-west Western Australia, generally west of the Nullarbor.

P. faaarkinelle sp. nov. is confined to a region north of the Nullarbor in South Australia in the south, bound by red dune country to the north, still in South Australia, extending south-east to the eastern edge of the Nullarbor.

P. wholeeecow sp. nov. is a species apparently confined to the Tomkinson Ranges and adjacent elevated and hilly areas in north-west South Australia and immediately adjacent parts of the Northern Territory and Western Australia. This area is in turn surrounded by mainly red dunes or flooding black or red soil areas.

P. overlookedit sp. nov. is confined to the Selwyn Ranges and Mount Isa area in northwest Queensland. This region is surrounded by black soil plains on three sides and the Gulf of Carpentaria on the north.

P. lopezi sp. nov. is a distinctive dark coloured form found in the area of the Northern Territory and South Australian border, south of the Amadeus basin and its south-eastern extension, and north of the low-lying plains between Coober Pedy in South Australia and the Northern Territory border. To the east and west, this species is constrained by significant areas of hard packed flooding plains.

P. greatfind sp. nov. is a relict form confined to the elevated parts of the Sturt National Park and environs in far northwest New South Wales, separated on all sides by flooding black or cracking soil lowlands.

P. laylawangae sp. nov. is a taxon found in the region occupying most of central South Australia, excluding the far south-east, far west, and far north.

P. gottabekidding sp. nov. is found in north-central Western Australia, east of the Pilbara district and west of the main Gibson Desert. It is closely related to the Pilbara species within the genus *Phaneropis*, rather than the east Australian lineage, which includes the other species named herein.

Of relevance also is that *P. timida* of the type form is herein confined to most of New South Wales, west of the Great Dividing Range, excluding the far south-west and far northwest, as well as being found in most of south Queensland west of the Great Divide, bound in the west by the lowland flooding black soil areas of the Lake Eyre drainage system.

P. rhodonoides is found in the region of north-west Victoria, adjacent south-west New South Wales and adjacent parts of South Australia to the east of the Flinders Ranges.

P. goerlingi is a distinctive lighter coloured form found in central west Western Australia, generally near the coast, south of the Pilbara district.

PUTATIVE *PHANEROPIS MUELLERI* (FISCHER, 1881) IS SPLIT INTO TWO

The concept of *Phaneropis muelleri* Fischer, 1881 (type species for that genus named at the same time) has changed significantly in recent years.

From 1983 to 2026, almost all authors have followed on from Cogger *et al*. (1983) by placing the relevant species within the better-known genus *Lerista* Bell, 1833.

Wells and Wellington, 1985, shifted the relevant species to *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, but Wells (2012) shifted the relevant species to *Phaneropis* Fischer, 1881 based in particular on a recent molecular study published by Skinner *et al.* (2008).

Hoser (2024c) and this paper follow on from that position.

P. muelleri, was originally viewed as a pan-Australian species found across most of the continent, except for the extremities, especially in the north, south and far east.

Various studies and papers have basically cut the putative species up so that in most recent years, since the publication of Smith and Adams (2007) *P. muelleri* has been essentially confined to the Pilbara district of Western Australia.

However, the phylogeny of Torkkola *et al.* (2026) showed two very divergent species level lineages within this population. These populations were divided across the Fortescue River, a well-known biogeographical barrier of known antiquity. Furthermore, they are consistently morphologically

divergent and so I have no hesitation in naming the unnamed southern form as a new species herein.

Phaneropis muelleri Fischer, 1881 has a type locality of Nichol Bay, Western Australia, Australia, (lectotype designation by designated by Smith and Adams (2007)), being the part of the Pilbara District that is north of the Fortescue River.

P. oxy sp. nov. is the species from the main Pilbara District, south of the Fortescue River.

RHODONA CONNIVENS (STORR, 1971) IS SPLIT THREE WAYS

Until now, no subspecies of *Rhodona connivens* (Storr, 1971) have been recognised.

Singhal *et al*. (2025) in their supplementary data recognised three so-called "Operational species" within putative "*Lerista connivens*" of west Western Australia, Australia.

These were the nominate subspecies from the immediate vicinity of Shark Bay, including islands and the type locality of Salutation Island, Western Australia, Latitude 26.32 S., Longitude 113.46 E. (Holotype WAM R25777), a form from the coast further north in the area north of Carnarvon and generally south of the Cape Range, and a population to the south found in a zone extending from Billabong Roadhouse to Kalbarri in Western Australia.

The subspecies *R. connivens nanda subsp. nov.* is found in a zone from Billabong Roadhouse to Kalbarri in Western Australia. *R. connivens yinggarda subsp. nov*. is the taxon found from Carnarvon in the south to Cape Range in the north, of Western Australia. Around and in Shark Bay, Western Australia one finds nominate *R. connivens*.

A SPECIES ASSOCIATED WITH _TELCHINOSCINCUS NICHOLLSI_ (LOVERIDGE, 1933) IS FORMALLY NAMED AS _T. CALISTACLEMENTSAE SP. NOV._

Until now, *Telchinoscincus calistaclementsae sp. nov.* has been treated as a divergent northern population of the better known *Telchinoscincus nichollsi* (Loveridge, 1933) *sensu* Hoser (2024c) and previously placed by most earlier authors in the greater genus *Lerista* Bell, 1833, but notably excluding Wells and Wellington (1984) who first erected the genus *Telchinoscincus*.

Telchinoscincus calistaclementsae sp. nov. is a relatively restricted taxon found within 200 km of the Western Australian coastline in the arid area mainly to the south of the Wooramei River and north of the Murchison River. South of the Murchison River one finds the closely related taxon *T. nichollsi* (Loveridge, 1933) and in the region bounded by the the Wooramei River and Gascoyne River to the north one finds the closely related taxon *T. gascoynensis* (Storr, 1986). All other species in the genus *Telchinoscincus* are concentrated within the same general region of Western Australia.

TELCHINOSCINCUS ONSLOVIANA (STORR, 1984) GETS TWO NEW SUBSPECIES.

Telchinoscincus onsloviana ah subsp. nov. is a newly named taxon apparently restricted to the eastern side of the Exmouth Gulf in Western Australia.

T. onsloviana har subsp. nov. is known from 14 km north of Giralia, to 2 km south of Marrilla, extending about 50 km east in the adjacent sandy zone, all in west, Western Australia, Australia.

The nominate form of *Telchinoscincus onsloviana* (Storr, 1984), is a taxon found further north in the vicinity of Onslow, extending along the coast about 40 km to the south in Western Australia.

TELCHINOSCINCUS PETERSONI (STORR, 1976) GETS TWO NEW SUBSPECIES.

Until now, no subspecies of *Telchinoscincus petersoni* (Storr, 1976) (a west Western Australian taxon) have been recognised since Aplin *et al.* (2022) synonymised "*Lerista talpina* Storr, 1991", type locality Bullara, Western Australia, Australia (Holotype: WAM R74947) with "*L. petersoni* Storr, 1976", type locality Yinnietharra, Western Australia, Australia, (Holotype: WAM R46197).

Singhal *et al*. (2025) in their supplementary data recognised four so-called "Operational species" within putative "*Lerista petersoni*". On the basis of consistent morphological divergences and the fact that each relevant population is geographically isolated and separated from one another, with each being centred on a given range of hills, it is appropriate that each be given formal taxonomic recognition.

Nominate *Telchinoscincus petersoni* occupies most of the known range for the putative species, including the Mount Augustus area of Western Australia.

T. petersoni talpina appears to be a Cape Range endemic.

T. petersoni sheeta subsp. nov. is a taxon found in the Barlee Range.

T. petersoni faaark subsp. nov. is a taxon found in the Kennedy Range.

TISM KALUMBURU (STORR, 1976) GETS TWO NEW SUBSPECIES

Tism kalumburu bestoff subsp. nov. is known only from the Carson's Escarpment, Western Australia, Australia.

Tism kalumburu whatareya subsp. nov. is known only from the vicinity of Theda Station in the north Kimberley District of Western Australia.

Tism kalumburu of the nominate form is from the Kalumburu area and near coastal environs in Western Australia.

INFORMATION RELEVANT TO THE FORMAL DESCRIPTIONS THAT FOLLOW

There is no conflict of interest in terms of this paper or the conclusions arrived at herein.

Several people including anonymous peer reviewers who revised the manuscript prior to publication are also thanked as are relevant staff at museums who made specimens and records available in line with international obligations.

In terms of the following formal descriptions, spelling of names should not be altered in any way for any purpose unless expressly and exclusively called for by the rules governing Zoological Nomenclature as administered by the *International Commission of Zoological Nomenclature* (Ride *et al.* 1999 and ICZN 2012).

Material downloaded from the internet and cited anywhere in this paper was downloaded and checked most recently as of 8 June 2026, unless otherwise stated and were accurate in terms of the context cited herein as of that date.

Unless otherwise stated explicitly, colour descriptions apply to living adult male specimens of generally good health, with an original tail (not shed or regenerated) and not under any form of stress by means such as excessive cool, heat, dehydration or abnormal skin reaction to chemical or other input.

Females tend to be slightly larger and with slightly more elongate bodies as a rule within this assemblage.

Readers should be mindful of the fact that specimens pre-sloughing skin may have dulled or obscured markings, juveniles often may have brighter or different markings to adults and regenerated tails as a rule lack the pattern or markings of original ones.

While numerous texts and references were consulted prior to publication of this paper, the criteria used to separate the relevant species has already been spelt out and/or is done so within each formal description and does not rely on material within publications not explicitly cited herein.

Material within descriptions may be repeated from others in this paper in order to ensure compliance with the *International Code of Zoological Nomenclature* as amended (Ride *et al.* 1999 and ICZN 2012).

If more than one taxon described herein is considered to be of the same species, then the name to be used in preference by a first reviser is that in terms of page priority herein as seen in the abstract keywords.

Material, items or references cited herein merely as “online” without a URL, can be easily found by copying and pasting the name or detail into a search engine such as “Google” and it will appear either as the first result or in the first page of results on a computer desktop screen.

It should be noted that as of 2026, almost all species discussed within this paper would have until now been treated by most publishing authors in the previous 25 years as being within the genus *Lerista* Bell, 1833, with the notable exceptions of myself, Wells, Wellington or Wells and Wellington.

It has been published on “Facebook” that the Wüster / Britton / Kaiser gang will be renaming the relevant genera at some stage in one or more of their narcissistic acts of taxonomic vandalism (as detailed in Hoser 2019a-b). Those junior synonyms should obviously not be used as correct. Their only use is to clutter up synonyms lists.

Acceptance and use of the these names as published in this paper, being a reflection of the scientific reality, should be done as rapidly as possible, as failure to do so may have serious adverse consequences for their long-term conservation. See Hoser (2019a, 2019b) for further details.

The descriptions are published in alphabetical order in terms of genera names and then species and subspecies names within the same genera. To a limited extent, this has resulted in some cases of most closely related species not being formally described in closest proximity in this paper.

This is notably the case in the genus *Gaia* Wells, 2012, where species from several species groups were named for the first time in this paper.

However, each relevant description does refer to the closest and most relevant species explicitly.

AAAH EDWARDSAE GOTIT SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:6E0E2DDC-DC26-47DE-96DA-620A09993D1A

Holotype: A preserved male specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R49052 collected from 3.6 km south of Marsella Hill, Arcoona Station, South Australia, Australia, Latitude -31.2614 S., Longitude 136.9283 E.

This facility allows access to its holdings.

Paratype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R49050 collected from 3.6 km south of Marsella Hill, Arcoona Station, South Australia, Australia, Latitude -31.2614 S., Longitude 136.9283 E.

Diagnosis: Hoser (2024c) formally named *Aaah edwardsae skink* Hoser, 2024 which had previously been treated as the eastern population of *Aaah edwardsae* (Storr, 1982), better known as “*Lerista edwardsae*” with a type locality of Streaky Bay. South Australia, Australia (Latitude -32.50 S., Longitude 134.15 E.), being on the western edge of the Eyre Peninsula.

Aaah edwardsae skink Hoser, 2024 is a taxon of the

Spencer Gulf and the lower Eyre Peninsula in South Australia.

Nominate *Aaah edwardsae* (Storr, 1982) is a taxon found roughly from Streaky Bay in the east to near Penong in the West in South Australia.

Two other subspecies are formally named for the first time within this paper.

These are *Aaah edwardsae gotit subsp. nov.* from the arid region west of Lake Torrens, South Australia and *Aaah edwardsae hellyeah subsp. nov.* from a 20 km radius around Yalata, South Australia.

The four subspecies are separated from one another by the following combinations of characters:

Nominate *Aaah edwardsae* (Storr, 1982) has:

1/ A beige coloured dorsum (versus light brown in *Aaah edwardsae skink* Hoser, 2024), 2/ Very light, broken or no dark etching between the scales on the upper surface of the head as also seen in *A. edwardsae gotit subsp. nov.* (versus heavy in *A. edwardsae skink* Hoser, 2024 and *A. edwardsae hellyeah subsp. nov.*), 3/ A dark lateral band that is medium thickness (versus thick *A. edwardsae skink* Hoser, 2024 or thin in *A. edwardsae hellyeah subsp. nov.*) 4/ Yellow upper labials, that are incompletely barred or etched on the lower parts (of some), but etched black at the tops and on adjoining scales, and 5/ Most specimens of this subspecies and *A. edwardsae hellyeah subsp. nov.* have two supraoculars in contact with the frontal, versus the reverse (one in contact) in *A. edwardsae skink* Hoser, 2024 and *A. edwardsae gotit subsp. nov.*

Aaah edwardsae skink Hoser, 2024 has:

1/ A generally darker coloured dorsum than the other three subspecies (being light brown rather than beige), 2/ Thick dark etching between the scales on the upper surface of the head (versus light or none in nominate *A. edwardsae* and *Aaah edwardsae gotit subsp. nov.*), 3/ A dark lateral band that is very thick (versus medium in *A. edwardsae* and *Aaah edwardsae gotit subsp. nov.* or relatively thin in *A. edwardsae hellyeah subsp. nov.*), 4/ Yellow upper labials, that are heavily barred black on the bottom (etchings) becoming black entirely around the upper edge and on adjoining scales, and 5/ Most specimens of this subspecies and *A. edwardsae gotit subsp. nov.* have only one supraocular in contact with frontal, versus the reverse in nominate *A. edwardsae* and *A. edwardsae hellyeah subsp. nov.* (2 in contact).

Aaah edwardsae gotit subsp. nov. has:

1/ A beige coloured dorsum (versus light brown in *Aaah edwardsae skink* Hoser, 2024), 2/ Light, broken or no dark etching between the scales on the upper surface of the head, like seen in nominate *A. edwardsae* (versus heavy in *Aaah edwardsae skink* Hoser, 2024 and *A. edwardsae hellyeah subsp. nov.*), 4/ White or whitish yellow upper labials, without dark etching on the lower edges, versus obviously yellow labials in the other three subspecies, and in this subspecies still with black entirely around the upper edge and on adjoining scales, and 5/ Most specimens of this subspecies and *A. edwardsae skink* Hoser, 2024 have only one supraocular in contact with the frontal, versus the reverse in nominate *A. edwardsae* and *A. edwardsae hellyeah subsp. nov.* (2 in contact).

Aaah edwardsae hellyeah subsp. nov. has:

1/ A beige coloured dorsum (versus light brown in *Aaah edwardsae skink* Hoser, 2024), 2/ Heavy dark etching between the scales on the upper surface of the head, like seen in *Aaah edwardsae skink* Hoser, 2024 (versus light, broken or not present in nominate *A. edwardsae* and *A. edwardsae gotit subsp. nov.*), 4/ Yellow upper labials, with dark etching in the form of triangles, the point on the lower edges, and in this subspecies still with black entirely around the upper edge and on adjoining scales, and 5/ Most specimens of this subspecies and nominate *A. edwardsae* have two supraoculars in contact with the frontal, versus the reverse (one in contact) in *A. edwardsae skink* Hoser, 2024 and *A. edwardsae gotit subsp. nov.*

A. edwardsae of the nominate subspecies is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/54817712249/

and

https://www.flickr.com/photos/127392361@N04/54817719148/

A. edwardsae skink Hoser, 2024 is depicted in life in Cogger (2014) on page 603 at top left and online at:

https://www.flickr.com/photos/reptileshots/55168787713/

and

https://www.flickr.com/photos/ryanfrancis/16587119909/

and

https://www.inaturalist.org/observations/325999588

and

https://www.inaturalist.org/observations/206751709

A. edwardsae gotit subsp. nov. is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/49231437681/

specimen from Pernatty Lagoon, South Australia, Australia, and.

https://www.inaturalist.org/observations/269583256

specimen from Unincorp, South Australia, Australia.

A. edwardsae hellyeah subsp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/154750972

specimen from the Eyre Highway, Yalata, South

Australia, Australia.

A. edwardsae and the other preceding named subspecies are separated from the closely related *A. picturata* (Fry, 1914) by having a much shorter foreleg, fewer dorsal lines, as in just 2 instead of four and usually less mid body rows (18-20 versus 20-22).

The preceding taxa are in turn separated from *A. baynesi* (Storr, 1972) (of both subspecies as identified in Hoser 2024c, being nominate *A. baynesi* (Storr, 1972) and *A. baynesi ngandatha* Hoser, 2024) by having a strong rather than faded dorsal colouration and 9-14 rather than 8-10 subdigital lamellae under the second toe.

The preceding species, being the entirety of the genus *Aaah gen. nov*. are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) and assuming placement of all relevant species within these genera, (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) by the following unique combination of characters:

All three species (and four additional subspecies) in the genus are thick set for burrowing skinks. They have one or other of 2 front digits (tiny first digit), or just one or sometimes none with a style, as well as always 2 digits at the rear; movable eyelid, nasals forming a median suture, frontoparietals paired, and an enlarged second supraocular excluding or nearly excluding first from contact with the frontal. Adult snout-vent length 64-95 mm. Length of appendages as a percent of snout vent length (SVL) is: fore-leg 0.5-1.7; hind-leg 11.1-18.9; snout to foreleg 21.9-25.7.

Nasals in short to long contact. Prefrontals widely separated. Frontoparietals separated, much smaller than the interparietal. 2-5 nuchals, 3 supraoculars, second always and first sometimes in contact with the frontal. Supraciliaries usually 0+1 or rarely 0+2 or 0+3. 6 supralabials. 2-3 temporals, usually 3. Upper secondary temporal largest, lower secondary temporal much the smallest.

18-22 Midbody scale-rows (usually 20). 8-15 lamellae under the longer toe.

Dorsally the colour is pale fawn with the following dark-brown markings: 4 or 2 dorsal lines from neck to middle of tail (on which they break up into series of dots), blotches on the head and/or dark etching between the scales, and a bold dark stripe from the lores (the area between eyes and nostril) through the orbit to the base of the tail. Dorsal colour is sometimes faded. Lips, ventrolateral surfaces, and belly are whitish. Belly sometimes very lightly dark flecked.

Under the toes is dark grey rather than the white of the other ventral surfaces.

Distribution: *A. edwardsae gotit subsp. nov.* is a taxon from the arid region west of Lake Torrens, South Australia.

It is relatively range restricted but due to the minimal human activity in the area, is not believed to be under any known threat.

Etymology: In 2010 I did a reptile survey in conjunction with Tom Cotton in the arid region west of Lake Torrens, South Australia and when he grabbed a fleeing specimen he yelled "*gotit*" and hence the subspecies name.

AAAH EDWARDSAE HELLYEAH SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:EFA05FF2-C968-4141-BBC6-7B29F72DB620

Holotype: A preserved female specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R67130 collected from 21.4 km west north-west of Mount Tabby (summit), South Australia, Australia, Latitude -31.39854 S., Longitude 131.2811 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the South Australian Museum, Adelaide, South Australia,

Australia, specimen number R67133 collected from 21.4 km west north-west of Mount Tabby (summit), South Australia, Australia, Latitude -31.39854 S., Longitude 131.2811 E.

Diagnosis: Hoser (2024c) formally named *Aaah edwardsae skink* Hoser, 2024 which had treated as the eastern population of *Aaah edwardsae* (Storr, 1982), better known as "*Lerista edwardsae*" with a type locality of Streaky Bay. South Australia, Australia (Latitude -32.50 S., Longitude 134.15 E.), being on the western edge of the Eyre Peninsula.

Aaah edwardsae skink Hoser, 2024 is a taxon of the Spencer Gulf and the lower Eyre Peninsula in South Australia.

Nominate *Aaah edwardsae* (Storr, 1982) is a taxon found roughly from Streaky Bay in the east to near Penong in the West in South Australia.

Two other subspecies are formally named for the first time within this paper.

These are *Aaah edwardsae gotit subsp. nov.* from the arid region west of Lake Torrens, South Australia and *Aaah edwardsae hellyeah subsp. nov.* from a 20 km radius around Yalata, South Australia.

The four subspecies are separated from one another by the following combinations of characters:

Nominate *Aaah edwardsae* (Storr, 1982) has:

1/ A beige coloured dorsum (versus light brown in *Aaah edwardsae skink* Hoser, 2024), 2/ Very light, broken or no dark etching between the scales on the upper surface of the head as also seen in *A. edwardsae gotit subsp. nov.* (versus heavy in *A. edwardsae skink* Hoser, 2024 and *A. edwardsae hellyeah subsp. nov.*), 3/ A dark lateral band that is medium thickness (versus thick *A. edwardsae skink* Hoser, 2024 or thin in *A. edwardsae hellyeah subsp. nov.*) 4/ Yellow upper labials, that are incompletely barred or etched on the lower parts (of some), but etched black at the tops and on adjoining scales, and 5/ Most specimens of this subspecies and *A. edwardsae hellyeah subsp. nov.* have two supraoculars in contact with the frontal, versus the reverse (one in contact) in *A. edwardsae skink* Hoser, 2024 and *A. edwardsae gotit subsp. nov.*

Aaah edwardsae skink Hoser, 2024 has:

1/ A generally darker coloured dorsum than the other three subspecies (being light brown rather than beige), 2/ Thick dark etching between the scales on the upper surface of the head (versus light or none in nominate *A. edwardsae* and *Aaah edwardsae gotit subsp. nov.*), 3/ A dark lateral band that is very thick (versus medium in *A. edwardsae* and *Aaah edwardsae gotit subsp. nov.* or relatively thin in *A. edwardsae hellyeah subsp. nov.*), 4/ Yellow upper labials, that are heavily barred black on the bottom (etchings) becoming black entirely around the upper edge and on adjoining scales, and 5/ Most specimens of this subspecies and *A. edwardsae gotit subsp. nov.* have only one supraocular in contact with frontal, versus the reverse in nominate *A. edwardsae* and *A. edwardsae hellyeah subsp. nov.* (2 in contact).

Aaah edwardsae gotit subsp. nov. has:

1/ A beige coloured dorsum (versus light brown in *Aaah edwardsae skink* Hoser, 2024), 2/ Light, broken or no dark etching between the scales on the upper surface of the head, like seen in nominate *A. edwardsae* (versus heavy in *Aaah edwardsae skink* Hoser, 2024 and *A. edwardsae hellyeah subsp. nov.*), 4/ White or whitish yellow upper labials, without dark etching on the lower edges, versus obviously yellow labials in the other three subspecies, and in this subspecies still with black entirely around the upper edge and on adjoining scales, and 5/ Most specimens of this subspecies and *A. edwardsae skink* Hoser, 2024 have only one supraocular in contact with the frontal, versus the reverse in nominate *A. edwardsae* and *A. edwardsae hellyeah subsp. nov.* (2 in contact).

Aaah edwardsae hellyeah subsp. nov. has:

1/ A beige coloured dorsum (versus light brown in *Aaah edwardsae skink* Hoser, 2024), 2/ Heavy dark etching between the scales on the upper surface of the head, like seen in *Aaah edwardsae skink* Hoser, 2024 (versus light, broken or not present in nominate *A. edwardsae* and *A. edwardsae gotit subsp. nov.*), 4/ Yellow upper labials, with dark etching in the form of triangles, the point on the lower edges, and in this subspecies still with black entirely around the upper edge and on adjoining scales, and 5/ Most specimens of this subspecies and nominate *A. edwardsae* have two supraoculars in contact with the frontal, versus the reverse (one in contact) in *A. edwardsae skink* Hoser, 2024 and *A. edwardsae gotit subsp. nov.*

A. edwardsae of the nominate subspecies is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/54817712249/

and

https://www.flickr.com/photos/127392361@N04/54817719148/

A. edwardsae skink Hoser, 2024 is depicted in life in Cogger (2014) on page 603 at top left and online at:

https://www.flickr.com/photos/reptileshots/55168787713/

and

https://www.flickr.com/photos/ryanfrancis/16587119909/

and

https://www.inaturalist.org/observations/325999588

and

https://www.inaturalist.org/observations/206751709

A. edwardsae gotit subsp. nov. is depicted in life

online at:
https://www.flickr.com/photos/127392361@N04/49231437681/

specimen from Pernatty Lagoon, South Australia, Australia, and.

https://www.inaturalist.org/observations/269583256

specimen from Unincorp, South Australia, Australia.

A. edwardsae hellyeah subsp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/154750972

specimen from the Eyre Highway, Yalata, South Australia, Australia.

A. edwardsae and the other preceding named subspecies are separated from the closely related *A. picturata* (Fry, 1914) by having a much shorter foreleg, fewer dorsal lines, as in just 2 instead of four and usually less mid body rows (18-20 versus 20-22).

The preceding taxa are in turn separated from *A. baynesi* (Storr, 1972) (of both subspecies as identified in Hoser 2024c) by having a strong rather than faded dorsal colouration and 9-14 rather than 8-10 subdigital lamellae under the second toe.

The preceding species, being the entirety of the genus *Aaah gen. nov*. are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) and assuming placement of all relevant species within these genera, (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) by the following unique combination of characters:

All three species (and four additional subspecies) in the genus are thick set for burrowing skinks.

They have one or other of 2 front digits (tiny first digit), or just one or sometimes none with a style, as well as always 2 digits at the rear; movable eyelid, nasals forming a median suture, frontoparietals paired, and an enlarged second supraocular excluding or nearly excluding first from contact with the frontal. Adult snout-vent length 64-95 mm. Length of appendages as a percent of snout vent length (SVL) is: fore-leg 0.5-1.7; hind-leg 11.1-18.9; snout to foreleg 21.9-25.7.

Nasals in short to long contact. Prefrontals widely separated. Frontoparietals separated, much smaller than the interparietal. 2-5 nuchals, 3 supraoculars, second always and first sometimes in contact with the frontal. Supraciliaries usually 0+1 or rarely 0+2 or 0+3. 6 supralabials. 2-3 temporals, usually 3. Upper secondary temporal largest, lower secondary temporal much the smallest.

18-22 Midbody scale-rows (usually 20). 8-15 lamellae under the longer toe.

Dorsally the colour is pale fawn with the following dark-brown markings: 4 or 2 dorsal lines from neck to middle of tail (on which they break up into series of dots), blotches on the head and/or dark etching between the scales, and a bold dark stripe from the lores (the area between eyes and nostril) through the orbit to the base of the tail. Dorsal colour is sometimes faded. Lips, ventrolateral surfaces, and belly are whitish. Belly sometimes very lightly dark flecked.

Under the toes is dark grey rather than the white of the other ventral surfaces.

Distribution: *A. edwardsae hellyeah subsp. nov.* is a taxon known only from a small area within a 20 km radius around Yalata, South Australia, Australia.

It is relatively range restricted but due to the minimal human activity in the area, is not believed to be under any known threat.

Etymology: When I mentioned to a colleague that I was going to formally name this subspecies he exclaimed "*hellyeah*"!

ACDC THUNDERSTRUCK SP. NOV.

LSIDurn:lsid:zoobank.org:act:36025BC7-8AB8-42BF-B2DF-1F66BA8B4A90

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia,

Australia, specimen number R25268 collected from leaf litter in a wall crevice in Kookhabinna Gorge, Barlee Range, Western Australia, Australia, Latitude -23.266667 S., Longitude 116.033333 E.

This government-owned facility allows access to its holdings.

Paratypes: Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R102228 and R102229 both collected from the Barlee Range Nature Reserve, Western Australia, Australia, Latitude -23.411389 S., Longitude 115.893889 E.

Diagnosis: Until now, both *Acdc tnt sp. nov*. and *Acdc thunderstruck sp. nov*. were treated as outlier populations of the Pilbara (Australia) taxon *Acdc flammicauda* (Storr, 1985) (type locality Marandoo, Western Australia, Australia, Latitude 22.37 S., Longitude 118.08E.), which was originally placed by Storr and most subsequent authors in the genus *Lerista* Bell, 1833.

The three species (as defined herein) are distributionally disjunct, being separated by major known biogeographical barriers, have clearly been separated for millions of years (certainly in excess of 1.5 MYA based on the age of the barriers in their present forms and available molecular data that is publicly available) and are morphologically divergent.

Therefore, I have no hesitation in formally describing and naming the outlier populations as new species.

Acdc tnt sp. nov. occupies the north-west Pilbara District of Western Australia, generally north of the Fortescue River.

Acdc flammicauda (Storr, 1985) occupies the main Pilbara District south of the Fortescue River and north of the Ashburton River.

Acdc thunderstruck sp. nov. appears to be restricted to the Barlee Range area, south of the Ashburton River.

The three species are separated from one another by the following character combinations:

Acdc flammicauda (Storr, 1985) has an (original) tail that is flame red with distinct and well-defined black spots on the upper and lateral surfaces that are closely spaced. The upper and mid flank is occupied by a bold black stripe. Beneath this the scales on the lower flank are white with dull grey spots or flecks, the most prominent forming a vague dull grey line. Lower edges of anterior and middle upper labials are thinly etched dark or black.

Acdc tnt sp. nov. has an (original) tail that is orange and without any or obvious spots or flecks on the upper surface of the tail and scattered, often dull spots on the lateral parts of the tail. The upper and mid flank is occupied by a bold black stripe. Beneath this the scales on the lower flank are white with distinct grey spots or flecks, the most prominent forming a grey line along the otherwise white lower lateral edge. Lower edges of anterior and middle upper labials are thickly etched dark or black, becoming thicker as one moves up the scale, giving the dark parts a triangular shape (the point at the lower edge and the base at the top of the labial).

Acdc thunderstruck sp. nov. is an overall more yellowish coloured lizard than the two preceding species. The (original) tail is a yellowish-orange colour, versus flame red. There are no black spots or flecks on the upper surface of the tail and those on the side are generally around the mid-lateral line, where they are tiny and closely spaced. Anterior edge of each lower lateral scale is dull grey, the rest being white. Thin dark etching (not forming triangles) on two or three of the anterior upper labials only.

The only other species in the genus *Acdc* Hoser, 2014 is *A. zietzi* (Wells and Wellington, 1985) (AKA the junior synonym "*Lerista chalybura* Storr, 1985").

It is separated from the preceding three species by having a bluish or greyish tail (instead, of red or orange), and a dark upper lateral stripe that is inferiorly ill-defined and less than one scale wide, versus sharp edged and more than one scale wide in the other three species.

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(2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Four fingers, four toes, movable eyelids, three supraoculars (first two in contact with the frontal) and five supraciliaries (second and fifth being the smallest). Ear aperture is smaller than the nostril (occasionally as large); foreleg-hindleg ratio 0.33-0.56; Dorsal pattern of two or four dark stripes or lines of spots usually well developed; under tail pink, usually immaculate; black upper lateral stripe wide and sharply defined. Nasals usually narrowly separated, but sometimes in short contact. Widely separated prefrontals. Frontoparietals in short to long contact. 3 supraoculars with the first two in contact with the frontal. 5 supraciliaries with the second and last the smallest, or rarely 4 or 6. Six or rarely seven upper labials, 16-22 midbody rows, (usually 18). 12-21 lamellae under the third toe.

Ear aperture is about twice as large as the nostril and the foreleg-hindleg ratio is 0.56-0.69.

These last two characters separate this genus from the morphologically similar genus *Oh gen. nov*. from the central Australian Ranges and southern Australia, which instead has an ear aperture that is smaller than the nostril (rarely occasionally as large) and a foreleg-hindleg ratio of 0.33-0.56.

The genus *Acdc* Hoser, 2024 is found in North-west Australia in the region of the Pilbara and just south of there.

Relevant Comments: The name "*Lerista zietzi* Wells and Wellington, 1985", type locality of 9 miles south of Wittenoom, Western Australia, Australia (in the Hamersley Ranges within Karijini National Park), Latitude -22.3734 S., Longitude 118.3362 E., has date priority over the name "*Lerista chalybura* Storr, 1985" type locality, Weeli Wolli Spring, Western Australia, Australia, Latitude -22.55 S., Longitude 119.12 E. published later that year.

Both are of the same taxon.

"*Lerista zietzi* Wells and Wellington, 1985" is the senior synonym of "*Lerista chalybura* Storr, 1985".

Hence the correct ICZN species nomen to use is "*zietzi*".

A number of non ICZN sites such as Peter Uetz's "The Reptile Database" deliberately use the junior synonym instead, lying and pretending that it is the correct ICZN name, knowing that it is not and also pretends that "*Lerista zietzi* Wells and Wellington, 1985" does not exist. This remained the case as recently as last checked in early 2026 and again on 8 June 2026.

Wells (2012) placed the species in this genus (*Acdc gen. nov*.) and the genus *Oh gen. nov*. into his genus *Tychismia* Wells, 2012 (type species *Lerista chordae* Amey, Kutt and Hutchinson 2005). They are not closely related based on the phylogeny published by Skinner *et al*. (2008) and on my reassessment of the relevant species, two new genera were erected in Hoser (2024c) to accommodate the relevant taxa.

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See Uetz (2022a, 2022b) for details.

A close Peter Uetz associate, and member of the Wüster / Britton / Kaiser gang, Adam Britton of McMinns Lagoon, Darwin, Northern Territory Australia pled guilty to multiple offences involving the torture, rape and killing of dogs, and the possession of child abuse material.

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More details at:
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The commonality of taxonomic vandalism, egregious animal abuse and other serious crimes are a direct pathology of the Narcissistic Personality Disorder (NPD) that affects these gang members in the trait of zero regards for rules and welfare of others. They are standard traits of these people, although I note that as a rule they take extraordinary steps to conceal their acts of evil from other people.

This is done by false narratives, lies, deflections, blame shifting, controlling others and ruthlessly stopping others from exposing their ill deeds. See

also the etymology for *Gaia narcissismawareness sp. nov*. in this paper.

Distribution: *Acdc thunderstruck sp. nov*. occupies the north-west Pilbara District of Western Australia, generally north of the Fortescue River.

Etymology: The genus name is in honour of the Australian rock band AC/DC formed in Sydney in 1973. One of their greatest hit songs was "Thunderstruck". It was known for Angus Young's blistering, tapping intro riff and one of their most highly certified hits globally. It was released in 1990. This is the etymology for this species.

ACDC TNT SP. NOV.

LSIDurn:lsid:zoobank.org:act:572FF076-7FF6-47A0-A884-0E0B78BC0E9D

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R157775 collected from 4 km southwest of Zebra Hill, Western Australia, Australia, Latitude -21.218056 S., Longitude 116.825833 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R131007 collected from the Millstream-Chichester National Park, Western Australia, Australia, Latitude -21.312778 S., Longitude 117.233889 E.

Diagnosis: Until now, both *Acdc tnt sp. nov*. and *Acdc thunderstruck sp. nov*. were treated as outlier populations of the Pilbara (Australia) taxon *Acdc flammicauda* (Storr, 1985) (type locality Marandoo, Western Australia, Australia, Latitude 22.37 S., Longitude 118.08E.), which was originally placed by Storr and most subsequent authors in the genus *Lerista* Bell, 1833.

The three species (as defined herein) are distributionally disjunct, being separated by major known biogeographical barriers, have clearly been separated for millions of years (certainly in excess of 1.5 MYA based on the age of the barriers in their present forms and available molecular data that is publicly available) and are morphologically divergent.

Therefore, I have no hesitation in formally describing and naming the outlier populations as new species.

Acdc tnt sp. nov. occupies the north-west Pilbara District of Western Australia, generally north of the Fortescue River.

Acdc flammicauda (Storr, 1985) occupies the main Pilbara District south of the Fortescue River and north of the Ashburton River.

Acdc thunderstruck sp. nov. appears to be restricted to the Barlee Range area, south of the Ashburton River.

The three species are separated from one another by the following character combinations:

Acdc flammicauda (Storr, 1985) has an (original) tail

that is flame red with distinct and well-defined black spots on the upper and lateral surfaces that are closely spaced. The upper and mid flank is occupied by a bold black stripe. Beneath this the scales on the lower flank are white with dull grey spots or flecks, the most prominent forming a vague dull grey line. Lower edges of anterior and middle upper labials are thinly etched dark or black.

Acdc tnt sp. nov. has an (original) tail that is orange and without any or obvious spots or flecks on the upper surface of the tail and scattered, often dull spots on the lateral parts of the tail. The upper and mid flank is occupied by a bold black stripe. Beneath this the scales on the lower flank are white with distinct grey spots or flecks, the most prominent forming a grey line along the otherwise white lower lateral edge. Lower edges of anterior and middle upper labials are thickly etched dark or black, becoming thicker as one moves up the scale, giving the dark parts a triangular shape (the point at the lower edge and the base at the top of the labial).

Acdc thunderstruck sp. nov. is an overall more yellowish coloured lizard than the two preceding species. The (original) tail is a yellowish-orange colour, versus flame red. There are no black spots or flecks on the upper surface of the tail and those on the side are generally around the mid-lateral line, where they are tiny and closely spaced. Anterior edge of each lower lateral scale is dull grey, the rest being white. Thin dark etching (not forming triangles) on two or three of the anterior upper labials only.

The only other species in the genus *Acdc* Hoser, 2014 is *A. zietzi* (Wells and Wellington, 1985) (AKA the junior synonym "*Lerista chalybura* Storr, 1985").

It is separated from the preceding three species by having a bluish or greyish tail (instead, of red or orange), and a dark upper lateral stripe that is inferiorly ill-defined and less than one scale wide, versus sharp edged and more than one scale wide in the other three species.

The genus *Acdc* Hoser, 2024 is separated from all other species within the putative genus *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

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The genus *Acdc* Hoser, 2024 is found in North-west Australia in the region of the Pilbara and just south of there.

Relevant Comments: The name "*Lerista zietzi* Wells and Wellington, 1985", type locality of 9 miles south of Wittenoom, Western Australia, Australia (in the Hamersley Ranges within Karijini National Park), Latitude -22.3734 S., Longitude 118.3362 E., has date priority over the name "*Lerista chalybura* Storr, 1985" type locality, Weeli Wolli Spring, Western

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This is done by false narratives, lies, deflections, blame shifting, controlling others and ruthlessly stopping others from exposing their ill deeds. See also the etymology for *Gaia narcissismawareness sp. nov*. in this paper.

Distribution: *Acdc tnt sp. nov*. occupies the north-west Pilbara District of Western Australia, generally north of the Fortescue River.

Etymology: The genus name is in honour of the Australian rock band AC/DC formed in Sydney in 1973. One of their greatest hit songs was "TNT". It was an explosive, crowd-pumping staple from the band's early pub-rock days and was released in 1975. This is the etymology for this species.

A REDEFINITION OF THE GENUS *AH* HOSER, 2024.

The molecular results of Torkkola *et al.* (2026) showed three lineages (AKA species) of lizards previously treated as northern populations of *Miculia elegans* Gray, 1845 as defined in Hoser (2024c), were in fact part of the lineage of the genus *Ah* Hoser, 2020 (type species *Ah ha* Hoser, 2024), and also including the better known species *Ah christinae* (Storr, 1979), that was originally placed in the genus *Lerista* Bell, 1833.

The closest related to genus to *Ah* Hoser, 2024 is *Miculia* Gray, 1845 and as a result of the identification of three new species within *Ah* Hoser, 2024 that are convergent in form with species within *Miculia* (all formally named for the first time in this paper) the genus *Ah* Hoser, 2024 had to be redefined. This is done below:

***AH* HOSER, 2024.**

Type species: *Ah ha* Hoser, 2024.

Diagnosis: The species within the genus *Ah gen. nov*. are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa*

boulengeri Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024 (in this case excluded), *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

A slender burrowing skink with 4 digits on front and back limbs and a fixed eyelid. Under toes are greyish brown, the rest of the venter is white, becoming buffy under the tail.

Nasals in medium to long contact. Prefrontals widely separated. Frontoparietals in medium to long contact, smaller than interparietal. 2-5 nuchals. 3 supraoculars, the first two in contact with the frontal. 4 supraciliaries, first three are subequal and last by far the smallest. 6 supralabials, rarely 7. 3 temporals, upper secondary temporal much the largest, lower secondary much the smallest. 16, 18 or more midbody scale-rows. long and narrow interparietal, 16-18 lamellae under the fourth toe and one or other of the following:

A/ Dorsum and flanks light brown to whitish and boldly patterned with 6 sharp edged moderately thick longitudinal lines and 16 midbody scale rows, or:

B/ In the alternative, 18 or more midbody rows and a dorsum that is essentially unicolour on top; there is a relatively thin, sometimes broken, black line that runs from behind the eye, along the upper flank and to the anterior part of the (original) tail, where it ends, this being different to the thick unbroken line that runs along the upper flank and the entire length of the (original) tail in morphologically similar *Miculia* species from south-west Australia (e.g. *M. elegans* Gray, 1845, *M. distinguenda* (Werner, 1910), *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.*).

Note that the divergent taxon *M. ruficauda* Hoser, 2024, of the northern Eyre Peninsula in South Australia is morphologically similar to species of *Ah* Hoser, 2024, but separated from them by the fact that the white on the lower flank has an orange tinge. It is the only species within *Miculia* Gray, 1845 in which the upper lateral band is narrow (rather than thick) and unlike others in the genus, it does not run along the tail. Furthermore, there are four well defined rows of dots running down the dorsum versus two (well defined rows) in the other species of *Miculia.*

Species within *Ah* Hoser, 2024 are separated from the morphologically similar *Miculia distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by having nasals forming a median suture, long and narrow interparietal (versus separated nasals and a shorter and broader interparietal).

Distribution: Western Australia in a restricted coastal and near coastal zone between Perth / Rottnest Island in the South-west, (and possibly further south), extending north along the coast to Onslow, Western Australia and also including Barrow Island and the Monte Bello Islands to the north.

Etymology: From Hoser (2024c): Recalling the first time I saw a new species in the genus, I exclaimed "*ah ha*", hence both the genus name and the name of the new species described immediately following this (genus) description.

The genus and species name are deliberately chosen to ignite interest among people in this generally neglected group of reptiles.

Content: *Ah ha* Hoser, 2024 (type species); *Ah christinae* (Storr, 1979), *Ah yess sp. nov.* (this paper); *Ah pfuk sp. nov.* (this paper); *Ah sheat sp. nov.* (this paper).

AH YESS SP. NOV.

LSIDurn:lsid:zoobank.org:act:C8966E7A-B53C-4DD6-B84D-A0F5E317BFDB

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R121393 collected from Kalbarri, Western Australia, Australia, Latitude -27.7 S., Longitude 114.166667 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ Three preserved specimens at the Australian Museum, Sydney, Australia, specimen numbers R.134247, R.134202 and R.134248 all collected from a site roughly 21 km south of Kalbarri via Balline-Kalbarri Road, Western Australia, Australia, Latitude -27.866667 S., Longitude 114.15 E.

2/ Three preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, being specimen number R141851, collected from 5 km east of Arrowsmith, Western Australia, Australia, Latitude -29.542222 S., Longitude 115.0675 E., specimen number R88770 collected from beneath Acacia litter 25 km south of Dongara, Western Australia, Australia, Latitude -29.583333 S., Longitude 115.0 E., and specimen number R73114 collected from 16 km north of Eneabba, Western Australia, Australia, Latitude -29.683333 S., Longitude 115.233333 E.

Diagnosis: It has been known for many years that there is significant species-level divergence between populations of putative "*Lerista elegans* (Gray, 1845)" a taxon restricted to the coast and hinterland in Western Australia, Australia.

Wells and Wellington (1984, 1985) restored the putative species to the genus *Miculia* Gray, 1845, agreed by Wells (2012) and then Hoser (2024c), who as a cohort are alone in publishing herpetologists in accepting the reality of herpetological classification and nomenclature in Australia, without resorting to petty boycotting of works of people outside the Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang's name censorship club (see Hoser 2024c for further details and citations).

The description for Gray's holotype in Gray (1845) was vague, but did mention the following:

"*Olive, with a dark streak on each side, with a narrow white edge below; beneath pale whitish; tail brown-dotted; eyelid interrupted above*."

The only specimens of putative *Miculia elegans* Gray, 1845, that match this description are those from the Perth area extending north to about Guilderton and Gingin, north of where they are significantly lighter in colour and with a reduced dark streak on either side making the "*narrow white edge below*" somewhat wider.

In addition to this, the holotype was from "*Mr. Gilbert's Collection*", which as far as is known (based on Gray, 1845) only included specimens from the Swan River (Perth), Houtman Abrolhos (Islands off the West Australian coast) and Port Essington (Northern Territory).

As putative *Miculia elegans* Gray, 1845 AKA "*Lerista elegans* (Gray, 1845)´ do not occur in either other location, this confirms that the type locality is Perth, Western Australia and the taxon from there is of the type form.

Papers of Singhal *et al.* (2025) and Torkkola *et al*. (2026) confirmed the presence of at least three other species within putative "*Lerista elegans*".

Significantly these other three species that as a cohort are found along the Western Australian coast from about Jurian Bay in the south to Onslow in the north and then beyond to Barrow Island and the Monte Bello Islands, while morphologically are convergent with *Miculia elegans* Gray, 1845, they are in fact most closely related to the species *Ah christinae* (Storr, 1979) and *Ah ha* Hoser, 2024.

These are the three new species that are named herein.

They are as follows:

Ah yess sp. nov. is found along the west coast and nearby hinterland of Western Australia from just north of Kalbarri, Western Australia south to about Jurian Bay (encompassing an area known as the lower Coral Coast).

Ah pfuk sp. nov. is found in the region from Nerren Nerren in the south, along the coast and nearby hinterland north to Exmouth, Onslow, Barrow Island and the Montebello Islands.

Ah sheat sp. nov. is apparently restricted to the Kennedy Range area of western Western Australia.

The morphologically convergent species *Miculia elegans* Gray, 1845 is in effect confined to the region south of Jurian Bay, along the West Australian coastline and nearby hinterland south through Perth to Augusta.

The four species are readily separated from one another by the following unique combinations of characters (in life):

Miculia elegans is a dark, almost chocolate brown lizard with a thick blackish brown stripe running from the eye, along the upper flank and onto the (original) tail, where it remains well defined and thick. The lower border of this stripe is a well-defined white line that is bounded below by an area of greyish scales, turning whitish below or alternatively whitish scales bordered by dull grey. The tail is usually the same colour as the body or alternatively dull orange posteriorly. Running down the dorsum are two rows of poorly defined black dots. There is limited black speckling on the upper surface of the head.

Ah yess sp. nov. is a light brown to beige lizard on top.

The dark streak running from eye along upper flank is slightly thinner than seen in *M. elegans* and in this species barely runs onto the (original) tail, instead dissipating quickly.

The tail itself is a dull orange colour, except for the very anterior part. Markings on the tail are semi distinct at best and mainly in the form of peppering.

The two rows of spots running down the dorsum are dark brown rather than black.

Below the dark line on the flank, the scales are white with dark brown etching, but of same intensity from top to bottom of the lower flank. There is a lot of dark speckling on the upper surface of the head.

Ah pfuk sp. nov. is a light greyish to greyish beige on top. The dark stripe from back of the eye along the flank is very thin along its entire length and becomes

an extremely thin line as it runs along the anterior of the side of the tail before dissipating. Tail is usually a bright orange colour except for the very anterior part and without any obvious flecks, peppering or markings, except the sides of the anterior part. There are limited, but distinct black flecks on the upper surface of the head.

The lower flank scales are immaculate white, or rarely etched, but if etched, they are etched indistinctly.

Some specimens have a distinctive row of black dots running along the lower flank.

Ah sheat sp. nov. is a light pinkish reddish beige on top and otherwise like *Ah pfuk sp. nov.* as described above but is separated from that species by its dorsal colouration combined with the fact that the upper labials are dark etched, versus immaculate or near immaculate in *Ah pfuk sp. nov.*.

The four preceding species *M. elegans, Ah yess sp. nov.*, *A. sheat sp. nov.* and *A. pfuk sp. nov.* are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 4 front and 4 back and a fixed eyelid.

Separated from the morphologically similar *Miculia distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by having 16 midbody scale-rows and nasals forming a median suture, long and narrow interparietal (versus 18 or more scale-rows separated nasals and shorter and broader interparietal).

The type form of *M. elegans* is depicted in life in Cogger (2014) on page 603 at top right, Wilson and Swan (2021) on page 363, second from bottom on right, and online at:
https://www.inaturalist.org/observations/112910784
and
https://www.inaturalist.org/observations/315138713
and
https://www.inaturalist.org/observations/130490997

A. yess sp. nov. is depicted in life online at:
https://www.flickr.com/photos/colonel_007/49087636698/
and
https://www.inaturalist.org/observations/351635991

A. pfuk sp. nov. is depicted in life online at:
https://www.flickr.com/photos/114192916@N07/54080412575/
and
https://www.flickr.com/photos/114192916@N07/54080200843/
and
https://www.flickr.com/photos/julesfarquhar/52681798571/
and
https://www.flickr.com/photos/127392361@N04/54012354380/
and
https://www.flickr.com/photos/114192916@N07/54079073647/
and
https://www.flickr.com/photos/58349528@N02/52258254730/
and
https://www.flickr.com/photos/128497936@N03/49113912641/
and
https://www.flickr.com/photos/114192916@N07/54079074552/

Distribution: *A. yess sp. nov*. is found along the west coast and nearby hinterland of Western Australia from just north of Kalbarri, Western

Australia to about Jurian Bay (encompassing an area known as the lower Coral Coast).

Etymology: "*Yess*" is a common exclamation a person makes when one grabs an escaping specimen successfully without causing it to shed its tail.

The spelling of the species name is deliberate, to avoid creation of a potential homonym and must not be amended.

AH PFUK SP. NOV.

LSIDurn:lsid:zoobank.org:act:DDC6DFBB-1076-4F4E-8B8C-C0123E371E6E

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R120425 collected from 40KM south south-east of Carnarvon, Western Australia, Australia, Latitude -25.125833 S., Longitude 113.822778 E., from an area with reddish brown sandy alluvium with low red sandy rises; *Atriplex* (on flats) with *Acacia* thickets (on rises).

This government-owned facility allows access to its holdings.

Paratypes: 1/ A preserved female specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R122983 collected from Nerren Nerren station, Western Australia, Australia, Latitude -27.005833 S., Longitude 114.799167 E., 2/ A preserved female specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R153823 collected from Bullara Station, Western Australia, Australia, Latitude -22.890556 S., Longitude 113.923611 E.

Diagnosis: It has been known for many years that there is significant species-level divergence between populations of putative "*Lerista elegans* (Gray, 1845)´ a taxon restricted to the coast and hinterland in Western Australia, Australia.

Wells and Wellington (1984, 1985) restored the putative species to the genus *Miculia* Gray, 1845, agreed by Wells (2012) and then Hoser (2024c), who as a cohort are alone in publishing herpetologists in accepting the reality of herpetological classification and nomenclature in Australia, without resorting to petty boycotting of works of people outside the Wolfgang Wüster / Adam Britton /Hinrich Kaiser gang's name censorship club (see Hoser 2024c for further details and citations).

The description for Gray's holotype in Gray (1845) was vague, but did mention the following:

"*Olive, with a dark streak on each side, with a narrow white edge below; beneath pale whitish; tail brown-dotted; eyelid interrupted above*."

The only specimens of putative *Miculia elegans* Gray, 1845, that match this description are those from the Perth area extending north to about Guilderton and Gingin, north of where they are significantly lighter in colour and with a reduced dark streak on either side making the "*narrow white edge below*" somewhat wider.

In addition to this, the holotype was from "*Mr. Gilbert's Collection*", which as far as is known (based on Gray, 1845) only included specimens from the Swan River (Perth), Houtman Abrolhos (Islands off the West Australian coast) and Port Essington (Northern Territory).

As putative *Miculia elegans* Gray, 1845 AKA "*Lerista elegans* (Gray, 1845)" do not occur in either other location, this confirms that the type locality is Perth, Western Australia and the taxon from there is of the type form.

Papers of Singhal *et al.* (2025) and Torkkola *et al*. (2026) confirmed the presence of at least three other species within putative "*Lerista elegans*".

Significantly these other three species that as a cohort are found along the Western Australian coast from about Jurian Bay in the south to Onslow in the north and then beyond to Barrow Island and the Monte Bello Islands, while morphologically are convergent with *Miculia elegans* Gray, 1845, they are in fact most closely related to the species *Ah christinae* (Storr, 1979) and *Ah ha* Hoser, 2024.

These are three new species are named herein as follows:

Ah yess sp. nov. is found along the west coast and nearby hinterland of Western Australia from just north of Kalbarri, Western Australia south to about Jurian Bay (encompassing an area known as the lower Coral Coast).

Ah pfuk sp. nov. is found in the region from Nerren Nerren in the south, along the coast and nearby hinterland north to Exmouth, Onslow, Barrow Island and the Montebello Islands.

Ah sheat sp. nov. is apparently restricted to the Kennedy Range area of western Western Australia.

The morphologically convergent species *Miculia elegans* Gray, 1845 is in effect confined to the region south of Jurian Bay, along the West Australian coastline and nearby hinterland south through Perth to Augusta.

The four species are readily separated from one another by the following unique combinations of characters (in life):

Miculia elegans is a dark, almost chocolate brown lizard with a thick blackish brown stripe running from the eye, along the upper flank and onto the (original) tail, where it remains well defined and thick. The lower border of this stripe is a well-defined white line that is bounded below by an area of greyish scales, turning whitish below or alternatively whitish scales bordered by dull grey. The tail is usually the same colour as the body or alternatively dull orange posteriorly. Running down the dorsum are two rows

of poorly defined black dots. There is limited black speckling on the upper surface of the head.

Ah yess sp. nov. is a light brown to beige lizard on top.

The dark streak running from eye along upper flank is slightly thinner than seen in *M. elegans* and in this species barely runs onto the (original) tail, instead dissipating quickly.

The tail itself is a dull orange colour, except for the very anterior part. Markings on the tail are semi distinct at best and mainly in the form of peppering.

The two rows of spots running down the dorsum are dark brown rather than black.

Below the dark line on the flank, the scales are white with dark brown etching, but of same intensity from top to bottom of the lower flank. There is a lot of dark speckling on the upper surface of the head.

Ah pfuk sp. nov. is a light greyish to greyish beige on top. The dark stripe from back of the eye along the flank is very thin along its entire length and becomes an extremely thin line as it runs along the anterior of the side of the tail before dissipating. Tail is usually a bright orange colour except for the very anterior part and without any obvious flecks, peppering or markings, except the sides of the anterior part. There are limited, but distinct black flecks on the upper surface of the head.

The lower flank scales are immaculate white, or rarely etched, but if etched, they are etched indistinctly.

Some specimens have a distinctive row of black dots running along the lower flank.

Ah sheat sp. nov. is a light pinkish reddish beige on top and otherwise like *Ah pfuk sp. nov.* as described above but is separated from that species by its dorsal colouration combined with the fact that the upper labials are dark etched, versus immaculate or near immaculate in *Ah pfuk sp. nov.*.

The four preceding species *M. elegans, A. yess sp. nov.*, *A. sheat sp. nov.* and *A. pfuk sp. nov.* are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 4 from and 4 back and a fixed eyelid.

Separated from the morphologically similar *Miculia distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by having 16 midbody scale-rows and nasals forming a median suture, long and narrow interparietal (versus 18 or more scale-rows separated nasals and shorter and broader interparietal).

The type form of *M. elegans* is depicted in life in Cogger (2014) on page 603 at top right, Wilson and Swan (2021) on page 363, second from bottom on right, and online at:
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A. pfuk sp. nov. is depicted in life online at:
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and
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and
https://www.flickr.com/photos/

julesfarquhar/52681798571/
and
https://www.flickr.com/photos/127392361@N04/54012354380/
and
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and
https://www.flickr.com/photos/58349528@N02/52258254730/
and
https://www.flickr.com/photos/128497936@N03/49113912641/
and
https://www.flickr.com/photos/114192916@N07/54079074552/

Distribution: *Ah pfuk sp. nov*. is found in the region from Nerren Nerren in the south, along the coast and nearby hinterland north to Exmouth, Onslow, Barrow Island and the Montebello Islands.

Etymology: When trying to catch this lizard as it escapes and the only thing you get is a wriggling tail, the common exclamation made is "*Ah pfuk*" and hence the species name "*pfuk*". The spelling name is deliberate to avoid a potential homonym and should not be amended or changed.

AH SHEAT SP. NOV.

LSIDurn:lsid:zoobank.org:act:BCFEE3D4-0770-4E2A-8B96-3AAADBE349DE

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R108629 collected from the number 1 oil well at the Kennedy Range National Park, Western Australia, Australia, Latitude -24.333333 S., Longitude 114.983333 E.

This government-owned facility allows access to its holdings.

Paratypes: Seven preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R120374, R120629, R121566, R121570, R122797, R122798 and R125394 all collected from 8 km northwest of Mardathuna Homestead, Western Australia, Australia in an area of red dunes with Acacia and Grevillea over spinifex.

Diagnosis: It has been known for many years that there is significant species-level divergence between populations of putative "*Lerista elegans* (Gray, 1845)´ a taxon restricted to the coast and hinterland in Western Australia, Australia.

Wells and Wellington (1984, 1985) restored the putative species to the genus *Miculia* Gray, 1845, agreed by Wells (2012) and then Hoser (2024c), who as a cohort are alone in publishing herpetologists in accepting the reality of herpetological classification and nomenclature in Australia, without resorting to petty boycotting of works of people outside the Wolfgang Wüster / Adam Britton / Hinrich Kaiser gang's name censorship club (see Hoser 2024c for further details and citations).

The description for Gray's holotype in Gray (1845) was vague, but did mention the following:

"*Olive, with a dark streak on each side, with a narrow white edge below; beneath pale whitish; tail brown-dotted; eyelid interrupted above*."

The only specimens of putative *Miculia elegans* Gray, 1845, that match this description are those from the Perth area extending north to about Guilderton and Gingin, north of where they are significantly lighter in colour and with a reduced dark streak on either side making the "*narrow white edge below*" somewhat wider.

In addition to this, the holotype was from "*Mr. Gilbert's Collection*", which as far as is known (based on Gray, 1845) only included specimens from the Swan River (Perth), Houtman Abrolhos (Islands off the West Australian coast) and Port Essington (Northern Territory).

As putative *Miculia elegans* Gray, 1845 AKA "*Lerista elegans* (Gray, 1845)" do not occur in either other location, this confirms that the type locality is Perth, Western Australia and the taxon from there is of the type form.

Papers of Singhal *et al.* (2025) and Torkkola *et al*. (2026) confirmed the presence of at least three other species within putative "*Lerista elegans*".

Significantly these other three species that as a cohort are found along the Western Australian coast from about Jurian Bay in the south to Onslow in the north and then beyond to Barrow Island and the Monte Bello Islands, while morphologically are convergent with *Miculia elegans* Gray, 1845, they are in fact most closely related to the species *Ah christinae* (Storr, 1979) and *Ah ha* Hoser, 2024.

These are three new species are named herein as follows:

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Ah pfuk sp. nov. is found in the region from Nerren Nerren in the south, along the coast and nearby hinterland north to Exmouth, Onslow, Barrow Island and the Montebello Islands.

Ah sheat sp. nov. is apparently restricted to the Kennedy Range area of western Western Australia.

The morphologically convergent species *Miculia elegans* Gray, 1845 is in effect confined to the region south of Jurian Bay, along the West Australian coastline and nearby hinterland south through Perth

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The four species are readily separated from one another by the following unique combinations of characters (in life):

Miculia elegans is a dark, almost chocolate brown lizard with a thick blackish brown stripe running from the eye, along the upper flank and onto the (original) tail, where it remains well defined and thick. The lower border of this stripe is a well-defined white line that is bounded below by an area of greyish scales, turning whitish below or alternatively whitish scales bordered by dull grey. The tail is usually the same colour as the body or alternatively dull orange posteriorly. Running down the dorsum are two rows of poorly defined black dots. There is limited black speckling on the upper surface of the head.

Ah yess sp. nov. is a light brown to beige lizard on top.

The dark streak running from eye along upper flank is slightly thinner than seen in *M. elegans* and in this species barely runs onto the (original) tail, instead dissipating quickly.

The tail itself is a dull orange colour, except for the very anterior part. Markings on the tail are semi distinct at best and mainly in the form of peppering.

The two rows of spots running down the dorsum are dark brown rather than black.

Below the dark line on the flank, the scales are white with dark brown etching, but of same intensity from top to bottom of the lower flank. There is a lot of dark speckling on the upper surface of the head.

Ah pfuk sp. nov. is a light greyish to greyish beige on top. The dark stripe from back of the eye along the flank is very thin along its entire length and becomes an extremely thin line as it runs along the anterior of the side of the tail before dissipating. Tail is usually a bright orange colour except for the very anterior part and without any obvious flecks, peppering or markings, except the sides of the anterior part. There are limited, but distinct black flecks on the upper surface of the head.

The lower flank scales are immaculate white, or rarely etched, but if etched, they are etched indistinctly.

Some specimens have a distinctive row of black dots running along the lower flank.

Ah sheat sp. nov. is a light pinkish reddish beige on top and otherwise like *Ah pfuk sp. nov.* as described above but is separated from that species by its dorsal colouration combined with the fact that the upper labials are dark etched, versus immaculate or near immaculate in *Ah pfuk sp. nov.*.

The four preceding species *M. elegans, A. yess sp. nov.*, *A. sheat sp. nov.* and *A. pfuk sp. nov.* are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 4 from and 4 back and a fixed eyelid.

Separated from the morphologically similar *Miculia distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by having 16 midbody scale-rows and nasals forming a median suture, long and narrow interparietal (versus 18 or more scale-rows separated nasals and shorter and broader interparietal).

The type form of *M. elegans* is depicted in life in Cogger (2014) on page 603 at top right, Wilson and Swan (2021) on page 363, second from bottom on right, and online at:
https://www.inaturalist.org/observations/112910784
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https://www.inaturalist.org/observations/130490997

A. yess sp. nov. is depicted in life online at:

https://www.flickr.com/photos/colonel_007/49087636698/

and

https://www.inaturalist.org/observations/351635991

A. pfuk sp. nov. is depicted in life online at:

https://www.flickr.com/photos/114192916@N07/54080412575/

and

https://www.flickr.com/photos/114192916@N07/54080200843/

and

https://www.flickr.com/photos/julesfarquhar/52681798571/

and

https://www.flickr.com/photos/127392361@N04/54012354380/

and

https://www.flickr.com/photos/114192916@N07/54079073647/

and

https://www.flickr.com/photos/58349528@N02/52258254730/

and

https://www.flickr.com/photos/128497936@N03/49113912641/

and

https://www.flickr.com/photos/114192916@N07/54079074552/

Distribution: *Ah sheat sp. nov.* is apparently restricted to the Kennedy Range area of western Western Australia, Australia.

Etymology: When trying to catch one of these lizards in the sand and it escapes, a common exclamation is "*Ah sheat*" and hence the species name "sheat". The spelling is deliberate and should not be changed as it may otherwise become a homonym.

APHRODITIA GERRARDII BADIMAYA SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:6A75408E-8C58-4C57-AB77-D18F96E3720A

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R95290 collected from under exfoliated granite at the base of an outcrop at Walganna Rock, Western Australia, Australia, Latitude -27.4 S., Longitude 117.466667 E.

This government-owned facility allows access to its holdings.

Paratype: Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R97028 and R97029 both collected from litter at Woolgerong Rock, Western Australia, Australia, Latitude -27.4 S., Longitude 117.383333 E.

Diagnosis: Until now, no subspecies have been recognised in the putative species *Aphroditia gerrardii* (Gray, 1864), with a type locality of Swan River, north-east of Perth, Western Australia, Australia.

The species as presently known occurs north to near Shark Bay and extending inland about 400 km.

Singhal *et al*. (2025) in their supplementary data recognised three so-called "Operational species" within putative "*Lerista gerrardii*" of west Western Australia, Australia.

The nominate form occupies most of the range, with outlier populations in the north-west (on the coast) and north-east herein formally recognised and named as new morphologically divergent subspecies.

The three subspecies are separated from one another by the following unique combinations of characters:

Nominate (type form) *Aphroditia gerrardii* is characterised with a C-shaped dark brown marking at the back of the parietal scale; light-brown dorsum, mainly greyish tail and a thick brown vertebral line that is generally slightly thinner than the light brown stripes on either side on the dorsum; tip of snout is not white.

A. gerrardii badimaya subsp. nov. has dark brown speckling on the rear of the head (no obvious "C" shape or big blotch of more-or-less circular shape); a thick brown vertebral stripe on the dorsum that is slightly wider than the light-yellow longitudinal stripes on either side. Upper surfaces of the hind limbs and the tail have a strong purplish colour; tip of snout is not white.

A. gerrardii nhanta subsp. nov. has a large dark brown spot occupying the region of the parietal scale on the head; a chocolate brown vertebral line that is slightly thinner than the light whitish yellow stripes on either side that occupy the rest of the dorsum; the vertebral line forming two rows of purplish-brown dots along the anterior of the upper part of the tail; tip of snout is white.

The three preceding species are separated from all other species within *Lerista* Bell, 1833, *sensu lato sensu* Hoser (2024c), or Cogger (2014), by the following combination of characters:

Large species to 90 mm snout vent length (usually about 75 mm), with digits 1 + 2 (finger sometimes reduced to style; rarely 2 fingers). Movable eyelid, separated nasals; interparietal free; frontoparietals paired, and dark brown vertebral stripe in turn bordered with light brown beige, yellow or light yellow stripes running down the rest of the dorsum of the body.

A. gerrardii gerrardii of the nominate form is depicted in life online at:

https://www.inaturalist.org/observations/109719936

A. gerrardii badimaya subsp. nov. is depicted in life online at:

https://www.flickr.com/photos/brian_busho/14452342191/

and

https://www.inaturalist.org/observations/363346300

A. gerrardii nhanta subsp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/317160205

Distribution: *A. gerrardii badimaya subsp. nov.* is only known from the vicinity of within 50 km of the type locality Walganna Rock, Western Australia, Australia, Latitude -27.4 S., Longitude 117.466667 E. but may be more widespread. In terms of conservation status, it should be regarded as insufficiently known.

Etymology: *A. gerrardii badimaya subsp. nov.* is named in recognition of the Badimaya tribe of Aboriginal Australians, as the original inhabitants of the area the species occurs. The subspecies name "*badimaya*" is deliberate and should not be Latinised to the form of "badimayaorum" unless mandated by ICZN ruling or similar binding rule.

APHRODITIA GERRARDII NHANTA SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:D6AF0C64-98FF-4A32-BCDA-9406FDA6C01E

Holotype: A preserved specimen at the Australian Museum, Sydney, New South Wales, Australia, specimen number R.134136 collected from the rubbish tip just west of Northampton, Western Australia, Australia, Latitude -28.35 S., Longitude 114.6 E.

This government-owned facility allows access to its holdings.

Paratypes: Six preserved specimens at the Australian Museum, Sydney, New South Wales, Australia, specimen numbers R.134141, R.134143, R.134170, R.134171, R.134172 and R.134175 all collected from the rubbish tip just west of Northampton, Western Australia, Australia, Latitude -28.35 S., Longitude 114.6 E.

Diagnosis: Until now, no subspecies have been recognised in the putative species *Aphroditia gerrardii* (Gray, 1864), with a type locality of Swan River, north-east of Perth, Western Australia, Australia.

The species as presently known occurs north to near Shark Bay and extending inland about 400 km.

Singhal *et al*. (2025) in their supplementary data recognised three so-called "Operational species" within putative "*Lerista gerrardii*" of west Western Australia, Australia.

The nominate form occupies most of the range, with outlier populations in the north-west (on the coast) and north-east herein formally recognised and named as new morphologically divergent subspecies.

The three subspecies are separated from one another by the following unique combinations of characters:

Nominate (type form) *Aphroditia gerrardii* is characterised with a C-shaped dark brown marking at the back of the parietal scale; light-brown dorsum, mainly greyish tail and a thick brown vertebral line that is generally slightly thinner than the light brown stripes on either side on the dorsum; tip of snout is not white.

A. gerrardii badimaya subsp. nov. has dark brown speckling on the rear of the head (no obvious "C" shape or big blotch of more-or-less circular shape); a thick brown vertebral stripe on the dorsum that is slightly wider than the light-yellow longitudinal stripes on either side. Upper surfaces of the hind limbs and the tail have a strong purplish colour; tip of snout is not white.

A. gerrardii nhanta subsp. nov. has a large dark brown spot occupying the region of the parietal scale on the head; a chocolate brown vertebral line that is slightly thinner than the light whitish yellow stripes on either side that occupy the rest of the dorsum; the vertebral line forming two rows of purplish-brown dots along the anterior of the upper part of the tail; tip of snout is white.

The three preceding species are separated from all other species within *Lerista* Bell, 1833, *sensu lato sensu* Hoser (2024c), or Cogger (2014), by the following combination of characters:

Large species to 90 mm snout vent length (usually about 75 mm), with digits 1 + 2 (finger sometimes reduced to style; rarely 2 fingers). Movable eyelid, separated nasals; interparietal free; frontoparietals paired, and dark brown vertebral stripe in turn bordered with light brown beige, yellow or light yellow stripes running down the rest of the dorsum of the body.

A. gerrardii gerrardii of the nominate form is depicted in life online at:

https://www.inaturalist.org/observations/109719936

A. gerrardii badimaya subsp. nov. is depicted in life online at:

https://www.flickr.com/photos/brian_busho/14452342191/

and

https://www.inaturalist.org/observations/363346300

A. gerrardii nhanta subsp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/317160205

Distribution: *A. gerrardii nhanta subsp. nov.* is only known from the vicinity of the type locality

Northampton, Western Australia, Australia, Latitude -28.35 S., Longitude 114.6 E, including adjacent coastal areas south of Shark Bay, but may be more widespread. In terms of conservation status, it should be regarded as insufficiently known.

Etymology: *A. gerrardii nhanta subsp. nov.* is named in recognition of the Nhanta tribe of Aboriginal Australians, as the original inhabitants of the area the species occurs. The subspecies name "*nhanta*" is deliberate and should not be Latinised to the form of "nhantaorum" unless mandated by ICZN ruling or similar binding rule.

APHRODITIA HARRIGANORUM SP. NOV.

LSIDurn:lsid:zoobank.org:act:0997E19C-8C8D-4050-801C-F76344BC776A

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R87456 collected from under rubbish at the Altona Homestead, Western Australia, Australia, Latitude -27.533333 S., Longitude 119.983333 E.

This government-owned facility allows access to its holdings.

Paratypes: Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R82572 and R82573 both collected from under mulga litter 18 km north-west of Nuendah Homestead, Western Australia, Australia, Latitude -26.883333 S., Longitude 120.3 E.

Diagnosis: *Aphroditia harriganorum sp. nov*. has until now been treated as a western population (from west-central western Australia) of the relatively widespread species *A. desertorum* (Sternfeld, 1919), type locality of Hermannsburg Mission, upper Finke River, N. T., designated by Mertens (1967).

A. desertorum occurs in most of the southern half of the Northern Territory, most of South Australia, except for the far south and south-east and into inland southern Western Australia extending west to north of the Goldfields (Kalgoorlie).

The species *Aphroditia harriganorum sp. nov*. appears to be restricted to a relatively small area bound by Leinster Downs Station, Latitude -27.975556 S., Longitude 120.299444 E. and Carnarvon Range, Latitude -25.317778 S., Longitude 120.870556 E. in the north, broadly extending about 150 km to the west from these two points.

Aphroditia harriganorum sp. nov. is readily separated from *A. desertorum* as follows:

A dorsum that is yellow, rather than beige-brown, bold, unbroken brown paravertebral lines, versus small and semi-distinct spots (well separated) forming the two paravertebral lines in *A. desertorum*, and an original tail that is a mottled white and brown, rather than white and red, white and grey or white and reddish grey. The top of the head is yellow and brown, versus brown with black spots or dark peppering in *A. desertorum.* Upper labials are yellow, versus whitish in *A. desertorum.* Anterior forelimbs are mainly yellow, versus mainly dark in *A. desertorum.*

Aphroditia harriganorum sp. nov. and *A. desertorum* are separated from all other species of skinks in the genus *Aphroditia* Wells, 2012 and all other species within the genus *Lerista* Bell, 1833 *sensu lato sensu* Hoser (2024c), or Cogger (2014), by the following unique combination of characters:

2 fingers, 3 toes, dark upper lateral stripe and 2 paravertebral rows of small dark spots (or as an unbroken line) on back and onto the tail.

The two preceding species (which were treated by Storr (1991) as one species) are further diagnosed (by Storr 1991) as follows:

Snout-vent length (mm): 39-93 (N 113, mean 73.4). Length of appendages (SVL): foreleg 2.1-5.8 (N 53, mean 3.9); hind leg 13.0-18.9 (N 55, mean 15.7); tail 85-119 (N 33, mean 103.2); snout to foreleg 22.0-28.7 (N 54, mean 24.6). Nasals narrowly to moderately separated (N 61) or in point contact (1). Prefrontals widely separated (fused to frontal on one side of one specimen). Frontoparietals narrowly to moderately separated (N 72), in point contact (9) or in short to moderate contact (10), slightly to considerably smaller than interparietal. Nuchals 1-4 (N 78, mean 2.8). Supraoculars 3, first two in contact with frontal. Supraciliaries 0 + 3 (N 105), I + 3 (4), 0+ 2 (2), I + I (I), I + 2 (1), 3 (1), 5 (I). Midbodyscale rows 18 (N 6), 19 (1), 20 (50), 21 (3) or 22 (5). Lamellae under longest toe 9-14 (N 77, mean 11.4).

Aphroditia harriganorum sp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/85836500

from Leonora, Western Australia, Australia.

A. desertorum is depicted in life in Cogger (2014) on page 600 at bottom from Mount Olga, Northern Territory, Wilson and Swan (2021) on page 361 at bottom, from Alice Springs, Northern Territory, Australia, Storr (1991) at Fig. 6 from Big Shot Bore, Western Australia and online at:

https://www.inaturalist.org/observations/247970903

and

https://www.inaturalist.org/observations/328874724

and

https://www.inaturalist.org/observations/273640557

The morphological differences between *Aphroditia harriganorum sp. nov*. and *A. desertorum* are reflected in the species-level genetic differences identified in Singhal *et al.* (2025).

Distribution: The species *Aphroditia harriganorum sp. nov*. appears to be restricted to a relatively small area bound by Leinster Downs Station, Latitude -27.975556 S., Longitude 120.299444 E. and

Carnarvon Range, Latitude -25.317778 S., Longitude 120.870556 E. in the north, broadly extending about 150 km to the west from these two points.

Etymology: This species is named in honour of the Harrigan family of Mildura, Victoria in recognition of their contributions to herpetology. This is Liz Harrigan, Darren Harrigan, their children and grandchildren.

GAIA AMAZING SP. NOV.

LSIDurn:lsid:zoobank.org:act:FF6F2DFB-BD0F-4651-A2B5-052F4542AB61

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R172360 collected from 80 km north of Broome, Western Australia, Australia, Latitude -17.460833 S., Longitude 122.2025 E.

This government-owned facility allows access to its holdings.

Paratypes: Three preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R168517, R168518 and R16851 all collected from Coulomb Point, Western Australia, Australia, Latitude -17.501389 S., Longitude 122.147222 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans, salt flats and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

The thirteen species in the *G. bipes* complex as identified herein are as follows:

G. bipes is a taxon of the Pilbara region, found only north of the Fortescue River in Western Australia in the region of Nickol Bay, the Burrup Peninsula and near the coast.

G. kunja is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin.

G. arrernte is herein restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

G. amazing sp. nov. is a coastal taxon restricted to a small area from around Broome to about 100 km north of Broome, in Western Australia.

G. wow sp. nov. is a taxon found from Port Hedland to around Sandfire Flat/Mandora in northwest Western Australia.

G. robwatsoni sp. nov. is known only from around the Derby area in northwestern Western Australia.

G. vellai sp. nov. is a species of the west coastal south Pilbara, south of the Fortescue River and including nearby dune country to the south.

G. questionthenarrative sp. nov. is a species of the eastern Great Sandy Desert in Western Australia.

G. narcissismawareness sp. nov. is a species with a centre of distribution being the Little Sandy Desert and nearby parts of central Western Australia.

G. truthisscience sp. nov. is a species from the Goldfields area and nearby parts of south-east Western Australia, well north of the coast.

G. liesarenotscience sp. nov. is confined to the central west of South Australia, north and east of the Nullarbor (but not near the coast) and south of the floodplain areas in north-west South Australia. The Lake Eyre system forms a hard barrier to the east.

G. anangu sp. nov. is restricted to central Australia south of the Amadeus basin and north of the flooding areas in far north South Australia.

G. owe sp. nov. is confined to the Simpson Desert in the south-east of the Northern Territory.

G. anangu sp. nov. and *G. owe sp. nov.* are the only two relevant species named in the absence of molecular data confirming or denying their status.

However, both occur in well-defined areas with established barriers on all sides and are among the most divergent of the relevant forms.

The thirteen preceding species are separated from one another by the following unique combinations of characters:

G. bipes is beige on top of the dorsum. Posterior to the rostral the dorsal head shields are marked with irregular dark brown blotches of irregular shape, in an asymmetrical pattern, which occupy about a quarter of the areas of the scales, with the percentage higher posteriorly and lower anteriorly. The two vertebral scale rows each have an elongated distinct dark brown spot on that cut across the suture lines of each scale (making the centre of each usually the break between the spots), in total forming a pair of paravertebral lines which extend along the entire length of the original tail.

About a quarter of the way down the tail's length it rapidly transforms in colour from beige to light grey.

From the nasal scale and backwards is a lateral stripe that runs along the top of the neck, the flank (where it becomes very broad) and ultimately the entire length of the upper side of the tail, this being same on each side of the lizard.

The colour along the vertebral line of the dorsum and tail is the same as that on either side of the paravertebral stripes. There is either no obvious bleeding of yellow or orange at the lower edge of the dark lateral stripe, or if so, it is very thin in width and slight, with the scales below this and on the venter being an immaculate white.

Upper labials anterior to the eye are brown. Those below the eye and posterior to it are white below and brown above. Along the lower margin of the original tail, midway along its length is a series of tiny purplish brown spots forming a line over the otherwise immaculate white of the lower sides of the tail and underneath. 1-2 nuchals, 2 supraoculars, 9-12 lamellae under the longer toe.

G. bipes is depicted in life online at:
https://www.inaturalist.org/observations/173727537
and
https://www.flickr.com/photos/julesfarquhar/52382226809/
and
https://www.flickr.com/photos/julesfarquhar/52382228074/

Gaia arrernte Hoser, 2024 is separated from *G. bipes* and others in the complex by having a dark yellow or yellow-brown dorsum versus beige in *G. bipes,* a particularly sharp and well-defined dorsal pattern, versus moderately well-defined in *G. bipes*, 3-4 nuchals, 3 supraoculars, 11- 15 lamellae under the longer toe, combined with a longer average length of limbs.

G. arrernte is depicted in life in Wilson and Swan (2021) on page 357 bottom and online at:
https://www.flickr.com/photos/127392361@N04/54010019533/

G. kunja Hoser, 2024 is similar in most respects to *G. arrernte* Hoser, 2024 as just described but is separated from that taxon by having 2 supraoculars (versus 3) and a dorsal pattern that is only moderately well-defined.

G. anangu sp. nov. is purplish orange on the top of the dorsum, being deepest in colour on the neck and lightest toward the tip of the snout, which is usually white. The paravertebral lines on the dorsum and tail, while broken, are thick and well defined and run almost the length of the tail.

Midway along the tail, the upper surface transitions from purplish orange to grey, but this is not the case between the paravertebral line on the top of the tail, where the colour remains purplish orange.

Upper surfaces of hind limbs are light orange with widely scattered smallish to medium-sized dark brown spots.

Anterior head shields are immaculate, but those from the eyes back have a small number of well-defined dark spots arranged in a symmetrical manner on the head.

Upper labials (both anterior and posterior) are white at the bottom and darker (orange, brown or both) on top.

G. anangu sp. nov. is depicted in life online at:
https://www.flickr.com/photos/58349528@N02/51817809803/
and
https://www.inaturalist.org/observations/273640548
and
https://www.flickr.com/photos/ryanfrancis/52578696391/

G. owe sp. nov. is similar in most respects to *G. anangu sp. nov*. as just described but separated from that taxon by being a light orange on top, with a reddish tinge, and the spots on the scales at the rear of the head are tiny. The outer edges of the upper surface of the rear of the tail are extremely light grey or white.

Upper labials (both anterior and posterior) are white at the bottom and orange on top.

G. truthisscience sp. nov. is a beige coloured lizard on the dorsum. The spots forming the paravertebral line are square in shape, rather than rectangular.

At the back of the top of the head, from the eye back are 8 medium-sized brown spots on the scales arranged to form a V-shaped arrangement.

Most of the tail is also beige and there is no difference in colour or intensity between the paravertebral lines on the body or on the tail.

Anterior labials are a dull orange colour, while posterior ones are an immaculate white.

There is a very slight bleeding of beige below the dark upper lateral line.

Within the dark lateral line, the centre of each otherwise dark brown scale is a dull orange colour, this central part being centred on the anterior edge of

each scale.

The white scales on the lower flank have dull pink etchings on the posterior edges.

Upper surfaces of the (hind) limbs are white or whitish, either immaculate or with scattered tiny slightly darker spots or bars.

G. truthisscience sp. nov. is depicted in life online at:

https://www.flickr.com/photos/zimny_anders/48711141222/

and

https://www.flickr.com/photos/zimny_anders/48711101492/

G. liesarenotscience sp. nov. is similar in most respects to *G. truthisscience sp. nov*. as detailed above but is separated from that taxon by having a generally more washed-out colouration including no obvious markings on otherwise light limbs. Below the dark upper lateral line is an obvious bleeding of yellow onto the white below.

All upper labials are immaculate white.

G. narcissismawareness sp. nov. is orange on the top of the head fading to become yellow on the rear dorsum. The tail is beige on top for the entire length. The paravertebral lines on the dorsum are reduced in size of spots and intensity of them and in some specimens these lines are absent. Upper surfaces of limbs are heavily speckled dark over a dark pink background, giving them an overall dark purple colour.

The dark upper lateral stripe on the flank is bold and sharp edged both top and bottom, generally straight in line (as in with few if any jagged edges on the top). There is no colour bleeding over white on the lower edge. Upper labials are immaculate white. Head shields are either immaculate or if with spots, these are tiny and irregularly spaced. There is no spotting of any sort on the white lower parts of the tail.

G. narcissismawareness sp. nov. is depicted in life online at:

https://www.flickr.com/photos/171250498@N08/54023557237/

and

https://www.flickr.com/photos/114192916@N07/54129318165/

G. questionthenarrative sp. nov. is white at the anterior snout on top, with the posterior head shields being an immaculate light orange colour, usually without any speckling or spots. They are also thinly etched dark. The back of the head, top of neck and dorsum are a dark deep reddish-brown colour, grading to dark yellowish brown at the anterior part of the tail and purplish at the distal end.

Paravertebral lines are either reduced in intensity so as to be barely noticeable or alternatively absent.

The dark upper lateral stripe disintegrates at the anterior part of the tail and the main part of the tail top and sides is a combination of dark, orange and pinkish speckling and spots, giving it a unicolour non-patterned appearance.

There is a thick orange bleed over the white below the dark upper lateral stripe. The bleed itself is bold and makes a thin orange line at the lower edge of the dark upper lateral stripe.

Upper surfaces of limbs are a dull yellow pink in colour.

Upper labials are all immaculate white.

G. amazing sp. nov. is a dark yellow-brown lizard on top.

The dorsal head shields are all immaculate beige. The paravertebral lines are reduced in intensity along most of the length of the body, or otherwise not present. In some specimens, at the anterior part, the scales within the paravertebral line are also darker, giving the appearance of a dull vertebral stripe.

The dark upper lateral stripe is a medium brown colour, rather than the dark brown seen in most other species in the complex. It is also relatively thin, usually only occupying the upper third of the flank.

the dark lower boundary is abrupt with the white below and there is no evidence of colour bleed here. Upper surfaces of limbs are heavily spotted a purplish colour on a whitish purple background.

All upper labials are immaculate white.

G. amazing sp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/117096144

G. robwatsoni sp. nov. is slightly brownish on the dorsum anteriorly, with most of the dorsum being beige and the tail rapidly turning light grey on top.

The upper lateral line breaks on the neck and on the body is a slight bleed of brownish from the lower edge of the upper lateral line onto the immaculate white below. Head shields lack spots or etching of any sort, except those at the back of the head which have about 4 symmetrically placed small dark brown spots.

Upper surfaces of the legs are a light pinkish colour, with scattered tiny, semi-distinct dark purple spots.

The upper lateral stripe continues in a bold manner to the end of the tail. Below this is immaculate white.

All upper labials are immaculate white.

G. wow sp. nov. is a variable taxon, but invariably with the following characters:

It is an intensely coloured lizard.

The head and neck is usually a deep, orange colour. The dark upper lateral band is of reduced size and intensity on the body (often starting some way down the body and not on the head) and what separates this species from all others in the complex is that this bad is most bold on most of the tail, rather than on the body (versus the reverse in all the other species in the complex).

Hoser 2026 - Australasian Journal of Herpetology 81-82:1-128.

Anterior upper labials are usually orange, while the posterior ones are usually white below and orange above.

The dorsum of the lizard is mainly a yellowish orange colour, while the upper surface of the tail abruptly changes to a light whitish grey colour along its entire length on top.

The upper part of each side of the tail has a bold black stripe, below being immaculate white and with no spotting or marks of any sort.

Upper surfaces of the limbs are mainly a dark purplish brown, being comprised of close spaced bars, spots or peppering, overlaying a pinkish purple background.

G. wow sp. nov. is depicted in life online at:

https://www.flickr.com/photos/114192916@N07/54079664425/

G. vellai sp. nov. is a generally light orange coloured lizard on top of the body with a mainly light grey tail.

As a rule, the dorsal head shields are immaculate orange and with no, or very few tiny black dots. The paravertebral lines formed by the dots on the scales are relatively indistinct, the individual spots being small and circular in shape.

From the snout, all the way along the body to the hind legs there is a significant bleed of orange below the lower edge of the dark upper lateral stripe. Unusual among species in this complex, is the fact that between the paravertebral lines of dots the scales colour is a dark orange, versus a light orange outside these lines of spots. This differential in colouration is most intense above the pelvic girdle.

These paravertebral lines remain for most of the length of the tail, but inside these spotted lines and outside the intensity of the grey of the tail and colour is the same.

The dark upper lateral line continues in prominent form along most of the length of the tail. Beneath this line there is dark spotting on the white lower parts of the tail.

Upper labials are mainly orange except for the white lower edges of the posterior ones.

G. vellai sp. nov. is depicted in life online at:

https://www.flickr.com/photos/euprepiosaur/54567679222/

and

https://www.flickr.com/photos/114192916@N07/54080275589/

The preceding species are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows

and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. amazing sp. nov*. is a coastal taxon restricted to a small area from around Broome to about 100 km north of Broome, in Western Australia.

Etymology: In 1983 I showed a specimen of one of these lizards to a Yawuru Aboriginal elder from the same area. I asked him their tribal name for it and he simply looked at me and said "*Amazing*".

GAIA ANANGU SP. NOV.

LSIDurn:lsid:zoobank.org:act:0FDDD17C-7AC2-4FB4-A0C9-FBCCDD7E41EA

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R68361 collected from Uluru, Northern Territory, Australia, Latitude -25.3547 S., Longitude 131.0425 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ Two preserved specimens at the South Australian Museum, Adelaide, South Australia, Australia, specimen numbers R68362 and R68363 both collected from Uluru, Northern Territory, Australia, Latitude -25.3547 S., Longitude 131.0425 E., 2/ Four preserved specimens at the Museum and Art Gallery of the Northern Territory Reptile Collection, Darwin, Northern Territory, Australia, specimen numbers R14238, R14239, R14240 and R14241 all collected from the southern boundary of the Uluru-Kata Tjuta National Park, south of Uluru (AKA Ayers Rock), Northern Territory, Australia, Latitude -25.35 S., Longitude 131.017 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. anangu sp. nov. is restricted to central Australia south of the Amadeus basin and north of the flooding areas in far north South Australia.

G. anangu sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. anangu sp. nov. is purplish orange on the top of the dorsum, being deepest in colour on the neck and lightest toward the tip of the snout, which is usually white. The paravertebral lines on the dorsum and tail, while broken, are thick and well defined and run almost the length of the tail.

Midway along the tail, the upper surface transitions from purplish orange to grey, but this is not the case between the paravertebral line on the top of the tail, where the colour remains purplish orange.

Upper surfaces of hind limbs are light orange with widely scattered smallish to medium-sized dark brown spots.

Anterior head shields are immaculate, but those from the eyes back have a small number of well-defined dark spots arranged in a symmetrical manner on the head.

Upper labials (both anterior and posterior) are white at the bottom and darker (orange, brown or both) on top.

G. anangu sp. nov. is depicted in life online at: https://www.flickr.com/photos/58349528@N02/51817809803/

and

https://www.inaturalist.org/observations/273640548

and

https://www.flickr.com/photos/ryanfrancis/52578696391/

The 13 species within the *G. bipes* complex are

separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. anangu sp. nov*. is restricted to central Australia south of the Amadeus basin and north of the flat and flooding areas in far north South Australia.

Etymology: *G. anangu sp. nov*. is named in recognition of the Anangu Aboriginal people native to the area that this taxon occurs. There is no truth in the rumour that Aboriginals there wanted to contract my eldest daughter Adelyn Hoser for their “Welcome to Country” smoking ceremonies because she farted so much and could better ignite the flames.

GAIA AVIYEMENII SP. NOV.

LSIDurn:lsid:zoobank.org:act:84479BC7-6A70-4426-9074-DAD7F7402FC7

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, specimen number R102785 collected from the Little Sandy Desert, Western Australia, Australia, Latitude -24.531944 S., Longitude 120.291111 E.

This government-owned facility allows access to its holdings.

Paratypes: Five preserved specimens at the Western Australian Museum, Perth, Western Australia, specimen numbers R102625, R102700, R137557, R137558 and R137592 all collected a short distance north of the type locality and well within the Little Sandy Desert, Western Australia, Australia.

Diagnosis: Until now, each of *Gaia dezifreemani sp. nov.* and *Gaia aviyemenii sp. nov*. have been treated as outlier populations of *Gaia ips* (Storr, 1980) originally placed in the greater genus *Lerista* (Bell, 1833), with a type species of *Lerista lineata* Bell, 1833 by monotypy.

The type locality of *Gaia ips* is Lake Auld, Western Australia, Australia, Latitude 22.07 S., Longitude 123.52 E., which is within the Great Sandy Desert bioregion, being where that species as recognised herein occurs.

Where *Gaia dezifreemani sp. nov.* occurs to the east in far east Western Australia and the adjacent southwest parts of the Northern Territory is in the western part of the Tanami region and separated by a zone of unsuitable habitat dividing the two taxa.

Gaia aviyemenii sp. nov. is a species of the Little Sandy Desert, separated by hills extending from the Pilbara being a barrier separating the Little Sandy Desert from the Great Sandy Desert to the north and northeast of the Pilbara.

The three populations of each species are physically isolated from one another, genetically divergent from one another, as shown in Singhal *et al.* (2025) (see also their supplementary data) and so morphologically divergent from one another that it is entirely appropriate that they are recognised as three full species rather than as one highly variable one.

The three species are most readily separated from one another by dorsal colouration in adults.

They are separated from one another by the following unique colour combinations:

Gaia ips (Storr, 1980) is essentially a light reddish or pinkish coloured animal (accurately described by Storr (1980) as being earthworm pink) all over the dorsum and upper flank. There is barely any hint of a mid-vertebral line running along the dorsum.

The (original) tail is essentially the same colour as the dorsum. Snout tip is lighter than the rest of the dorsum, even being white in some specimens. Hindlegs are pink in colour.

Storr (1980) publishes a photo of the holotype of *Gaia ips* in life on page 442. The same species is depicted in Wilson and Knowles (1988) on page 299 at top right from the Sandfire area, Western Australia.

Gaia dezifreemani sp. nov. by contrast is a mainly silvery-grey coloured lizard, or silvery grey with a pinkish hue. Most of the top of the head is white or whitish. In contrast to both *Gaia ips* and *Gaia aviyemenii sp. nov*. the demarcation line from darker dorsum to white venter is not particularly well defined, versus well-defined in the other two species. Hindlegs are also silvery-grey like the dorsum.

Gaia dezifreemani sp. nov. is depicted in life online at:
https://www.flickr.com/photos/171250498@N08/53325254467/
and
https://www.inaturalist.org/observations/254059106

Gaia aviyemenii sp. nov. is a mainly yellowish colour along the vertebral line, which is in effect a wide vertebral stripe commencing on the neck and running onto the tail, which then turns white for most of its length. Head, lateral edges and upper flanks are also orange in colour. Hindlegs are mainly white.

It is depicted in life in Wilson and Swan (2021) on page 369 at second from top.

The three preceding species, (*Gaia ips*, *G. dezifreemani sp. nov.* and *G. aviyemenii sp. nov*.) are separated from all other species in the genus *Gaia* Wells, 2012, type species *Rhodona bipes* Fischer, 1882 by having a stout body, broader head, no dark upper lateral stripe and a single loreal.

Species within the genus *Gaia* Wells, 2012 are separated from all other species within *Lerista sensu lato* as defined by Hoser (2024c), by the following unique combination of characters:

Snout depressed, very sharp in profile and extending well beyond the mouth.

Eye very small. Eyelid movable. No trace of foreleg (including groove). Two toes. Nasals separated moderately widely. No prefrontals. Frontal a little wider than long. Frontoparietals and interparietal fused into a large triangular shield. Parietals long and narrow, in short contact. Nuchals 1 or 2. Supraoculars 2, both in contact with frontal. Supraciliaries 0+1. Preoculars 1. Postoculars 3. Temporals 3, Upper labials usually 6, Ear aperture minute.

It should be mentioned that the description for *Gaia ips* in Hoser (2024c) on page 19, second column is incorrect as it was based on the taxon regarded herein as *G. aviyemenii sp. nov*. and not *Gaia ips.*

The relevant descriptions herein are the correct ones.

Unlike narcissists, I will own and correct my mistakes when I become aware of them.

For the record, all corrections in this paper as compared to Hoser (2024c) came from my own investigations and not those of any third party. However, if they had come from a third party source, the errors would have been corrected all the same.

Distribution: *Gaia aviyemenii sp. nov.* is presently only known from the Little Sandy Desert area south of the Pilbara District in Western Australia, Australia. It is unlikely to extend out of this area.

Etymology: Named in honour of Avi Yemeni, in recognition of his services to journalism.

Avraham Shalom Yemini is an Australian-Israeli political commentator who regularly exposes corruption in government and the legacy media. Since 2020 he has worked as the Australian correspondent for Rebel News, a Canadian based website. Yemini has been repeatedly sued in the corrupt Australian courts by those who seek to silence him.

The State controlled media in Australia (ABC, News Corporation, Nine Media) brand him as a far-right lunatic as a means to discredit him. The narrative is false.

GAIA DEZIFREEMANI SP. NOV.

LSIDurn:lsid:zoobank.org:act:4426009E-547C-466D-8106-3907992B4F6D

Holotype: A preserved adult male specimen at the Western Australian Museum, Perth, Western Australia, specimen number R133216 collected from Lake Mackay, Western Australia, Australia, Latitude -22.444167 S., Longitude 128.290833 E.

This government-owned facility allows access to its holdings.

Paratypes: Four preserved adult male specimens at the Western Australian Museum, Perth, Western Australia, specimen numbers R133272, R133371, R133585, R133603 and R133604 all collected from Lake Mackay, Western Australia, Australia, Latitude -22.444167 S., Longitude 128.290833 E.

Diagnosis: Until now, each of *Gaia dezifreemani sp. nov.* and *Gaia aviyemenii sp. nov*. have been treated as outlier populations of *Gaia ips* (Storr, 1980) originally placed in the greater genus *Lerista* (Bell, 1833), with a type species of *Lerista lineata* Bell, 1833 by monotypy.

The type locality of *Gaia ips* is Lake Auld, Western Australia, Australia, Latitude 22.07 S., Longitude 123.52 E., which is within the Great Sandy Desert bioregion, being where that species as recognised herein occurs.

Where *Gaia dezifreemani sp. nov.* occurs to the east in far east Western Australia and the adjacent southwest parts of the Northern Territory is in the western part of the Tanami region and separated by a zone of unsuitable habitat dividing the two taxa.

Gaia aviyemenii sp. nov. is a species of the Little Sandy Desert, separated by hills extending from the Pilbara from the Great Sandy Desert to the north and northeast of the Pilbara.

The three populations of each species are physically isolated from one another, genetically divergent from one another, as shown in Singhal *et al.* (2025) (see also their supplementary data) and so morphologically divergent from one another that it is entirely appropriate that they are recognised as three full species rather than as one highly variable one.

The three species are most readily separated from one another by dorsal colouration in adults.

They are separated from one another by the following unique colour combinations:

Gaia ips (Storr, 1980) is essentially a light reddish or pinkish coloured animal (accurately described by Storr (1980) as being earthworm pink) all over the dorsum and upper flank. There is barely any hint of a mid-vertebral line running along the dorsum.

The (original) tail is essentially the same colour as the dorsum. Snout tip is lighter than the rest of the dorsum, even being white in some specimens. Hindlegs are pink in colour.

Storr (1980) publishes a photo of the holotype of *Gaia ips* in life on page 442. The same species is depicted in Wilson and Knowles (1988) on page 299 at top right from the Sandfire area, Western Australia.

Gaia dezifreemani sp. nov. by contrast is a mainly silvery-grey coloured lizard or silvery grey with a pinkish hue. Most of the top of the head is white or whitish. In contrast to both *Gaia ips* and *Gaia aviyemenii sp. nov*. the demarcation line from darker dorsum to white venter is not particularly well defined, versus well-defined in the other two species. Hindlegs are also silvery-grey like the dorsum.

Gaia dezifreemani sp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/254059106

Gaia aviyemenii sp. nov. is a mainly yellowish colour along the vertebral line, which is in effect a wide vertebral stripe commencing on the neck and running onto the tail, which then turns white for most of its length. Head, lateral edges and upper flanks are also orange in colour. Hindlegs are mainly white.

It is depicted in life in Wilson and Swan (2021) on page 369 at second from top.

The three preceding species, (*Gaia ips*, *G. dezifreemani sp. nov.* and *G. aviyemenii sp. nov*.) are separated from all other species in the genus *Gaia* Wells, 2012, type species *Rhodona bipes* Fischer, 1882 by having a stout body, broader head, no dark upper lateral stripe and a single loreal.

Species within the genus *Gaia* Wells, 2012 are separated from all other species within *Lerista sensu lato* as defined by Hoser (2024c), by the following unique combination of characters:

Snout depressed, very sharp in profile and extending well beyond the mouth.

Eye very small. Eyelid movable. No trace of foreleg (including groove). Two toes. Nasals separated moderately widely. No prefrontals. Frontal a little wider than long. Frontoparietals and interparietal fused into a large triangular shield. Parietals long and narrow, in short contact. Nuchals 1 or 2. Supraoculars 2, both in contact with frontal. Supraciliaries 0+1. Preoculars 1. Postoculars 3. Temporals 3, Upper labials usually 6, Ear aperture minute.

It should be mentioned that the description for *Gaia ips* in Hoser (2024c) on page 19, second column is incorrect as it was based on the taxon regarded herein as *G. aviyemenii sp. nov*. and not *Gaia ips.*

The relevant descriptions herein are the correct ones.

It is important that mistakes identified in papers be made known and corrected expeditiously.

Distribution: *Gaia dezifreemani sp. nov.* is presently only known from the Tanami Desert region of the Northern Territory and immediately adjacent parts of northern Western Australia, Australia. It is unlikely to be found beyond this area.

Etymology: Named in honour of Dezi Freeman for services to the Australian public. For several years he tried tirelessly to expose corruption in the Victorian Police and the cocaine addicted judiciary in the state.

Rather than dealing with the serious corruption

problem, it was organised that the Victorian police shot him execution-style on 30 March 2026, at a rural property in Thologolong, Victoria (near Walwa, close to the New South Wales border), being in a remote part of north-east Victoria.

In recent legal proceedings he had exposed a corrupt Victorian magistrate for serious crimes in a case that garnered wide publicity.

Cocaine addiction, bribe taking and general corruption are endemic in the Victorian judiciary (Fraser, 2011).

GAIA LIESARENOTSCIENCE SP. NOV.

LSIDurn:lsid:zoobank.org:act:706BBADB-37C3-4065-AB36-EB3DF45A530B

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R57936 collected from 47.1 km east southeast of Emu, South Australia, Australia, Latitude -28.8644 S., Longitude 132.6058 E.

This government-owned facility allows access to its holdings.

Paratypes: Three preserved specimens at the South Australian Museum, Adelaide, South Australia, Australia, specimen numbers R47468, R47469 and R47474 all collected from 5.9 km south of the Oolarinna East Bore, South Australia, Australia, Latitude -27.6717 S., Longitude 132.9164 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans, salt lakes and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. truthisscience sp. nov. is a species from the Goldfields area and nearby parts of south-east Western Australia, well north of the coast.

G. liesarenotscience sp. nov. is confined to the central west of South Australia, north and east of the Nullarbor (but not near the coast) and south of the floodplain areas in north-west South Australia. The Lake Eyre system forms a hard barrier to the east.

G. truthisscience sp. nov. and *G. liesarenotscience sp. nov*. is separated from the other 11 species in the *G. bipes* complex by the following unique combinations of characters:

G. truthisscience sp. nov. is a beige coloured lizard on the dorsum. The spots forming the paravertebral line are square in shape, rather than rectangular.

At the back of the top of the head, from the eye back are 8 medium-sized brown spots on the scales arranged to form a V-shaped arrangement.

Most of the tail is also beige and there is no difference in colour or intensity between the paravertebral lines on the body or on the tail.

Anterior labials are a dull orange colour, while posterior ones are an immaculate white.

There is a very slight bleeding of beige below the dark upper lateral line.

Within the dark lateral line, the centre of each otherwise dark brown scale is a dull orange colour, this central part being centred on the anterior edge of each scale.

The white scales on the lower flank have dull pink etchings on the posterior edges.

Upper surfaces of the (hind) limbs are white or whitish, either immaculate or with scattered tiny slightly darker spots or bars.

G. truthisscience sp. nov. is depicted in life online at:

https://www.flickr.com/photos/zimny_anders/48711141222/

and

https://www.flickr.com/photos/zimny_anders/48711101492/

G. liesarenotscience sp. nov. is similar in most respects to *G. truthisscience sp. nov*. as detailed

above but is separated from that taxon by having a generally more washed-out colouration including no obvious markings on otherwise light limbs. Below the dark upper lateral line is an obvious bleeding of yellow onto the white below.

All upper labials are immaculate white.

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. liesarenotscience sp. nov*. is confined to the central west of South Australia, north and east of the Nullarbor (but not near the coast) and south of the floodplain areas in north-west South Australia. The Lake Eyre system forms a hard barrier to the east.

Etymology: Lies are not a part of science, or at least they should not be. It is that simple.

GAIA OTIT SP. NOV.

LSIDurn:lsid:zoobank.org:act:7FA45616-2599-481E-A782-E37469E5CDBF

Holotype: A preserved female specimen at the Western Australian Museum, Perth, Western Australia, Australia specimen number R171610 collected from Storr Island, West Kimberley District, Western Australia, Australia, Latitude -15.949722 S., Longitude 124.561111.

This government-owned facility allows access to its holdings.

Paratype: A preserved male specimen at the Western Australian Museum, Perth, Western Australia, Australia specimen number R171613 collected from Storr Island, West Kimberley District, Western Australia, Australia, Latitude -15.949722 S., Longitude 124.561111.

Diagnosis: Hoser (2024c) split the species *Gaia greeri* (Storr, 1982) as recognized then (being that of Storr, 1982) into two species based on very significant and obvious divergences, distributional disjuncture and known antiquated biogeographical boundaries.

Singhal *et al.* (2025) in their supplementary Fig S5 showed that Storr's concept of *G. greeri* included four putative species implying that Hoser (2024c) had not gone far enough.

This view was clarified somewhat by Torkkola *et al.*

(2026) at Fig 1., where they showed that the Western type form was in fact split into two closely related species (one north form from Storr Island, West Kimberley and the south Type form of *G. greeri*, from 8 km south southeast of Derby, Western Australia, Latitude 17.22 S., Longitude 123.40 E.).

Additionally, Torkkola *et al.* (2026) found that an eastern sample, being West Australian Museum specimen number WAM R137387 collected from Fowlhouse Junction, Osmond Range, East Kimberley district of Western Australia, Latitude -17.266667 S., Longitude 128.483333 E. was a very, very divergent taxon.

This essentially was in line with the concept of Hoser (2024c) for *G. oomph* Hoser, 2024, supporting the erection of that species in the strongest possible terms.

It should be noted that the type form of *G. oomph* from the Tanami Desert, while similar in most respects to other East Kimberley animals previously referred to *G. greeri,* did differ from some.

Were it not for my inherently conservative nature, I would have recognised at least one further species or subspecies on the basis of the differences. However, myself being ever (over) sensitive to ongoing unsupported claims by detractors that I have over split reptile species, I refrained from doing so.

In hindsight, taking notice of unscientific keyboard warriors, working as flying monkeys for the Wüster / Britton / Kaiser gang, generally from fake accounts, was a serious mistake on my part.

To confirm my mistake in listening to trolls, Singhal *et al.* (2025) in their supplementary Fig S5 also indicated that there were in fact two species occupying the same general region, this being the inland south and east Kimberley districts.

In other words, only two of the relevant four species had been formally named until now, although this did include the two main lineages.

Rather than having the less divergent, but still unnamed species-divergent lineages expire through general indifference I formally name them both herein. This is so that proper conservation measures can be implemented by governments, non-governmental organizations and concerned individuals.

The south and southeast Kimberley form is herein named *Gaia rate sp. nov.*, while the Storr Island form is herein named *Gaia otit sp. nov*..

This means that the diagnoses and concepts for *Gaia greeri* and *Gaia oomph* as given in Hoser (2024c) must be amended slightly to accommodate the two other species formally named within this species complex.

Gaia oomph and the newly named and most closely related species *Gaia rate sp. nov.* are morphologically and genetically divergent from the type form of *G. greeri*, the closely related *Gaia otit sp. nov*. and the closely related taxa, *G. robusta* (Storr, 1990), *G. vermicularis* (Storr, 1982), *G. nomenclaturematters sp. nov.* (this paper), *G. ruleofpriority sp. nov*. (this paper), *G. taxonomymatters sp. nov*. (this paper), *G. labialis* Storr (1971) or *G. simillima* (Storr, 1984). See for example the relevant sequence data published in Skinner *et al.* (2008), Farquhar *et al*. (2024), Singhal *et al.* (2025) or Torkkola *et al.* (2026).

Gaia oomph Hoser, 2024 and *Gaia rate sp. nov.* are readily separated from the previously named species as follows:

G. greeri, *G. otit sp. nov*., *G. labialis* and *G. simillima* are lizards with a bold dark lateral band that is either black, very dark brown and with a sharp upper and lower edge.

G. vermicularis, G. nomenclaturematters sp. nov. (this paper), *G. ruleofpriority sp. nov*. (this paper) and *G. taxonomymatters sp. nov*. are lizards with an orange red dorsal surface, sometimes pinkish, yellowish or orange instead, including to the snout and with white on the mid and lower flanks (below an apper lateral line that may or may not be well-defined). The white of the under surface extends about half-way up the side of the flank.

G. robusta is the most similar species to *G. oomph sp. nov.* and *G. rate sp. nov.*. The dorsal colour is orange on top. There is brown stripe on the upper lateral surface, reasonably sharply defined top and bottom and this occupies most of the side of the flank. The white from below only encroaches the lower edge of the flank. The distal end of the tail is grey.

A more distantly related species within *Gaia* Wells, 2012 is *G. ips* (Storr, 1980), that is separated from the other species by having a moderately to well-defined and wide mid-vertebral line that is yellow in colour on a dorsum that is otherwise orange, with a moderately well-defined orange upper lateral line extending about halfway down the side of the flank. Most of the tail is ghost white.

G. oomph and *G. rate sp. nov.* in contrast to all the preceding species is separated from them and defined as follows:

It has a dorsum that is a light beige-brown colour, extending along the tail.

The snout and back of head is whitish, darkening on the neck, lightening on the rear body and then intensifying on the anterior tail before becoming near white at the distal end.

The dorsal colouration extends over the dorsolateral edge where a boundary of darker brown occurs.

This is formed by one or two scale rows having the main part of each scale a darker brown colour.

Below this line that is poorly defined above and well-defined below is the white of the majority of the flank

that extends to the white belly.

On the anterior half of the tail, the dark band of the upper lateral edge, remains 2 scales wide and becomes well defined on top as well as below, with sharper contrast in colour, giving the anterior tail a well-defined striped appearance. These colours reduce to become white or white with flecks at the distal end.

G. oomph and *G. rate sp. nov.* are further readily separated from *G. greeri* and *G. otit sp. nov*. by the fact that along the paravertebral lines spotting is small and distinct but in no way forming obvious lines in *G. oomph* or very obscured in the form of feint small dull spots or none at all in *G. otit sp. nov*., versus bold, black or blackish, enlarged or merged to form obvious paravertebral lines on the dorsum in both *G. greeri* and *G. otit sp. nov*.

G. rate sp. nov. is separated from *G. oomph* by not having a pink tinge on the dorsum (versus having one), yellow within the anterior part of the dark upper lateral band (versus pinkish orange) and by the fact that along the paravertebral lines spotting is small and distinct but in no way forming obvious lines in *G. oomph* versus very obscured in the form of feint small dull spots or none at all in *G. otit sp. nov*.,

G. greeri and the closely related *G. otit sp. nov*. are separated by the fact that the dorsum of *G. greeri* is dark yellowish or light-brown or even light greyish in colour, versus light greyish with a slight reddish or pinkish tinge in *G. otit sp. nov*. and upper surfaces of hind limbs of *G. greeri* that are mainly dark in colour, versus mainly light in *G. otit sp. nov*. (as well as *G. oomph* and *G. rate sp. nov.*).

G. oomph, *G. rate sp. nov.*, *G. greeri* and *G. otit sp. nov*. are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

No trace of a forelimb or forelimb groove; two toes; no prefrontals; nasals usually separated; 6 upper labials, 20 midbody rows, 2 supraoculars in contact with the frontal and 1-2 supraciliaries.

A detailed description of the type form of *G. greeri* is in Wells (2012) on pages 198-199 which is evidently largely applicable to *G. oomph* Hoser, 2024, *G. rate sp. nov.* and *G. otit sp. nov*. as well.

G. oomph Hoser, 2024 is depicted in life online at:

https://www.flickr.com/photos/58349528@N02/54678291202/

G. rate sp. nov. is depicted in life online at:

https://www.flickr.com/photos/euprepiosaur/53840611459/

and

https://www.flickr.com/photos/171250498@N08/53835283097/

G. greeri (of the nominate type form) is depicted in life in Wilson and Swan (2021) on page 367 at top and online at:

https://www.flickr.com/photos/brian_busho/14270539460/

Distribution: *G. greeri* is mainly known from near the type locality of Derby, Western Australia and *G. otit sp. nov*. is known principally from Storr Island further north. Both areas are near the mainland coast in the west Kimberley District, with Storr Island only being separated from the mainland since the most recent glacial minimum.

Of relevance *G. rate sp. nov.* is a taxon from the southern and southeast Kimberley District, well away from the coast in the driest parts of the region.

G. oomph Hoser, 2024 is a taxon presently only known from the north-west Tanami in far east Western Australia and also the Ord River region of the east Kimberley District, also in Western Australia, from there extending into the nearby part of the Northern Territory.

Etymology: *G. otit sp. nov.* is a play on words.

Combined in English the words “Got it” is what one says when one catches the lizard.

GAIA OWE SP. NOV.

LSIDurn:lsid:zoobank.org:act:A3447C02-6EE2-406A-8E40-BC96A5BD07E3

Holotype: A preserved specimen at the Museum and Art Gallery of the Northern Territory, Darwin, Northern Territory, Australia, specimen number R33392 collected from Hay River, Simpson Desert, Northern Territory, Australia, Latitude -23.917 S., Longitude 137.267 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ A preserved specimen at the Museum and Art Gallery of the Northern Territory, Darwin, Northern Territory, Australia, specimen number R33450 collected from Hay River, Simpson Desert, Northern Territory, Australia, Latitude -23.917 S., Longitude 137.267 E., 2/ A preserved specimen at the Queensland Museum, Brisbane, Queensland, Australia, specimen number J89536 collected from the Simpson Desert National Park, far west Queensland, Australia, Latitude -24.101667 S., Longitude 138.205833 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative “*Lerista bipes*” in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. anangu sp. nov. is restricted to central Australia south of the Amadeus basin and north of the flooding areas in far north South Australia.

G. owe sp. nov. is confined to the Simpson Desert in the south-east of the Northern Territory.

G. anangu sp. nov. and *G. owe sp. nov.* are separated from the other 11 species in the *G. bipes* complex by the following unique combination of characters:

G. anangu sp. nov. is purplish orange on the top of the dorsum, being deepest in colour on the neck and lightest toward the tip of the snout, which is usually white. The paravertebral lines on the dorsum and tail, while broken, are thick and well defined and run almost the length of the tail.

Midway along the tail, the upper surface transitions from purplish orange to grey, but this is not the case between the paravertebral line on the top of the tail, where the colour remains purplish orange.

Upper surfaces of hind limbs are light orange with widely scattered smallish to medium-sized dark brown spots.

Anterior head shields are immaculate, but those from the eyes back have a small number of well-defined dark spots arranged in a symmetrical manner on the head.

Upper labials (both anterior and posterior) are white at the bottom and darker (orange, brown or both) on top.

G. anangu sp. nov. is depicted in life online at: https://www.flickr.com/photos/58349528@N02/51817809803/

and

https://www.inaturalist.org/observations/273640548

and

https://www.flickr.com/photos/ryanfrancis/52578696391/

G. owe sp. nov. is similar in most respects to *G. anangu sp. nov*. as just described but separated from that taxon by being a light orange on top, with a reddish tinge, and the spots on the scales at the rear of the head are tiny. The outer edges of the upper surface of the rear of the tail are extremely light grey or white.

Upper labials (both anterior and posterior) are white at the bottom and orange on top.

The 13 species within the *G. bipes* complex are

separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. owe sp. nov.* is confined to the Simpson Desert in the south-east of the Northern Territory.

Etymology: *G. owe sp. nov.* is a simple and easy to remember name. As an Australian herpetologist, you simply "owe" it to yourself to try to see a live specimen in the flesh.

GAIA NARCISSISMAWARENESS SP. NOV.

LSIDurn:lsid:zoobank.org:act:79FBB13A-E927-42A0-AE22-1A62F0E50470

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R129955 collected from near Mount Joel, Western Australia, Australia, Latitude -27.216667 S., Longitude 121.05 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R129985 collected from near Mount Joel, Western Australia, Australia, Latitude -27.216667 S., Longitude 121.05 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas

surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. narcissismawareness sp. nov. is a species with a centre of distribution being the Little Sandy Desert and nearby parts of central Western Australia.

G. narcissismawareness sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. narcissismawareness sp. nov. is orange on the top of the head fading to become yellow on the rear dorsum. The tail is beige on top for the entire length. The paravertebral lines on the dorsum are reduced in size of spots and intensity of them and in some specimens these lines are absent. Upper surfaces of limbs are heavily speckled dark over a dark pink background, giving them an overall dark purple colour.

The dark upper lateral stripe on the flank is bold and sharp edged both top and bottom, generally straight in line (as in with few if any jagged edges on the top). There is no colour bleeding over white on the lower edge. Upper labials are immaculate white. Head shields are either immaculate or if with spots, these are tiny and irregularly spaced. There is no spotting of any sort on the white lower parts of the tail.

G. narcissismawareness sp. nov. is depicted in life online at:

https://www.flickr.com/photos/171250498@N08/54023557237/

and

https://www.flickr.com/photos/114192916@N07/54129318165/

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end

of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. narcissismawareness sp. nov*. is a species with a centre of distribution being the Little Sandy Desert and nearby parts of central Western Australia.

Etymology: Narcissism is a well-known personality trait, but in terms of the pathological disorder it remains relatively little known to most people.

Persons with Narcissism Personality Disorder (NPD) are effectively created, at least at the precursor stage within the first seven years of life (probably within the first five years for most people).

In adults with the mature form of the disorder, they have a sense of entitlement and self-importance that manifests in zero concern for the welfare of others and a desire to get what they want at any means because they must be seen as superior to others. This sense of self-importance (being better than others) and zero concern for the welfare of others are at the core of the disorder and most of the traits such as development of the habit of pathologically lying and manipulating others are products of these core traits.

The lying, manipulation and controlling of others is not necessarily done overtly. In fact, most narcissistic behaviour is covert, manipulative, often with the victims unaware of what is happening and all done while the perpetrator maintains a false image of niceness and honesty.

At their core narcissists are fundamentally dishonest and literally lie all the time, including when they do not need to, or even when they'd be better off telling the truth.

To explain why this habit develops is beyond the scope of this etymology, but it is real and common to most if not all people with NPD.

They lie overtly, covertly and by omission and to a degree that most non narcissists would be unable to comprehend unless they have actually studied the pathology and taken time to observe the behaviour closely.

However, once the pathology becomes known and fully understood, narcissistic individuals and their dishonest behaviour is more easily seen and even becomes quite predictable.

Understanding of the pathology of Narcissistic Personality Disorder (NPD), that afflicts roughly 1 in 5 persons in the modern (present day) era is of great benefit to scientists and taxonomists in particular.

In this field, NPD manifests as so-called scientists who fake findings, plagiarise the works of others and who without any moral compass engage in acts of taxonomic vandalism.

That is perhaps best described here as stealing the work of others, renaming taxa that already have names and then pretending that the earlier works and names never existed.

The most dishonest and narcissistic part of the act of taxonomic vandalism comes when these pseudoscientists peddle fake news stories to media outlets about how through their own scientific prowess they have discovered and named new species.

There are countless examples of this.

See for example:

https://news.mongabay.com/short-article/2025/06/four-new-snake-species-discovered-in-papua-new-guinea/

and

https://www.theguardian.com/australia-news/2022/jun/27/frogs-that-lay-eggs-on-land-new-genus-named-after-wa-teacher-whose-lab-was-a-campervan

both being cases of where NPD pseudoscientists (Grant Webster and Fred Kraus) have faked discoveries made by another person, but lied by omission to the media in pretending that they actually discovered the species.

This is the narcissistic trait of seeking of grandiosity and admiration by any means possible, with lying merely being an acceptable part of the pathway there.

Narcissism doesn't just show up in the area of science and taxonomy. These same individuals as a rule engage in pathological NPD behaviour in other areas of their lives, including manipulating and controlling others, lying, blame shifting, cheating in marriages and so on.

Examples of persons engaging in taxonomic vandalism, abusing members of the opposite sex, including by cheating in marriage, pathologically lying in herpetological publications and in their own self-promotion are common and I have detailed examples in Hoser (2015a-f).

Individuals operating in herpetology who display the pathological traits of NPD include Wolfgang Wüster, Adam Britton, Wulf Schleip, Grant Webster, Bryan Fry, Fred Kraus and Hinrich Kaiser.

This means that none of their so-called scientific publications should ever be accepted as factual in the absence of proper scrutiny of every minute detail (and when this has been done, they almost always invariably fail).

In the case of Adam Britton, his egregious actions in terms of herpetological taxonomy as a key member of the Wüster gang and his publicly documented actions of stealing people's pet dogs for anal sex,

while hiding this behaviour from his wife for more than 20 years are not separate and divorced actions (Mackay 2024).
They are both parts of the same Narcissistic Personality Disorder.
See also:

https://www.bbc.com/news/articles/c33nv8x64mlo

In case I missed it, I should mention that another hallmark of this disorder is a steadfast refusal of narcissists to admit wrong or liability for their actions, or to apologise for them even when caught and presented with irrefutable proof. In terms of the Wüster/Britton/Kaiser gang and their members, this is seen when these individuals are exposed for wrongdoings, be it taxonomic vandalism, lying to the tabloid media about discoveries they never made, running fake accounts to manipulate opinion online, stealing scientific works from others, or when raping women, children or pet dogs.
The damage caused by persons with NPD to the sciences is so huge that a greater awareness of this is required.

Therefore, I have no hesitation in using a relatively small and obscure lizard species as a vehicle to bring attention to this serious problem.

GAIA NOMENCLATUREMATTERS SP. NOV.

LSIDurn:lsid:zoobank.org:act:E6431EE1-F0F4-42AF-BAD8-7F91D563B86F

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R133803 collected from in soil under leaf litter on a dune at Lake Mackay, Western Australia, Australia, Latitude -22.444167 S., Longitude 128.290833 E.

This government-owned facility allows access to its holdings.

Paratypes: Three preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R130927, R130938 and R134424 all collected from Lake Mackay, Western Australia, Australia, Latitude -22.444167 S., Longitude 128.290833 E.

Diagnosis: As originally described and conceived "*Lerista vermicularis* Storr, 1982", herein transferred to the genus *Gaia* Wells, 2012 in line with Hoser (2024c) was a putative species originally only known from the Great Sandy Desert, in north-west Western Australia (see Storr, 1982 including "Figure 3").

More recently specimens that are morphologically similar from the northern extension of the Great Sandy Desert, Tanami Desert Region and the Gibson Desert Region have been collected, lodged in the Western Australian Museum and assigned to the same species.

The same applies to colour images of these lizards posted on photo sharing sites such as "Inaturalist" and "Flickr".

However, the lizards themselves (as seen in the images posted) are quite different in form to one another. They are separated by barriers of unsuitable non-dune habitat of significant antiquity, and the molecular results of Torkkola *et al.* (2026) have confirmed the existence of three very different forms.

The northern-most form was not included in the study of Torkkola *et al.* (2026) but is morphologically divergent from the others and also separated by areas of unsuitable (non-dune) habitat, implying lengthy isolation.

Therefore, the three hitherto unnamed forms are formally named herein for the first time.

Gaia nomenclaturematters sp. nov. is the form previously assigned to *G. vermicularis* from the Tanami Desert and North Gibson Desert in east Western Australia.

Gaia taxonomymatters sp. nov. is the form previously assigned to *G. vermicularis* from the middle Gibson Desert in east Western Australia.

Gaia ruleofpriority sp. nov. occurs in a confined area east of Baines Hills and Elsey Hills in the area between the Great Sandy Desert (to the south-west), southeast Kimberley District (to the north), the open grass plains to the east and the Gardiner Range to the southeast

The four species are separated from one another by the following combinations of characters:

G. vermicularis Storr, 1982 is orangish-red on the dorsum of the body. The upper lateral band commences on the neck and extends to the hind limb. It is poorly defined and medium brown in colour (being of similar intensity and colour to the dorsum, making it indistinct), being poorly defined on the upper edge and better defined on the lower edge. Below this the scales are either immaculate white or very faintly and thinly etched.

The (original) tail is whitish with a series of well-defined black spots, arrange in rows, sometimes nearly or actually merging to form longitudinal lines.

Upper labials are immaculate white at the lower edge and dark above.

Gaia nomenclaturematters sp. nov. is pinkish yellow on the dorsum, with a well-defined dark brown stripe on the upper flank (being very distinct and separate from the colour on the dorsum and below the stripe). This stripe commences at the back of the head and extends to the hind limb. It is well defined on both upper and lower edge.

The (original) tail is whitish with a series of well-defined black spots, arrange in rows, sometimes nearly or actually merging to form longitudinal lines.

Upper labials are orange, becoming whitish posterior to the eye.

Gaia taxonomymatters sp. nov. is a light greyish yellow on the dorsum, with a dark upper stripe along

the upper flank commencing from the back of the head and running to the tail. It is reasonably week defined at the anterior end and bold on the latter part of the body. The lower edge bleeds yellowish orange over the otherwise immaculate white scales below. This lateral stripe continues along the entire length of the (original) tail in essentially the same form, forming a boundary on the otherwise plain upper surface of the tail.

The tail is not whitish with a series of well-defined black spots, arranged in rows, sometimes nearly or actually merging to form longitudinal lines.

On the top the tail fades from greyish yellow to grey at the distal parts.

Anterior upper labials are orange. The middle and posterior upper labials are immaculate white.

Gaia ruleofpriority sp. nov. is most similar in most respects to *Gaia taxonomymatters sp. nov*. especially with respect of the colouration of the (original) tail.

However, *G. ruleofpriority sp. nov.* differs from the preceding three species by having a deep yellow colour on the dorsum and an upper lateral stripe that is absent on the neck and only forms on the anterior part of the body. The lower edge bleeds yellow over the otherwise immaculate white scales below.

The tail is not whitish with a series of well-defined black spots, arranged in rows, sometimes nearly or actually merging to form longitudinal lines.

The upper labials are all whitish in colour.

The four preceding species are separated from all other species within the genus *Gaia* Wells, 2012 by the following unique combination of characters:

They are relatively small slender species with immovable eyelid, two toes, and no trace of the forelimb (including groove). Lacking a preocular and lower secondary temporal, two smallish supraoculars, body angular at change from lateral to ventral surface, and dark upper lateral stripe that may or may not be defined. No supraciliaries, 1-2 temporals, five upper labialis and 16-18 midbody scale rows.

They are further diagnosed as follows:

Snout-vent length to about 42 mm. Length of appendages (as % SVL): hindleg 15.2-21.3 (N 5, mean 18.3), tail 73-89 (N 4, mean 80). Nasals separated. No prefrontals. Frontal slightly wider than long. Nuchals 1-2 (N 5) Supraoculars 2, small, first in contact with frontal. No supraciliaries. Loreals 2. No preocular (fused to second loreal). Temporals usually 2, primary the smaller and occasionally fused to the fourth labial. Lamellae under longer toe 7-11 (N 5, mean 8.8). Dorsal surface buff, pinkish, yellowish, yellow, beige or very pale reddish brown, marked with dark brown as follows: 1-5 small irregular spots on frontonasal and frontal; sometimes 0-4 lines of small spots on back, passing through centre of the dorsal scales, central (paravertebral) pair sometimes extending on to the tail, where spots become larger, darker and more closely spaced (and when this happens the tail is whitish); and upper lateral stripe from nasal, back of head, neck or anterior of body to the end of the tail, broken on the head (if present), variably defined on body, ranging from ill-defined in its entirety to well-defined in its entirety, sometimes conspicuous on the tail. Ventral and lower lateral surfaces white or whitish (modified from Storr, 1982).

Species within the genus *Gaia* Wells, 2012 are separated from all other species within *Lerista sensu lato* as defined by Hoser (2024c), by the following unique combination of characters:

Snout depressed, very sharp in profile and extending well beyond the mouth.

Eye very small. Eyelid movable. No trace of foreleg (including groove). Two toes. Nasals separated moderately widely. No prefrontals. Frontal a little wider than long. Frontoparietals and interparietal fused into a large triangular shield. Parietals long and narrow, in short contact. Nuchals 1 or 2. Supraoculars 2, both in contact with frontal. Supraciliaries 0+1. Preoculars 1. Postoculars 3. Temporals 3, Upper labials usually 6, Ear aperture minute (modified from Storr, 1982).

G. vermicularis Storr, 1982 is depicted in life in Wilson and Swan (2021) on page 389 middle left.

Gaia nomenclaturematters Hoser, 2026 is depicted in life online at:

https://www.flickr.com/photos/reptileshots/52565034530/

and

https://www.flickr.com/photos/127392361@N04/54018619237/

and

https://www.flickr.com/photos/127392361@N04/54018619237/

and

https://www.flickr.com/photos/114192916@N07/54131338535/

and

https://www.flickr.com/photos/114192916@N07/54130869871/

and

https://www.flickr.com/photos/171250498@N08/54024898275/

Gaia taxonomymatters sp. nov. is depicted in life online at:

https://www.flickr.com/photos/54876436@N08/53163291726/

Gaia ruleofpriority sp. nov. is depicted in life online at:

https://www.flickr.com/photos/jaricornelis/52932485847/

Distribution: *Gaia nomenclaturematters sp. nov*. is a taxon from the general areas of the Tanami Desert and North Gibson Desert in east Western Australia and adjacent Northern Territory, Australia.

Etymology: In light of the recent (years 2012-2026) epidemic of taxonomic vandalism (as defined in Hoser 2015a-f) it is important to highlight the correct taxonomy and nomenclature that is necessary for science to progress. Integral in all this is the rules as given in the *International Code of Nomenclature* (Ride *et al*. 1999), being the current "in force" edition.

The most important rules within that document (simplified here) are the rules of homonymy, meaning no species can have more than one correct name, and the associated and essential rule of priority, meaning that the first name applied to a taxon is the correct one and the only one that should be used as correct.

For example the name *Broghammerus* Hoser, 2004, for the Reticulated Python genus has priority over the later illegally coined name *Malayopython* Wüster *et al.* 2014 and the genus name *Wellingtondella* Hoser, 2020 has date priority over *Anstisia* Webster and Bool, 2023 for a genus of West Australian frogs.

GAIA NOTTOBEMISSED SP. NOV.

LSIDurn:lsid:zoobank.org:act:A743E907-DA20-4B52-8B33-446039C9D210

Holotype: A preserved adult female specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R87348 collected from leaf litter 13 km east of Anna Plains Homestead, Western Australia, Australia, Latitude -19.266667 S., Longitude 121.616667 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R87349 collected from leaf litter 13 km east of Anna Plains Homestead, Western Australia, Australia, Latitude -19.266667 S., Longitude 121.616667 E.

Diagnosis: Predating the publication of Hoser (2024c), *Gaia griffini* (Storr, 1983), with a type locality of Kununurra, Western Australia, Australia, Latitude -15.47 S., Longitude 128.44 E., had been treated as a species occurring across the Kimberley District of Western Australia and nearby areas, across a wide zone in the lower top end of the Northern Territory.

This distribution is also disjunct.

Relying primarily on morphological evidence, Hoser (2024c), split off the eastern population as *Gaia griffini asgicondi* Hoser, 2024.

Gaia griffini asgicondi Hoser, 2024 has a centre of distribution on the Sturt Plateau bioregion of the north central Northern Territory including nearby hilly areas, such as the uplands near the south-west Gulf of Carpentaria.

There is a distributional gap of over 100 km straight line between these populations.

Populations of putative *Gaia griffini* from all parts of the Kimberley District were erroneously treated by Hoser (2024c) as being a single taxon, when it was stated "*The Kimberley population (both east and west Kimberley) are of the type subspecies*."

This error was shown in the paper of Singhal *et al.* (2025) in which they found three molecular divergent forms inhabiting the Kimberley District.

This included the type form from Kununurra and environs in the east Kimberley, a form from the west Kimberley and a form from the Dampier Peninsula and south along the nearby coast in Western Australia.

In a second error, Hoser (2024c) recognised the species originally described as "*Lerista praefrontalis* Greer, 1986" and mistakenly followed Wells (2012) by placing it in the genus *Rhodona* Gray, 1839, when it should have correctly been placed in the genus *Gaia* Wells, 2012.

"*Lerista praefrontalis* Greer, 1986" corresponds to the west Kimberley population and so there is no need to engage in Wüster / Britton / Kaiser style taxonomic vandalism by giving it a junior synonym name.

On the basis of distinct gaps in the distributions of all four forms of putative *Gaia griffini* (Storr, 1983), morphological and molecular divergences, all are herein treated as full species.

In other words they are referred to as *Gaia griffini* (Storr, 1983), *Gaia asgicondi* (Hoser, 2024), *Gaia praefrontalis* (Greer, 1986) and *Gaia nottobemissed sp. nov.*.

Gaia nottobemissed sp. nov. is herein formally described.

The diagnosis for *Gaia asgicondi* (Hoser, 2024) is modified slightly from that publication to accommodate the three taxa formerly all treated as Kimberley *G. griffini.*

The four relevant species are separated from one another as follows:

Gaia asgicondi (Hoser, 2024) is separated from nominate *Gaia griffini* by having a medium brownish tinge to the dorsum versus yellow with a slight reddish tinge in *G. griffini*, pinkish red in *G. praefrontalis* and light brown in *Gaia nottobemissed sp. nov.*.

G. asgicondi is further separated from the other three species by the second loreal which is narrow and triangular in shape, versus squarish in *G. griffini*, *G. praefrontalis* and *G. nottobemissed sp. nov.* The narrowing is caused by the scales above pushing their boundaries lower and the expansion at the lower edge of the first loreal.

The 2-3 postoculars present in *G. griffini*, *G. praefrontalis* and *G. nottobemissed sp. nov.* are also

reduced or absent in *G. asgicondi.*

G. asgicondi has no bleeding of yellow beyond the lower boundary of the dark brown lateral stripe on the body.

G. griffini is notably lighter in colour than the other three species, with a dorsum that is yellow with a slight reddish tinge. *G. griffini* has an obvious bleeding of yellow (over the white) beyond the lower boundary of the dark brown lateral stripe on the body.

G. praefrontalis according to Greer (1986) has phalanges of fourth toe of pes four, rather than five as seen in the other species. It also has a dorsum that is pinkish red.

G. nottobemissed sp. nov. has a light brown dorsum, with no, or very little bleeding of yellow below the dark of the upper lateral line. More than in the other three species, the dark of the sides of the anterior snout in front of the eye is concentrated and confined to form part of a distinctive dark stripe that forms from tip of snout through eye, neck and along the upper flank forming the lateral line.

Below this line, the upper labials are immaculate white.

In the other three species, the dark at the anterior side of the snout, while predominantly on the top part, is not strictly constrained, irregular in form and does not form a part of any obvious stripe.

In common with *G. asgicondi* the black spotting of scales on the dorsum of *G. nottobemissed sp. nov.* is very prominent, meaning obvious lines run down the body, versus not so in *G. griffini* and *G. praefrontalis.*

G. griffini, *G. praefrontalis*, *G. asgicondi* and *G. nottobemissed sp. nov.* are separated from all other species within the putative genus *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

A movable eyelid, two toes and no trace of a forelimb (including groove). Stout in build, snout vent to 70 mm, tail is nearly as long as the body, or rarely slightly longer; relatively dark in body colouration including with well-defined paravertebral stripes dorsally and a well-defined dark band on the dorsolateral edge; no prefrontals; nasal usually separated; a relatively low flat-topped second loreal (especially in the species *G. asgicondi* (Hoser, 2024)), that is significantly larger than the first; no preocular and digits 0 + 2.

G. griffini is depicted in life in Wilson and Swan (2021) on page 367, second from bottom on left and online at:

https://www.inaturalist.org/observations/203040488

and

https://www.flickr.com/photos/58349528@N02/52655204336/

G. asgicondi is depicted in life online at:

https://www.flickr.com/photos/zimny_anders/51599000858/

and

https://www.flickr.com/photos/58349528@N02/50227974482/

and

https://www.flickr.com/photos/zimny_anders/51599000748/

and

https://www.flickr.com/photos/58349528@N02/50440947107/

G. nottobemissed sp. nov. is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/54010116724/

and

https://www.flickr.com/photos/reptileshots/25033418298/

and

https://www.flickr.com/photos/euprepiosaur/24770205518/
and
https://www.flickr.com/photos/114192916@N07/54041997149/
and
https://www.inaturalist.org/observations/364610609
and
https://www.inaturalist.org/observations/244653862
and
https://www.inaturalist.org/observations/123929586

Distribution: *Gaia nottobemissed sp. nov.* is a taxon from the Dampier Peninsula, Western Australia and south along the nearby coast in Western Australia, at least as far south as Wallal Caravan Park, Eighty Mile Beach, Western Australia, Australia, Latitude -19.756111 S., Longitude 120.67194 E.

Etymology: Taken from the English phrase "not to be missed" this is exactly the status of this species at the time I decided to formally identify it and give it an ICZN scientific name. The spelling of the species name should not be changed.

GAIA QUESTIONTHENARRATIVE SP. NOV.

LSIDurn:lsid:zoobank.org:act:F66A97DF-C3FD-4432-A771-A17DF4A22B10

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R137925 collected from Telfer, Western Australia, Australia, Latitude -21.761111 E., Longitude 122.228889 E. This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R137923 collected from Telfer, Western Australia, Australia, Latitude -21.727222 E., Longitude 122.168333 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species herein.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. questionthenarrative sp. nov. is a species of the eastern Great Sandy Desert in Western Australia.

G. questionthenarrative sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. questionthenarrative sp. nov. is white at the anterior snout on top, with the posterior head shields being an immaculate light orange colour, usually without any speckling or spots. They are also thinly etched dark. The back of the head, top of neck and dorsum are a dark deep reddish-brown colour, grading to dark yellowish brown at the anterior part of the tail and purplish at the distal end.

Paravertebral lines are either reduced in intensity so as to be barely noticeable or alternatively absent.

The dark upper lateral stripe disintegrates at the anterior part of the tail and the main part of the tail top and sides is a combination of dark, orange and pinkish speckling and spots, giving it a unicolour non-patterned appearance.

There is a thick orange bleed over the white below the dark upper lateral stripe. The bleed itself is bold and makes a thin orange line at the lower edge of the dark upper lateral stripe. Upper surfaces of limbs are a dull yellow pink in colour.

Upper labials are all immaculate white.

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans*

Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. questionthenarrative sp. nov*. is a species of the eastern Great Sandy Desert in Western Australia.

Etymology: The species name for *G. questionthenarrative sp. nov*. is a direct piece of advice for all scientists and herpetologists in particular, this advice being to question the narrative. With narcissistic lies and fabrications masquerading as science as executed by the members of the Wolfgang Wüster, Adam Britton gang, including their non-stop egregious acts of taxonomic vandalism and rewriting history to justify their crimes, the need to question their narrative and those of like-minded narcissistic persons is important.

This means bypassing their acts of censorship on their controlled platforms like “The Reptile Database”, “Frog ID Ap” and others to actively seek out all the original literature for all matters science (including that which they pretend does not exist).

This includes that original literature relating to taxonomy and nomenclature.

When one does these proper checks of the (false) narrative one realises that they (Wüster/Britton and their gang) have engaged in acts of “DARVO” by falsely alleging that myself and others have engaged in taxonomic vandalism and anti-science, when in fact this is a crime that they themselves have committed.

As a rule of thumb, if a member of the Wüster/Britton gang claim to have discovered and named a new species, it is a near certainty that they have merely cut and pasted a description from an earlier work and then pretended that the earlier work was never published.

This gang have unlawfully assigned junior synonym names to reptiles and frogs over 100 times (to early 2026). Their databases and “official lists” invariably ignore the earlier works and recklessly peddle their junior synonyms as the only correct and available ICZN names.

GAIA RATE SP. NOV.

LSIDurn:lsid:zoobank.org:act:0579E40B-3D6C-42F4-8419-25173E6B4A9B

Holotype: A preserved male specimen at the Western Australian Museum, Perth, Western Australia, Australia collected in soil under tin, being specimen number R108783 collected from Egan’s Bore, Western Australia, Australia, Latitude -18.916667 S., Longitude 125.033333 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ A preserved male specimen at the Western Australian Museum, Perth, Western Australia, Australia collected in soil under tin, being specimen number R108784 collected from Egan's Bore, Western Australia, Australia, Latitude -18.916667 S., Longitude 125.033333 E. and, 2/ A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, collected in soil under iron, being specimen number R108762 collected from Egan's Bore, 50 km west of Cheerabun Homestead, Western Australia, Australia, Latitude -18.933333 S., Longitude 125.066667 E.

Diagnosis: Hoser (2024c) split the species *Gaia greeri* (Storr, 1982) as recognized then (being that of Storr, 1982) into two species based on very significant and obvious divergences, distributional disjuncture and known antiquated biogeographical boundaries.

Singhal *et al.* (2025) in their supplementary Fig S5 showed that Storr's concept of *G. greeri* included four putative species implying that Hoser (2024c) had not gone far enough.

This view was clarified somewhat by Torkkola *et al.* (2026) at Fig 1., where they showed that the Western type form was in fact split into two closely related species (one north form from Storr Island, West Kimberley and the south Type form of *G. greeri*, from 8 km south southeast of Derby, Western Australia, Latitude 17.22 S., Longitude 123.40 E.).

Additionally, Torkkola *et al.* (2026) found that an eastern sample, being West Australian Museum specimen number WAM R137387 collected from Fowlhouse Junction, Osmond Range, East Kimberley district of Western Australia, Latitude -17.266667 S., Longitude 128.483333 E. was a very, very divergent taxon.

This essentially was in line with the concept of Hoser (2024c) for *G. oomph* Hoser, 2024, supporting the erection of that species in the strongest possible terms.

It should be noted that the type form of *G. oomph* from the Tanami Desert, while similar in most respects to other East Kimberley animals previously referred to *G. greeri,* did differ from some.

Were it not for my inherently conservative nature, I would have recognised at least one further species or subspecies on the basis of the differences. However, myself being ever (over) sensitive to ongoing unsupported claims by detractors that I have over split reptile species, I refrained from doing so.

In hindsight, taking notice of unscientific keyboard warriors, working as flying monkeys for the Wüster / Britton / Kaiser gang, generally from fake accounts, was a serious mistake on my part.

To confirm my mistake in listening to trolls, Singhal *et al.* (2025) in their supplementary Fig S5 also indicated that there were in fact two species occupying the same general region, this being the inland south and east Kimberley districts.

In other words, only two of the relevant four species had been formally named until now, although this did include the two main lineages.

Rather than having the less divergent, but still unnamed species-divergent lineages expire through general indifference I formally name them both herein. This is so that proper conservation measures can be implemented by governments, non-governmental organizations and concerned individuals.

The south and south east Kimberley form is herein named *Gaia rate sp. nov.*, while the Storr Island form is herein named *Gaia otit sp. nov*..

This means that the diagnoses and concepts for *Gaia greeri* and *Gaia oomph* as given in Hoser (2024c) must be amended slightly to accommodate the two other species formally named within this species complex.

Gaia oomph and the newly named and most closely related species *Gaia rate sp. nov.* are morphologically and genetically divergent from the type form of *G. greeri*, the closely related *Gaia otit sp. nov*. and the closely related taxa, *G. robusta* (Storr, 1990), *G. vermicularis* (Storr, 1982), *G. nomenclaturematters sp. nov.* (this paper), *G. ruleofpriority sp. nov*. (this paper), *G. taxonomymatters sp. nov*. (this paper), *G. labialis* Storr (1971) or *G. simillima* (Storr, 1984). See for example the relevant sequence data published in Skinner *et al.* (2008), Farquhar *et al*. (2024), Singhal *et al.* (2025) or Torkkola *et al.* (2026).

Gaia oomph Hoser, 2024 and *Gaia rate sp. nov.* are readily separated from the previously named species as follows:

G. greeri, *G. otit sp. nov*., *G. labialis* and *G. simillima* are lizards with a bold dark lateral band that is either black, very dark brown and with a sharp upper and lower edge.

G. vermicularis, G. nomenclaturematters sp. nov. (this paper), *G. ruleofpriority sp. nov*. (this paper) and *G. taxonomymatters sp. nov*. are lizards with an orange red dorsal surface, sometimes pinkish, yellowish or orange instead, including to the snout and with white on the mid and lower flanks (below an apper lateral line that may or may not be well-defined). The white of the under surface extends about half-way up the side of the flank.

G. robusta is the most similar species to *G. oomph sp. nov.* and *G. rate sp. nov.*. The dorsal colour is orange on top. There is brown stripe on the upper lateral surface, reasonably sharply defined top and bottom and this occupies most of the side of the flank. The white from below only encroaches the lower edge of the flank. The distal end of the tail is grey.

Hoser 2026 - Australasian Journal of Herpetology 81-82:1-128.

A more distantly related species within *Gaia* Wells, 2012 is *G. ips* (Storr, 1980), that is separated from the other species by having a moderately to well-defined and wide mid-vertebral line that is yellow in colour on a dorsum that is otherwise orange, with a moderately well-defined orange upper lateral line extending about halfway down the side of the flank. Most of the tail is ghost white.

G. oomph and *G. rate sp. nov.* in contrast to all the preceding species is separated from them and defined as follows:

It has a dorsum that is a light beige-brown colour, extending along the tail.

The snout and back of head is whitish, darkening on the neck, lightening on the rear body and then intensifying on the anterior tail before becoming near white at the distal end.

The dorsal colouration extends over the dorsolateral edge where a boundary of darker brown occurs.

This is formed by one or two scale rows having the main part of each scale a darker brown colour.

Below this line that is poorly defined above and well-defined below is the white of the majority of the flank that extends to the white belly.

On the anterior half of the tail, the dark band of the upper lateral edge, remains 2 scales wide and becomes well defined on top as well as below, with sharper contrast in colour, giving the anterior tail a well-defined striped appearance. These colours reduce to become white or white with flecks at the distal end.

G. oomph and *G. rate sp. nov.* are further readily separated from *G. greeri* and *G. otit sp. nov*. by the fact that along the paravertebral lines spotting is small and distinct but in no way forming obvious lines in *G. oomph* or very obscured in the form of feint small dull spots or none at all in *G. otit sp. nov*., versus bold, black or blackish, enlarged or merged to form obvious paravertebral lines on the dorsum in both *G. greeri* and *G. otit sp. nov*.

G. rate sp. nov. is separated from *G. oomph* by not having a pink tinge on the dorsum (versus having one), yellow within the anterior part of the dark upper lateral band (versus pinkish orange) and by the fact that along the paravertebral lines spotting is small and distinct but in no way forming obvious lines in *G. oomph* versus very obscured in the form of feint small dull spots or none at all in *G. otit sp. nov*.,

G. greeri and the closely related *G. otit sp. nov*. are separated by the fact that the dorsum of *G. greeri* is dark yellowish or light-brown or even light greyish in colour, versus light greyish with a slight reddish or pinkish tinge in *G. otit sp. nov*. and upper surfaces of hind limbs of *G. greeri* that are mainly dark in colour, versus mainly light in *G. otit sp. nov*. (as well as *G. oomph* and *G. rate sp. nov.*).

G. oomph, *G. rate sp. nov.*, *G. greeri* and *G. otit sp. nov*. are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

No trace of a forelimb or forelimb groove; two toes; no prefrontals; nasals usually separated; 6 upper labials, 20 midbody rows, 2 supraoculars in contact with the frontal and 1-2 supraciliaries.

A detailed description of the type form of *G. greeri* is in Wells (2012) on pages 198-199 which is evidently largely applicable to *G. oomph* Hoser, 2024, *G. rate sp. nov.* and *G. otit sp. nov*. as well.

G. oomph Hoser, 2024 is depicted in life online at:

https://www.flickr.com/photos/58349528@N02/54678291202/

G. rate sp. nov. is depicted in life online at:

https://www.flickr.com/photos/euprepiosaur/53840611459/

and

https://www.flickr.com/photos/171250498@

N08/53835283097/

G. greeri (of the nominate type form) is depicted in life in Wilson and Swan (2021) on page 367 at top and online at:

https://www.flickr.com/photos/brian_busho/14270539460/

Distribution: *G. rate sp. nov.* is a taxon from the southern and southeast Kimberley District, well away from the coast in the driest parts of the region.

Of relevance *G. oomph* Hoser, 2024 is a taxon presently only known from the north-west Tanami in far east Western Australia and also the Ord River region of the east Kimberley District, also in Western Australia, from there extending into the nearby part of the Northern Territory.

G. greeri is mainly known from near the type locality of Derby, Western Australia and *G. otit sp. nov*. is known principally from Storr Island further north. Both areas are near the mainland coast in the west Kimberley district, with Storr Island only being separated from the mainland since the most recent glacial minimum.

Etymology: *G. rate sp. nov.* is a play on words. Combined in English the word "Grate" or "Great" which means really good, is exactly how an enthusiastic field herpetologist feels after finding their first specimen of this species.

GAIA ROBWATSONI SP. NOV.

LSIDurn:lsid:zoobank.org:act:5686B6A9-1748-4EE4-A212-8C328D834038

Holotype: A preserved specimen at the Division of Reptiles and Amphibians, University of Michigan Museum of Zoology, 1105 North University Ave, Ann Arbor, Michigan, 48109, USA, specimen number UMMZ 242664 collected from Dampier Downs Station, Site 2, at elevation of 93 m ASL, Western Australia, Australia, Latitude -18.114 S., Longitude 123.551 E.

This facility allows access to its holdings.

Paratypes: Eight preserved specimens at the Division of Reptiles and Amphibians, University of Michigan Museum of Zoology, 1105 North University Ave, Ann Arbor, Michigan, 48109, USA, specimen numbers 242661, 242662, 242663, 242665, 242666, 242667, 242669 and 242671 all collected from Dampier Downs Station, Western Australia, Australia, with the approximate Latitude -18.114 S., Longitude 123.551 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. robwatsoni sp. nov. is known only from around the Derby area in northwestern Western Australia.

G. wow sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. robwatsoni sp. nov. is slightly brownish on the dorsum anteriorly, with most of the dorsum being beige and the tail rapidly turning light grey on top.

The upper lateral line breaks on the neck and on the body is a slight bleed of brownish from the lower edge of the upper lateral line onto the immaculate white below. Head shields lack spots or etching of any sort, except those at the back of the head which have about 4 symmetrically placed small dark brown spots.

Upper surfaces of the legs are a light pinkish colour, with scattered tiny, semi-distinct dark purple spots.

The upper lateral stripe continues in a bold manner to the end of the tail. Below this is immaculate white.

All upper labials are immaculate white.

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu*

lato this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. robwatsoni sp. nov.* is known only from around the Derby area in northwestern Western Australia.

Etymology: *G. robwatsoni sp. nov.* is named in honour of Rob Watson, snake catcher in Brisbane, Queensland, Australia, with a business trading name of South-eastern Reptiles, in recognition of his services for public safety and wildlife conservation spanning many years.

GAIA RULEOFPRIORITY SP. NOV.

LSIDurn:lsid:zoobank.org:act:2CC09C5C-1B1B-41B8-8036-DD40BAB9862B

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R83650 collected from 52 km east southeast of Billiluna, Western Australia, Australia, Latitude - 19.183333 S., Longitude 128.233333 E.

This government-owned facility allows access to its holdings.

Diagnosis: As originally described and conceived "*Lerista vermicularis* Storr, 1982", herein transferred to the genus *Gaia* Wells, 2012 in line with Hoser (2024c) was a species only known from the Great Sandy Desert, in north-west Western Australia (see Storr, 1982 including "Figure 3").

More recently specimens that are morphologically similar from the northern extension of the Great Sandy Desert, Tanami Desert Region and the Gibson Desert Region have been collected, lodged in the Western Australian Museum and assigned to the same species.

The same applies to colour images of these lizards posted on photo sharing sites such as "Inaturalist" and "Flickr".

However, the lizards themselves (as seen in the images posted) are quite different in form to one another. They are separated by barriers of unsuitable non-dune habitat of significant antiquity, and the molecular results of Torkkola *et al.* (2026) have confirmed the existence of three very different forms.

The northern-most form was not included in the study of Torkkola *et al.* (2026) but is morphologically divergent from the others and also separated by

areas of unsuitable (non-dune) habitat, implying lengthy isolation.

Therefore, the three hitherto unnamed forms are formally named herein for the first time.

Gaia nomenclaturematters sp. nov. is the form previously assigned to *G. vermicularis* from the Tanami Desert and North Gibson Desert in east Western Australia.

Gaia taxonomymatters sp. nov. is the form previously assigned to *G. vermicularis* from the middle Gibson Desert in east Western Australia.

Gaia ruleofpriority sp. nov. occurs in a confined area east of Baines Hills and Elsey Hills in the area between the Great Sandy Desert (to the south-west), southeast Kimberley District (to the north), the open grass plains to the east and the Gardiner Range to the southeast.

The four species are separated from one another by the following combinations of characters:

G. vermicularis Storr, 1982 is orangish-red on the dorsum of the body. The upper lateral band commences on the neck and extends to the hind limb. It is poorly defined and medium brown in colour (being of similar intensity and colour to the dorsum, making it indistinct), being poorly defined on the upper edge and better defined on the lower edge. Below this the scales are either immaculate white or very faintly and thinly etched.

The (original) tail is whitish with a series of well-defined black spots, arrange in rows, sometimes nearly or actually merging to form longitudinal lines.

Upper labials are immaculate white at the lower edge and dark above.

Gaia nomenclaturematters sp. nov. is pinkish yellow on the dorsum, with a well-defined dark brown stripe on the upper flank (being very distinct and separate from the colour on the dorsum and below the stripe). This stripe commences at the back of the head and extends to the hind limb. It is well defined on both upper and lower edge.

The (original) tail is whitish with a series of well-defined black spots, arrange in rows, sometimes nearly or actually merging to form longitudinal lines.

Upper labials are orange, becoming whitish posterior to the eye.

Gaia taxonomymatters sp. nov. is a light greyish yellow on the dorsum, with a dark upper stripe along the upper flank commencing from the back of the head and running to the tail. It is reasonably week defined at the anterior end and bold on the latter part of the body. The lower edge bleeds yellowish orange over the otherwise immaculate white scales below. This lateral stripe continues along the entire length of the (original) tail in essentially the same form, forming a boundary on the otherwise plain upper surface of the tail.

The tail is not whitish with a series of well-defined black spots, arranged in rows, sometimes nearly or actually merging to form longitudinal lines.

On the top the tail fades from greyish yellow to grey at the distal parts.

Anterior upper labials are orange. The middle and posterior upper labials are immaculate white.

Gaia ruleofpriority sp. nov. is most similar in most respects to *Gaia taxonomymatters sp. nov*. especially with respect of the colouration of the (original) tail.

However, *G. ruleofpriority sp. nov.* differs from the preceding three species by having a deep yellow colour on the dorsum and an upper lateral stripe that is absent on the neck and only forms on the anterior part of the body. The lower edge bleeds yellow over the otherwise immaculate white scales below.

The tail is not whitish with a series of well-defined black spots, arranged in rows, sometimes nearly or actually merging to form longitudinal lines.

The upper labials are all whitish in colour.

The four preceding species are separated from all other species within the genus *Gaia* Wells, 2012 by the following unique combination of characters:

They are relatively small slender species with immovable eyelid, two toes, and no trace of the forelimb (including groove). Lacking a preocular and lower secondary temporal, two smallish supraoculars, body angular at change from lateral to ventral surface, and dark upper lateral stripe that may or may not be defined. No supraciliaries, 1-2 temporals, five upper labialis and 16-18 midbody scale rows.

They are further diagnosed as follows:

Snout-vent length to about 42 mm. Length of appendages (as % SVL): hindleg 15.2-21.3 (N 5, mean 18.3), tail 73-89 (N 4, mean 80). Nasals separated. No prefrontals. Frontal slightly wider than long. Nuchals 1-2 (N 5) Supraoculars 2, small, first in contact with frontal. No supraciliaries. Loreals 2. No preocular (fused to second loreal). Temporals usually 2, primary the smaller and occasionally fused to the fourth labial. Lamellae under longer toe 7-11 (N 5, mean 8.8). Dorsal surface buff, pinkish, yellowish, yellow, beige or very pale reddish brown, marked with dark brown as follows: 1-5 small irregular spots on frontonasal and frontal; sometimes 0-4 lines of small spots on back, passing through centre of the dorsal scales, central (paravertebral) pair sometimes extending on to the tail, where spots become larger, darker and more closely spaced (and when this happens the tail is whitish); and upper lateral stripe from nasal, back of head, neck or anterior of body to the end of the tail, broken on the head (if present), variably defined on body, ranging from ill-defined in its entirety to well-defined in its entirety, sometimes conspicuous on the tail. Ventral and lower lateral

Hoser 2026 - Australasian Journal of Herpetology 81-82:1-128.

surfaces white or whitish (modified from Storr, 1982).

Species within the genus *Gaia* Wells, 2012 are separated from all other species within *Lerista sensu lato* as defined by Hoser (2024c), by the following unique combination of characters:

Snout depressed, very sharp in profile and extending well beyond the mouth. Eye very small. Eyelid movable. No trace of foreleg (including groove). Two toes. Nasals separated moderately widely. No prefrontals. Frontal a little wider than long. Frontoparietals and interparietal fused into a large triangular shield. Parietals long and narrow, in short contact. Nuchals 1 or 2. Supraoculars 2, both in contact with frontal. Supraciliaries 0+1. Preoculars 1. Postoculars 3. Temporals 3, Upper labials usually 6, Ear aperture minute (modified from Storr, 1982).

G. vermicularis Storr, 1982 is depicted in life in Wilson and Swan (2021) on page 389 middle left.

Gaia nomenclaturematters Hoser, 2026 is depicted in life online at:

https://www.flickr.com/photos/reptileshots/52565034530/

and

https://www.flickr.com/photos/127392361@N04/54018619237/

and

https://www.flickr.com/photos/127392361@N04/54018619237/

and

https://www.flickr.com/photos/114192916@N07/54131338535/

and

https://www.flickr.com/photos/114192916@N07/54130869871/

and

https://www.flickr.com/photos/171250498@N08/54024898275/

Gaia taxonomymatters sp. nov. is depicted in life online at:

https://www.flickr.com/photos/54876436@N08/53163291726/

Gaia ruleofpriority sp. nov. is depicted in life online at:

https://www.flickr.com/photos/jaricornelis/52932485847/

Distribution: *Gaia ruleofpriority sp. nov*. is presently only known from a confined area east of Baines Hills and Elsey Hills in the area between the Great Sandy Desert (to the south-west), southeast Kimberley District (to the north), the open grass plains to the east and the Gardiner Range to the southeast.

Etymology: In light of the recent (years 2012-2026) epidemic of taxonomic vandalism (as defined in Hoser 2015a-f) it is important to highlight the correct taxonomy and nomenclature that is necessary for science to progress. Integral in all this is the rules as given in the *International Code of Nomenclature* (Ride *et al*. 1999), being the current "in force" edition.

The most important rules within that document (simplified here) are the rules of homonymy, meaning no species can have more than one correct name, and the associated and essential rule of priority, meaning that the first name applied to a taxon is the correct one and the only one that should be used as correct.

For example the name *Leiopython hoserae* Hoser, 2000 for the Black Southern White Lipped Python, being the first available name for this iconic species often called the "Hoser's Python" has date priority over the illegally coined junior synonym of the Wüster *et al.* 2014 name *Leiopython meridionalis* Schleip, 2014.

The latter name should never be used as correct and merely wastes the time of scientists for generations to come who must now as a matter of formality include it in synonyms lists.

GAIA TAXONOMYMATTERS SP. NOV.

LSIDurn:lsid:zoobank.org:act:834A3225-5999-46BD-B949-5339895DDFBF

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R164271 collected from Mina Mina Clutterbuck Hills, Gibson Desert, Western Australia, Australia, Latitude -24.517222 S., Longitude 126.217778 E.

This government-owned facility allows access to its holdings.

Paratypes: Four preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R164248, R164286, R164287 and R164307, collected from Mina Mina Clutterbuck Hills, Gibson Desert, Western Australia, Australia, Latitude -24.517222 S., Longitude 126.217778 E.

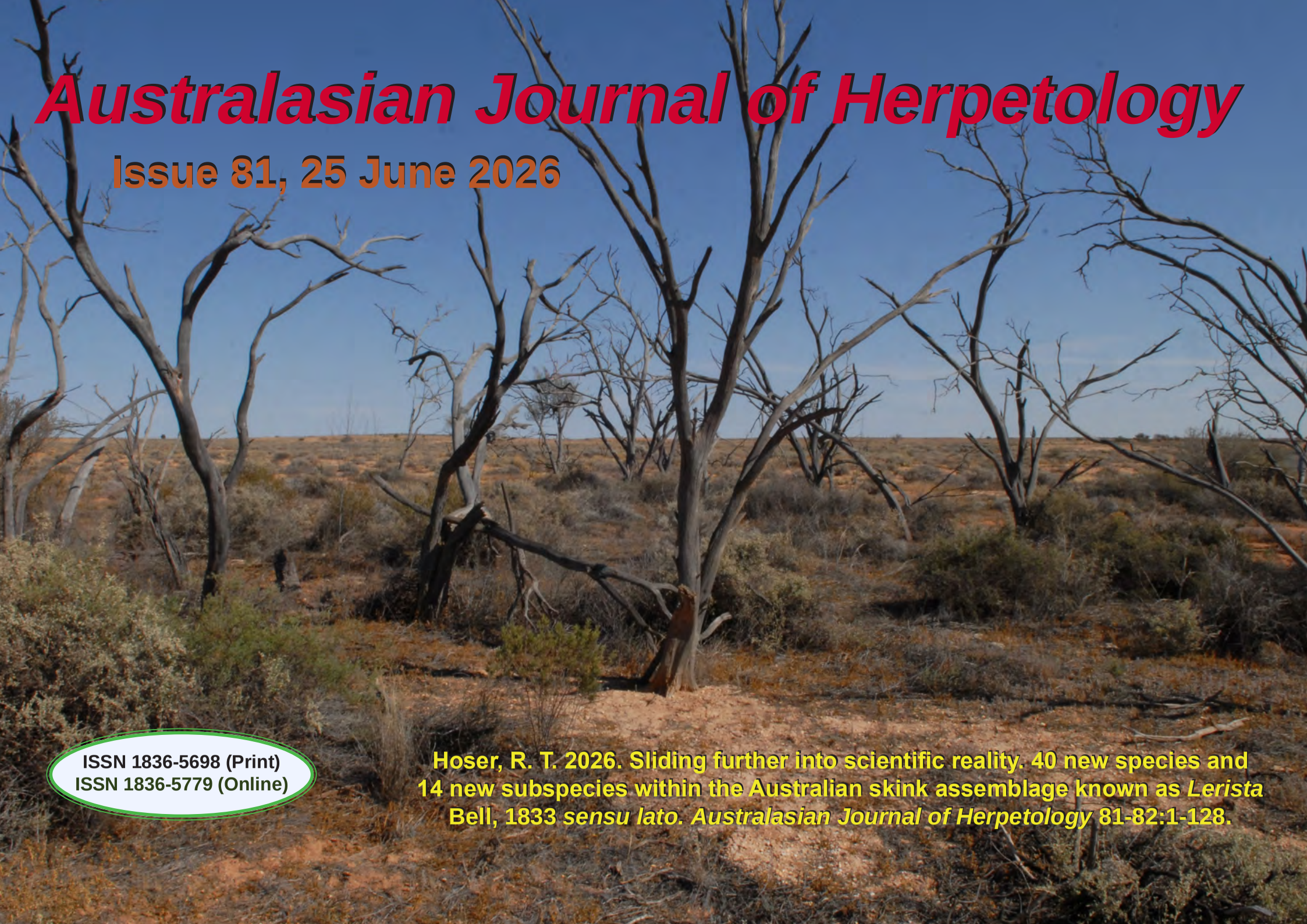
Diagnosis: As originally described and conceived "*Lerista vermicularis* Storr, 1982", herein transferred to the genus *Gaia* Wells, 2012 in line with Hoser (2024c) was a species only known from the Great Sandy Desert, in north-west Western Australia (see Storr, 1982 including "Figure 3").

More recently specimens that are morphologically similar from the northern extension of the Great Sandy Desert, Tanami Desert Region and the Gibson Desert Region have been collected, lodged in the Western Australian Museum and assigned to the same species. The same applies to colour images of these lizards posted on photo sharing sites such as "Inaturalist" and "Flickr".

... Continues in *Australasian Journal of Herpetology* Issue 82 ...

Australasian Journal of Herpetology

Issue 81, 25 June 2026



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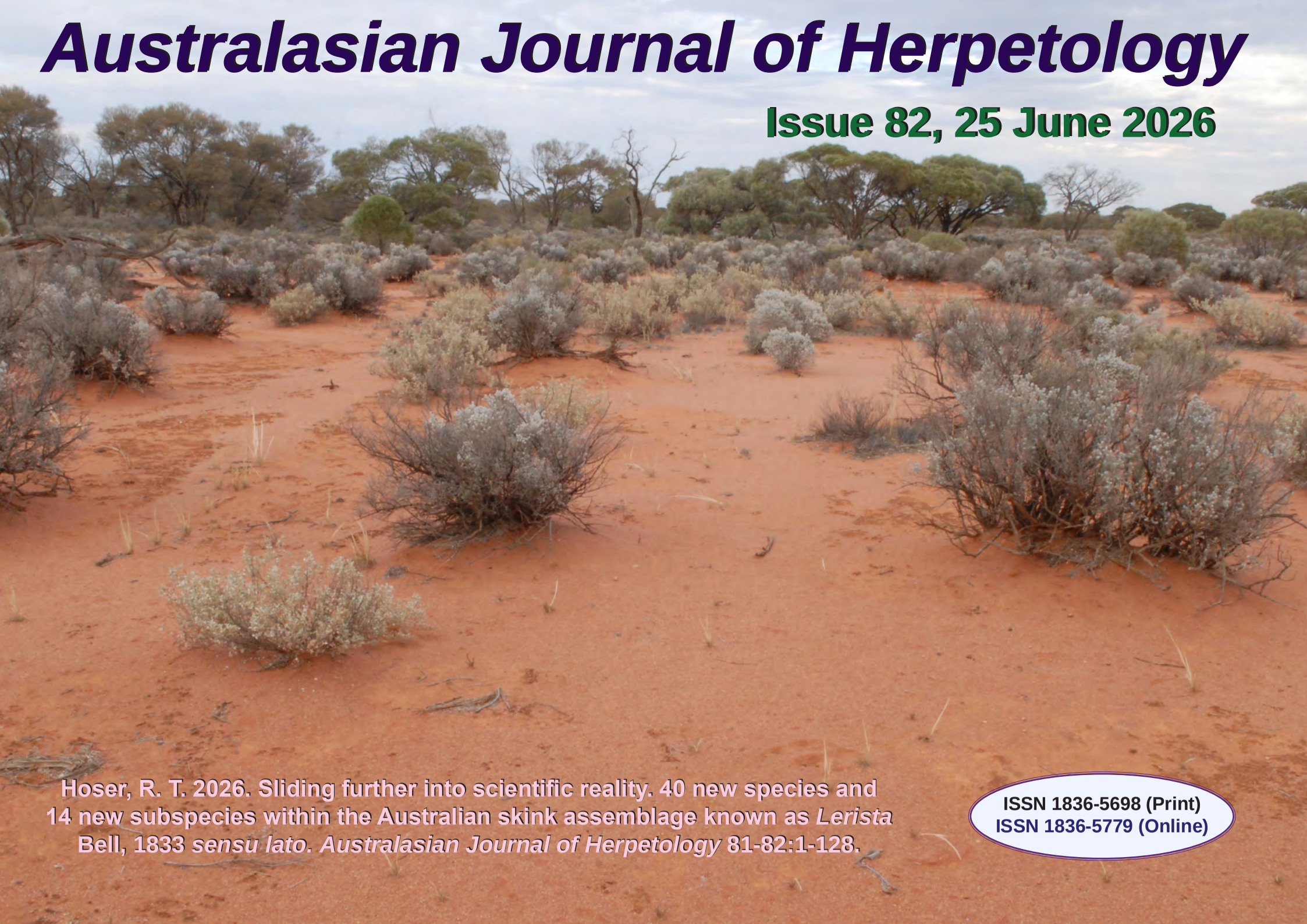
Australasian Journal of Herpetology

Issue 82, 25 June 2026



Hoser, R. T. 2026. Sliding further into scientific reality. 40 new species and 14 new subspecies within the Australian skink assemblage known as *Lerista* Bell, 1833 *sensu lato*. *Australasian Journal of Herpetology* 81-82:1-128.

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... Continued from *Australasian Journal of Herpetology* Issue 81 ...

GAIA TAXONOMYMATTERS SP. NOV.
CONTINUED ....

However, the lizards themselves (as seen in the images posted) are quite different in form to one another. They are separated by barriers of unsuitable non-dune habitat of significant antiquity, and the molecular results of Torkkola *et al.* (2026) have confirmed the existence of three very different forms.

The northern-most form was not included in the study of Torkkola *et al.* (2026) but is morphologically divergent from the others and also separated by areas of unsuitable (non-dune) habitat, implying lengthy isolation.

Therefore, the three hitherto unnamed forms are formally named herein for the first time.

Gaia nomenclaturematters sp. nov. is the form previously assigned to *G. vermicularis* from the Tanami Desert and North Gibson Desert in east Western Australia.

Gaia taxonomymatters sp. nov. is the form previously assigned to *G. vermicularis* from the middle Gibson Desert in east Western Australia.

Gaia ruleofpriority sp. nov. occurs in a confined area east of Baines Hills and Elsey Hills in the area between the Great Sandy Desert (to the south-west), southeast Kimberley District (to the north), the open grass plains to the east and the Gardiner Range to the southeast.

The four species are separated from one another by the following combinations of characters:

G. vermicularis Storr, 1982 is orangish-red on the dorsum of the body. The upper lateral band commences on the neck and extends to the hind limb. It is poorly defined and medium brown in colour (being of similar intensity and colour to the dorsum, making it indistinct), being poorly defined on the upper edge and better defined on the lower edge. Below this the scales are either immaculate white or very faintly and thinly etched.

The (original) tail is whitish with a series of well-defined black spots, arrange in rows, sometimes nearly or actually merging to form longitudinal lines.

Upper labials are immaculate white at the lower edge and dark above.

Gaia nomenclaturematters sp. nov. is pinkish yellow on the dorsum, with a well-defined dark brown stripe on the upper flank (being very distinct and separate from the colour on the dorsum and below the stripe). This stripe commences at the back of the head and extends to the hind limb. It is well defined on both upper and lower edge.

The (original) tail is whitish with a series of well-defined black spots, arrange in rows, sometimes nearly or actually merging to form longitudinal lines.

Upper labials are orange, becoming whitish posterior to the eye.

Gaia taxonomymatters sp. nov. is a light greyish yellow on the dorsum, with a dark upper stripe along the upper flank commencing from the back of the head and running to the tail. It is reasonably week defined at the anterior end and bold on the latter part of the body. The lower edge bleeds yellowish orange over the otherwise immaculate white scales below. This lateral stripe continues along the entire length of the (original) tail in essentially the same form, forming a boundary on the otherwise plain upper surface of the tail.

The tail is not whitish with a series of well-defined black spots, arranged in rows, sometimes nearly or actually merging to form longitudinal lines.

On the top the tail fades from greyish yellow to grey at the distal parts.

Anterior upper labials are orange. The middle and posterior upper labials are immaculate white.

Gaia ruleofpriority sp. nov. is most similar in most respects to *Gaia taxonomymatters sp. nov*. especially with respect of the colouration of the tail.

However, *G. ruleofpriority sp. nov.* differs from the preceding three species by having a deep yellow colour on the dorsum and an upper lateral stripe that is absent on the neck and only forms on the anterior part of the body. The lower edge bleeds yellow over the otherwise immaculate white scales below.

The tail is not whitish with a series of well-defined black spots, arranged in rows, sometimes nearly or actually merging to form longitudinal lines.

The upper labials are all whitish in colour.

The four preceding species are separated from all other species within the genus *Gaia* Wells, 2012 by the following unique combination of characters:

They are relatively small slender species with immovable eyelid, two toes, and no trace of the forelimb (including groove). Lacking a preocular and lower secondary temporal, two smallish supraoculars, body angular at change from lateral to ventral surface, and dark upper lateral stripe that may or may not be defined. No supraciliaries, 1-2 temporals, five upper labialis and 16-18 midbody scale rows.

They are further diagnosed as follows:

Snout-vent length to about 42 mm. Length of appendages (as % SVL): hindleg 15.2-21.3 (N 5, mean 18.3), tail 73-89 (N 4, mean 80). Nasals separated. No prefrontals. Frontal slightly wider than long. Nuchals 1-2 (N 5) Supraoculars 2, small, first in contact with frontal. No supraciliaries. Loreals 2. No

preocular (fused to second loreal). Temporals usually 2, primary the smaller and occasionally fused to the fourth labial. Lamellae under longer toe 7-11 (N 5, mean 8.8). Dorsal surface buff, pinkish, yellowish, yellow, beige or very pale reddish brown, marked with dark brown as follows: 1-5 small irregular spots on frontonasal and frontal; sometimes 0-4 lines of small spots on back, passing through centre of the dorsal scales, central (paravertebral) pair sometimes extending on to the tail, where spots become larger, darker and more closely spaced (and when this happens the tail is whitish); and upper lateral stripe from nasal, back of head, neck or anterior of body to the end of the tail, broken on the head (if present), variably defined on body, ranging from ill-defined in its entirety to well-defined in its entirety, sometimes conspicuous on the tail. Ventral and lower lateral surfaces white or whitish (modified from Storr, 1982).

Species within the genus *Gaia* Wells, 2012 are separated from all other species within *Lerista sensu lato* as defined by Hoser (2024c), by the following unique combination of characters:

Snout depressed, very sharp in profile and extending well beyond the mouth.

Eye very small. Eyelid movable. No trace of foreleg (including groove). Two toes. Nasals separated moderately widely. No prefrontals. Frontal a little wider than long. Frontoparietals and interparietal fused into a large triangular shield. Parietals long and narrow, in short contact. Nuchals 1 or 2. Supraoculars 2, both in contact with frontal. Supraciliaries 0+1. Preoculars 1. Postoculars 3. Temporals 3, Upper labials usually 6, Ear aperture minute (modified from Storr, 1982).

G. vermicularis Storr, 1982 is depicted in life in Wilson and Swan (2021) on page 389 middle left.

Gaia nomenclaturematters Hoser, 2026 is depicted in life online at:

https://www.flickr.com/photos/reptileshots/52565034530/

and

https://www.flickr.com/photos/127392361@N04/54018619237/

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https://www.flickr.com/photos/114192916@N07/54131338535/

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https://www.flickr.com/photos/114192916@N07/54130869871/

and

https://www.flickr.com/photos/171250498@N08/54024898275/

Gaia taxonomymatters sp. nov. is depicted in life online at:

https://www.flickr.com/photos/54876436@N08/53163291726/

Gaia ruleofpriority sp. nov. is seen in life online at:

https://www.flickr.com/photos/jaricornelis/52932485847/

Distribution: *Gaia taxonomymatters sp. nov*. is presently only known from the middle Gibson Desert in east Western Australia.

Etymology: In light of the recent (years 2009-2026) epidemic of taxonomic vandalism (as defined in Hoser 2015a-f) it is important to highlight the correct taxonomy and nomenclature that is necessary for science to progress. Integral in all this is the rules as given in the *International Code of Nomenclature* (Ride *et al*. 1999), being the current "in force" edition.

The most important rules within that document (simplified here) are the rules of homonymy, meaning no species can have more than one correct name, and the associated and essential rule of priority, meaning that the first name applied to a taxon is the correct one and the only one that should be used as correct.

Species need to be correctly identified, known and named so that everyone, from government departments, scientists, non-government organisations and individuals can work towards the same common objectives, including science and conservation.

Pretending that obvious divergent species do not exist, simply as a means to attack and discredit a "rival" scientist who named it, as has been done by the Wüster / Britton / Kaiser gang and their flying monkeys has caused serious negative wildlife conservation outcomes including the likely dooming of several species to unnecessary extinctions (see Hoser 2019a-b).

GAIA TRUTHISSCIENCE SP. NOV.

LSIDurn:lsid:zoobank.org:act:96BA5FD7-877B-49F9-804E-2612452832FD

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R166083 collected from Neale Junction Nature Reserve, Western Australia, (about 645 km by road northeast of Kalgoorlie), Australia, Latitude -28.102222 S., Longitude 125.959167 E.

This government-owned facility allows access to its holdings.

Paratype: Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R163389 and R163390 both collected from Neale Junction Nature Reserve, Western Australia, (about 645 km by road northeast of Kalgoorlie), Australia, Latitude -28.102222 S., Longitude 125.959167 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. truthisscience sp. nov. is a species from the Goldfields area and nearby parts of south-east Western Australia, well north of the coast.

G. truthisscience sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. truthisscience sp. nov. is a beige coloured lizard on the dorsum. The spots forming the paravertebral line are square in shape, rather than rectangular.

At the back of the top of the head, from the eye back are 8 medium-sized brown spots on the scales arranged to form a V-shaped arrangement.

Most of the tail is also beige and there is no difference in colour or intensity between the paravertebral lines on the body or on the tail.

Anterior labials are a dull orange colour, while posterior ones are an immaculate white.

There is a very slight bleeding of beige below the dark upper lateral line.

Within the dark lateral line, the centre of each otherwise dark brown scale is a dull orange colour, this central part being centred on the anterior edge of each scale.

The white scales on the lower flank have dull pink etchings on the posterior edges.

Upper surfaces of the (hind) limbs are white or whitish, either immaculate or with scattered tiny slightly darker spots or bars.

G. truthisscience sp. nov. is depicted in life online at:

https://www.flickr.com/photos/zimny_anders/48711141222/

and

https://www.flickr.com/photos/zimny_anders/48711101492/

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement

of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. truthisscience sp. nov*. is a species from the Goldfields area and nearby parts of south-east Western Australia, well north of the coast.

Etymology: Science is sometimes called a search for the truth. This is apt. Because lies in science get exposed eventually as fraud. If and when lies enter science and they are believed, the progress of science may be seriously impeded.

With taxonomy and nomenclature, the damage of lies doesn't just eventually show up in synonyms lists, but can also cause unwarranted extinctions of taxa either not recognised or improperly classified based on fraudulent claims.

Just as narcissism behaviours awareness will reduce the likelihood of lies being accepted by disconnected scientists who may otherwise believe false narratives, it is important to highlight the need for all scientists to rigidly stick to the truth when reporting their studies, results, theories or conclusions.

It is also important that mistakes made by a given scientist be corrected as soon as possible and this means and includes by the original publishing scientist if possible.

GAIA VELLAI SP. NOV.

LSIDurn:lsid:zoobank.org:act:74C90DDF-2535-4F38-B384-4721B771953A

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R116605 collected from 1 km northwest of Bullara Homestead, Western Australia, Australia, Latitude -22.683333 S., Longitude 114.05 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R116606 and R116608 both collected from 1 km northwest of Bullara Homestead, Western Australia, Australia, Latitude -22.683333 S., Longitude 114.05 E., 2/ Six preserved specimens at the Australian Museum, Sydney, New South Wales, Australia, specimen numbers R.101607, R.101608, R.101609, R.101610, R.101611 and R.101612 all collected from just west of Bullara Homestead, Western Australia, Australia, Latitude -22.7 S., Longitude 114.03333 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. vellai sp. nov. is a species of the west coastal south Pilbara, south of the Fortescue River and including nearby dune country to the south.

G. vellai sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. vellai sp. nov. is a generally light orange coloured lizard on top of the body with a mainly light grey tail. As a rule, the dorsal head shields are immaculate orange and with no, or very few tiny black dots. The paravertebral lines formed by the dots on the scales are relatively indistinct, the individual spots being small and circular in shape.

From the snout, all the way along the body to the hind legs there is a significant bleed of orange below the lower edge of the dark upper lateral stripe. Unusual among species in this complex, is the fact that between the paravertebral lines of dots the scales colour is a dark orange, versus a light orange outside these lines of spots. This differential in colouration is most intense above the pelvic girdle.

These paravertebral lines remain for most of the length of the tail, but inside these spotted lines and outside the intensity of the grey of the tail and colour is the same.

The dark upper lateral line continues in prominent form along most of the length of the tail. Beneath this line there is dark spotting on the white lower parts of the tail.

Upper labials are mainly orange except for the white lower edges of the posterior ones.

G. vellai sp. nov. is depicted in life online at:

https://www.flickr.com/photos/euprepiosaur/54567679222/

and

https://www.flickr.com/photos/114192916@N07/54080275589/

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some

Hoser 2026 - Australasian Journal of Herpetology 81-82:1-128.

specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *G. vellai sp. nov.* is a species of the west coastal south Pilbara, south of the Fortescue River and including nearby dune country to the south.

Etymology: *G. vellai sp. nov.* is named in honour of Matthew Vella of Regentville, New South Wales in recognition for his services to herpetology in Australia spanning many years including through the Australian Herpetological Society and Hawkesbury Herpetological Society.

GAIA WOW SP. NOV.

LSIDurn:lsid:zoobank.org:act:CA59B0F4-12CF-4D60-9F04-8B3FED3C8488

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R139070 collected from Mandora, Western Australia, Australia, Latitude -19.779444 S., Longitude 121.365833 E.

This government-owned facility allows access to its holdings.

Paratypes: Six preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R139069, R139071, R139117, R139118, R139119 and R139129 all collected from Mandora, Western Australia, Australia, Latitude -19.779444 S., Longitude 121.365833 E.

Diagnosis: Hoser (2024c) split the *Gaia bipes* (Fischer, 1882) complex into three species being the type form *G. bipes*, with a type locality of Nickol Bay, (Northwest Pilbara District of Western Australia), Australia, *G. kunja* Hoser, 2024 with a type locality of Naccowlah, 36 km west northwest of Jackson, in far south-west Queensland, Australia, Latitude -27.566667 S., Longitude 142.05 E. and *G. arrernte* Hoser, 2024 with a type locality of 42 km southwest of Wauchope, Northern Territory, Australia, Latitude -21.016667 S., Longitude 134.2 E.

The molecular phylogeny of Torkkola *et al.* (2026) showed no less than 7 species level populations of putative "*Lerista bipes*" in Western Australia alone. Singhal *et al*. (2025) found at least one more in south-west South Australia and so far there are no molecular results at all published for the complex in the Northern Territory, Queensland and northwest New South Wales.

As species within the complex are effectively tied to sand dune country, a plotting of the populations against habitat shows each has speciated in areas surrounded by zones of unsuitable habitat in the form of heavy dark soils, clay pans and the like.

The eight genetically divergent unnamed forms identified by Torkkola *et al.* (2026) and Singhal *et al*. (2025) are formally named as new species in this paper.

In addition to this *G. kunja* is herein restricted to far south-west Queensland and adjacent parts of northwest New South Wales and northeast South Australia, mainly east of the Lake Eyre basin. *G. arrernte* is restricted to most of the Northern Territory north of the Amadeus basin and south of the tropics.

Even with the new taxonomy and nomenclature for the *G. bipes* complex, actual species diversity is still quite likely underestimated.

This paper identifies and separates thirteen species in the *G. bipes* complex, the detail of which is in the formal description of *G. amazing sp. nov*. in this paper and that is relied upon explicitly as part of this formal description.

G. wow sp. nov. is a taxon found from Port Hedland to around Sandfire Flat/Mandora in northwest Western Australia.

G. wow sp. nov. is separated from the other 12 species in the *G. bipes* complex by the following unique combination of characters:

G. wow sp. nov. is a variable taxon, but invariably with the following characters:

It is an intensely coloured lizard.

The head and neck is usually a deep, orange colour. The dark upper lateral band is of reduced size and intensity on the body (often starting some way down the body and not on the head) and what separates this species from all others in the complex is that this bad is most bold on most of the tail, rather than on the body (versus the reverse in all the other species in the complex).

Anterior upper labials are usually orange, while the posterior ones are usually white below and orange above.

The dorsum of the lizard is mainly a yellowish orange colour, while the upper surface of the tail abruptly changes to a light whitish grey colour along its entire length on top.

The upper part of each side of the tail has a bold black stripe, below being immaculate white and with no spotting or marks of any sort.

Upper surfaces of the limbs are mainly a dark purplish brown, being comprised of close spaced bars, spots or peppering, overlaying a pinkish purple background.

G. wow sp. nov. is depicted in life online at:

https://www.flickr.com/photos/114192916@N07/54079664425/

The 13 species within the *G. bipes* complex are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans*

Gray, 1845, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small slender species with digits 0 + 2; snout depressed, very sharp in profile, extending well beyond the mouth; a moveable eyelid; frontoparietals and interparietal fused: no prefrontals (separating these species from the closely related, morphologically similar *G. nichollsi* (Loveridge, 1933) of the Gascoyne region in Western Australia, which has prefrontals) no trace of fore-limbs (including groove), 0 + 1 or 2 supraciliaries; two or three supraoculars in contact with the frontal (in contrast to the morphologically similar and closely related *G. labialis* (Storr, 1971) where they are not in contact).

Length of appendages (percentage of snout-vent length): hind-leg 12.9-21.8: tail 67-106.

Nasals narrowly separated (occasionally just touching, very rarely in short contact or widely separated). No prefrontals. 0-4 nuchals. 5 or rarely 6 upper labials. Primary and upper secondary temporals subequal, lower secondary much smaller. 18-20 midbody rows. 7-15 lamellae under the longer toe. Dorsally light yellowish to brown, often reddish or orange.

There is usually a line of brown dots through the paravertebral scales from the nape nearly to the end of the tail; less frequently there is an additional but fainter series of dorsal dots between the middle rows and the lateral edge. A dark-brown stripe usually runs from the nasal through the orbit nearly to the end of the tail, although this is sometimes reduced in some specimens or species. Lips (usually), lower flanks and belly are immaculate white or whitish (modified and corrected from Hoser 2024c).

Distribution: *Gaia wow sp. nov.* is a taxon found from Port Hedland to around Sandfire Flat/Mandora in northwest Western Australia.

Etymology: When I found my first specimen of this taxon in 1981, my companion, Dusty Brown the grader driver for Goldsworthy Mining exclaimed "Wow" and hence the species name.

GO FORIT SP. NOV.

LSIDurn:lsid:zoobank.org:act:2C4E5AB1-CAE6-4F31-B5E1-E9D64BB67F30

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R50411 collected from 1 km east southeast of Leigh Creek Homestead, Leigh Creek, South Australia, Australia, Latitude -30.5486 S., Longitude 138.4814 E.

This government-owned facility allows access to its holdings.

Paratypes: Three preserved specimens at the South Australian Museum, Adelaide, South Australia, Australia, being specimen number R50353 collected from 6.8 km south southeast of North Mooloolloo Homestead at North Mooloolloo Station, South Australia, Australia, Latitude -30.6842 S., Longitude 138.5383 E., specimen number R52510 collected from 0.2 km north of Warraweena Homestead, South Australia, Australia, Latitude -30.7833 S., Longitude 138.6389 E., and male specimen number R5269 collected from 1.5 km southeast of Warraweena Homestead, South Australia, Australia, Latitude -30.7794 S., Longitude 138.6481 E.

Diagnosis: For most of the past 160 years, the species originally described as "*Rhodona punctato-vittata* Günther, 1867" with a type locality of "Queensland" has been treated as a single species occupying the major part of inland eastern Australia.

This remained the case until relatively recently when Hoser (2024c) created the new genus *Go* Hoser, 2024 to accommodate this divergent lineage.

At the relevant time the three recognised species were:

Go punctatovittata (Günther, 1867) of the Murray Darling System, *Go emmotti* (Ingram, Couper and Donnellan, 1993) of the Cooper's Creek drainage and *Go colliveri* (Couper and Ingram, 1992) of the Burdekin River drainage basin in north-east Queensland), the genus being mainly west of the Great Dividing Range in Eastern Australia (except for the eastern distribution of *G. colliveri*).

Singhal *et al*. (2025) found three species within

putative *Go punctatovittata.* These were the type form from inland southern Queensland (and also most of western New South Wales, except for the far south-west), another from north-west Victoria and nearby South Australia as well as far southwest New South Wales and a third lineage from the northern Flinders Ranges, in the Lake Torrens drainage basin in South Australia.

The north-west Victorian population has an available name in the form of "*Rhodona officeri* McCoy, 1881" with a type locality of Swan Hill in Victoria.

Therefore, there is no need for me to engage in Wolfgang Wüster / Adam Britton / Hinrich Kaiser style taxonomic vandalism in renaming this taxon and pretending that McCoy's work never existed (see Hoser 2024c for details and relevant citations as well as the original work of McCoy (1881) cited herein).

Cogger *et al.* (1983) synonymised "*Rhodona officeri* McCoy, 1881" with their *Lerista punctatovittata* (Günther, 1867) and in this paper I formally reverse this by resurrecting it from synonymy.

The northern Flinders Ranges population in South Australia is genetically and morphology divergent and so I have no hesitation in naming it as a new species herein, being *Go forit sp. nov*..

Go forit sp. nov. is readily separated from all other species within *Go* Hoser, 2024 by the following unique combination of characters:

Barred upper labials that are more light than dark, etched shields on the upper parts of the head, the shields otherwise immaculate in colour, a brownish-red dorsum, which on the upper flank rapidly transitions to deep yellow (as in with a strong border between dark and light).

This contrasts with *Go officeri* which has a dorsum that is greyish in colour with a slight purplish tinge, mainly dark upper labials without obvious barring, dark etching on head shields which are not immaculate in that they have dark markings on the scales separate to the etchings and a gradual transition from dark to whitish on the lower flanks

Go punctatovittata is separated from the two preceding species by the combination of a pinkish orange dorsum (different to *Go forit sp. nov*. and *Go punctatovittata*); dark etching on the head shields that are otherwise immaculate in colour (in line with *Go forit sp. nov*.), barred labials that are more light than dark (similar to what is seen in *Go forit sp. nov*.), a gradual transition on the lower flank from dark to light yellow (in contrast to *Go forit sp. nov*. and *Go punctatovittata*).

The other two species in the genus *Go* Hoser, 2024 are separated from the preceding three by the following characters:

Go colliveri (Couper and Ingram, 1992) has a dorsum, flanks and tail that is medium brown in colour with a series of nine well-defined stripes running along the dorsum and flanks, these being formed by small blackish spots or alternatively fully merged to form distinctive lines. There is no light at all on the flanks and the upper labials are mainly dark. Dorsal head scales both etched and with dark markings on them.

Go emmotti (Ingram, Couper and Donnellan, 1993) has a dorsum, flanks and tail that is light brown in colour, poorly defined black spots or patches on scales on the dorsum or flanks, so that there are no obvious stripes running down the body or flank, although the lines can seen on close inspection, labials that are thickly barred dark, but with most of each labial white. Dorsal head shields are thinly, yet boldly etched dark, otherwise immaculate and either light brown or whitish in colour, being far lighter than the other upper surface scales on the lizard.

All the preceding named species within the genus *Go gen. nov.* are separated from all other species within *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, (in this case not *Go* Hoser, 2024) and *Get* Hoser, 2024) and assuming placement of all relevant species within

these genera, by the following unique combination of characters:

Relatively large and robust burrowing skink with an adult snout-vent to 100 mm. 4 limbs; forelimb is less than half the length of the hindlimb. Digits variable. Forelimb is stylar with 1, 2 fingers. 2 or 3 toes. Hindlimb is about 10 percent of snout-vent length.

Eyelid movable. Snout slightly protrusive. Pattern is absent, or consists of tiny dark spots aligned in rows, or joined to form longitudinal lines down the body. Lower flanks pale and the individual scales are edged dark brown. Belly is white or cream.

18 midbody rows; nasals broadly in contact; frontoparietals paired, not contacting and distinct from the interparietal; prefrontals are small and widely separated. Three supraoculars. Six supralabials.

Go forit sp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/107044466

Go officeri (McCoy, 1881) is depicted in life online at:

https://www.inaturalist.org/observations/81990734

and

https://www.inaturalist.org/observations/129115648

Go punctatovittata (Günther, 1867) is depicted in life in Wilson and Swan on page 381 on left, second from top and online at:

https://www.inaturalist.org/observations/259382349

and

https://www.inaturalist.org/observations/145822668

Distribution: The exact distribution of *Go forit sp. nov.* is uncertain, but believed to encompass the Flinders Ranges, north of the Spencer Gulf in South Australia, Australia, in particular the north-western facing side.

Etymology: In 2012, I was doing fieldwork at Leigh Creek with collaborator Tom Cotton.

This was after the local mine at Leigh Creek had contracted me to teach them how to handle venomous snakes.

I asked him to hit the brakes in the car he was driving so that I could flip a bit of tin on the side of the road in order to find a reptile.

He said, "*Go for it*".

Under the tin was a resting specimen of this species.

MICULIA CHRISMARTINI SP. NOV.

LSIDurn:lsid:zoobank.org:act:0B42144C-ABCE-4AB8-BEC8-E796167AF3D0

Holotype: A preserved adult male specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R168646 collected from Yetna, just north of Geraldton in Western Australia, Australia, Latitude -28.606667 S., Longitude 114.724722 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R130485 and R143520 both collected from the Bella Vista Nature Reserve, Western Australia, Australia, Latitude -28.531111 S., Longitude 114.738056 E.

2/ A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R130459 collected from the Howatharra Nature Reserve, Western Australia, Australia, Latitude -28.545556 S., Longitude 114.665278 E.

Diagnosis: Until 2024, the putative species *Ablepharus distinguendus* Werner, 1910, with a type locality of Freemantle (Perth), Western Australia, mostly placed in the genus *Lerista* Bell, 1833 was treated as a single species found from the Eyre Peninsula, South Australia west across southern Australia to the west coast and north to just past Geraldton in Western Australia.

In line with Wells and Wellington (1984, 1985) and Wells (2012), Hoser (2024c) placed the relevant species in the genus *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845.

The phylogeny of Torkkola *et al.* (2026) again confirmed a genus-level divergence of more than 10 MYA for the relevant group of species.

Hoser (2024c) also described the population from the northern two thirds of the Eyre Peninsula of South Australia as a separate and divergent species, namely *M. ruficauda* Hoser, 2024.

The published phylogeny of Torkkola *et al.* (2026) implied two other undescribed species in this complex, the first being the putative localised outlier population from the Geraldton district of the central west coast of Western Australia.

Because it is morphologically divergent, it is herein formally named as *Micula chrismartini sp. nov.*.

Another geographically separated and morphologically divergent population is found from about the Stokes National Park, which is roughly 70 km straight line west of Esperance, Western Australia, eastwards to about Madura on the western edge of the Nullarbor Plain proper, also in Western Australia, with outlier populations stretching inland and west to at least as far west as Hyden/Kondinin Cells (roughly 280 km in a straight line east of Perth).

That species is herein formally named as *Micula coldplayorum sp. nov.*.

The four relevant species, being *Miculia distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* are separated from one another by the following unique combinations of characters:

M. ruficauda Hoser, 2024 is readily separated from the other three species, being *M. distinguendus*, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.*

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by the fact that the black (in this species) dots on the scales that form the two lines running on either side of the vertebral line of the back are evenly spaced in forming the lines, versus not evenly spaced dots (black or brown) in the other three species. Furthermore, in *M. ruficauda* Hoser, 2024 the thick black line running along the upper flank is shrunken slightly as compared to the same line in *M. distinguendus*, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* with a consequence being a slight bleeding of brown, onto the white on the lower edge. The upper surfaces of the limbs are both dark and light in *M. ruficauda* Hoser, 2024 and *M. coldplayorum sp. nov.*, versus mainly dark in *M. distinguendus* and *M. chrismartini sp. nov.*.

M. ruficauda Hoser, 2024 is further separated from the other three species being *M. distinguendus*, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by the absence of a relatively bold and distinct lateral line on the anterior part of the tail.

M. distinguenda (Werner, 1910), is readily separated from the other three species, being *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by the combination of the following:

A dark chocolate or medium brown dorsum, strongly etched/barred upper labials and a thick jagged line on the upper lateral edge of the tail.

M. chrismartini sp. nov. is readily separated from the other three species, being *M. distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024 and *M. coldplayorum sp. nov.* by the combination of the following:

A relatively light, yellowish-brown dorsum and with tiny brown dots forming lines on the dorsum versus black dots in the other three species. There is a thick dark lateral stripe on the anterior tail that fades rapidly posteriorly.

Upper labials are thinly etched darker.

M. coldplayorum sp. nov. is readily separated from the other three species, being *M. distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024 and *M. chrismartini sp. nov.* by the combination of the following:

Weakly barred upper labials and a very thin line on the anterior upper lateral edge of the tail (versus absent in *M. ruficauda* Hoser, 2024 and thick in *M. distinguenda* (Werner, 1910) and *M. chrismartini sp. nov.*).

The four species *M. distinguendus*, *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

A small slender species with four digits on each limb, 6 supralabials; 3-4 supraciliaries and a fixed eyelid. They are separated from the closely related *Miculia elegans* Gray, 1845 species complex by having separated nasals (versus nasals joined to form a median suture) and more numerous midbody scale-rows (18 or more versus 16).

Distribution: *M. chrismartini sp. nov.* is essentially confined to the Geraldton area in Western Australia, as in within an 80 km radius, as is presently known. There is a gap of about 250 km before *M. distinguenda* (Werner, 1910) of the type form is found.

Etymology: The species is named in honour of Chris Martin, the front man for the rock band “Coldplay” in recognition of his services to the music industry globally as well as in outing narcissist marriage cheaters as he did to two narcissist marriage cheaters at a concert on Wednesday 16 July 2025.

During a Coldplay concert at Gillette Stadium at Foxborough, Massachusetts (greater Boston) USA, the venue’s JumboTron “Kiss Cam” panned to a couple cuddling and swaying intimately. When the

pair realized they were being broadcast to 65,000 fans, they panicked; the woman covered her face and the man ducked completely out of frame to hide.

From the stage, lead singer Chris Martin jokingly remarked, *"Either they're having an affair, or they're just very shy,"* unaware of how accurate his first guess was.

Internet sleuths quickly identified the couple as top executives from the billion-dollar New York tech startup, "Astronomer".

They were Andy Byron, the company's Chief Executive Officer (CEO), who was married and Kristin Cabot who was the company's Chief People Officer (CPO / Head of HR) who was also married. They were not married to each other!

Within days of the concert, the intense public backlash forced both executives to resign from their high-profile corporate roles.

Both lied in justifying their infidelity in their marriages.

In line with Narcissistic Personality Disorder (NPD) both were upset about being caught, but showed no remorse for doing the wrong thing by their partners. See relevant discussion at:

https://www.reddit.com/r/popculturechat/comments/1pptg2h/exec_caught_on_coldplay_kiss_cam_with_married/

MICULIA COLDPLAYORUM SP. NOV.

LSIDurn:lsid:zoobank.org:act:7D1B66E4-6AD7-45C4-BC3D-E804247FF89A

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R77776 collected from "in sand" 7 km north of Eyre Homestead, Western Australia, Australia, Latitude -32.184722 S., Longitude 126.301389 E.

This government-owned facility allows access to its specimens.

Paratypes: 1/ Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R93790 and R93791 both collected from under Eucalyptus litter 11 km east of Burnabbie Homestead, Western Australia, Australia, Latitude -32.122222 S., Longitude 126.456944 E., 2/ A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R91417 collected from leaf litter 2.5 km north of Eyre Homestead, Western Australia, Australia, Latitude -32.214444 S., Longitude 126.301667 E., 3/ A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R91418 collected from leaf litter 6 km north of Eyre Homestead, Western Australia, Australia, Latitude -32.177778 S., Longitude 126.300278 E., 4/ A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R34477 collected 24 km southeast of Cocklebiddy, Western Australia, Australia, Latitude -32.133333 S., Longitude 126.333333 E.

Diagnosis: Until 2024, the putative species *Ablepharus distinguendus* Werner, 1910, with a type locality of Freemantle (Perth), Western Australia, mostly placed in the genus *Lerista* Bell, 1833 was treated as a single species found from the Eyre Peninsula, South Australia west across southern Australia to the west coast and north to just past Geraldton in Western Australia.

In line with Wells and Wellington (1984, 1985) and Wells (2012), Hoser (2024c) placed the relevant species in the genus *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845.

The phylogeny of Torkkola *et al.* (2026) again confirmed a genus-level divergence of more than 10 MYA for the relevant group of species.

Hoser (2024c) also described the population from the northern two thirds of the Eyre Peninsula of South Australia as a separate and divergent species, namely *M. ruficauda* Hoser, 2024.

The published phylogeny of Torkkola *et al.* (2026) implied two other undescribed species in this complex, the first being the putative localised outlier population from the Geraldton district of the central west coast of Western Australia.

Because it is morphologically divergent, it is herein formally named as *Micula chrismartini sp. nov.*.

Another geographically separated and morphologically divergent population is found from about the Stokes National Park, which is roughly 70 km straight line west of Esperance, Western Australia, eastwards to about Madura on the western edge of the Nullarbor Plain proper, also in Western Australia, with outlier populations stretching inland and west to at least as far west as Hyden/Kondinin Cells (roughly 280 km in a straight line east of Perth).

That species is herein formally named as *Micula coldplayorum sp. nov.*.

The four relevant species, being *Miculia distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* are separated from one another by the following unique combinations of characters:

M. ruficauda Hoser, 2024 is readily separated from the other three species, being *M. distinguendus*, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by the fact that the black (in this species) dots on the scales that form the two lines running on either side of the vertebral line of the back are evenly spaced in forming the lines, versus not evenly spaced dots (black or brown) in the other three species. Furthermore, in *M. ruficauda* Hoser, 2024 the thick black line running along the upper flank is shrunken slightly as compared to the same line in

M. distinguendus, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* with a consequence being a slight bleeding of brown, onto the white on the lower edge. The upper surfaces of the limbs are both dark and light in *M. ruficauda* Hoser, 2024 and *M. coldplayorum sp. nov.*, versus mainly dark in *M. distinguendus* and *M. chrismartini sp. nov.*.

M. ruficauda Hoser, 2024 is further separated from the other three species being *M. distinguendus*, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by the absence of a relatively bold and distinct lateral line on the anterior part of the tail.

M. distinguenda (Werner, 1910), is readily separated from the other three species, being *M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* by the combination of the following:

A dark chocolate or medium brown dorsum, strongly etched/barred upper labials and a thick jagged line on the upper lateral edge of the tail.

M. chrismartini sp. nov. is readily separated from the other three species, being *M. distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024 and *M. coldplayorum sp. nov.* by the combination of the following:

A relatively light, yellowish-brown dorsum and with tiny brown dots forming lines on the dorsum versus black dots in the other three species. There is a thick dark lateral stripe on anterior tail that fades rapidly posteriorly.

Upper labials are thinly etched darker.

M. coldplayorum sp. nov. is readily separated from the other three species, being *M. distinguenda* (Werner, 1910), *M. ruficauda* Hoser, 2024 and *M. chrismartini sp. nov.* by the combination of the following:

Weakly barred upper labials and a very thin line on the anterior upper lateral edge of the tail (versus absent in *M. ruficauda* Hoser, 2024 and thick in *M. distinguenda* (Werner, 1910) and *M. chrismartini sp. nov.*).

The four species *M. distinguendus, M. ruficauda* Hoser, 2024, *M. chrismartini sp. nov.* and *M. coldplayorum sp. nov.* are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

A small slender species with four digits on each limb, 6 supralabials; 3-4 supraciliaries and a fixed eyelid. They are separated from the closely related *Miculia elegans* Gray, 1845 species complex by having separated nasals (versus nasals joined to form a median suture) and more numerous midbody scale-rows (18 or more versus 16).

Distribution: *M. coldplayorum sp. nov.* is found from about the Stokes National Park, which is roughly 70 km straight line west of Esperance, Western Australia, eastwards to about Madura on the western edge of the Nullarbor Plain proper, also in Western Australia, with outlier populations stretching inland and west to at least as far west as Hyden/Kondinin Cells (roughly 280 km in a straight line east of Perth).

Etymology: The species is named in honour of the internationally famous rock band “Coldplay” in recognition of their services to the music industry globally as well as in outing narcissist marriage cheaters as frontman Chris Martin did on Wednesday 16 July 2025 at a concert in Foxborough, Massachusetts (greater Boston) USA. See also the etymology for *M. chrismartini sp. nov.*.

MICULIA AERICEPS HOWZAT SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:63C270CD-C44D-4415-91E2-0B6C3BEA36CE

Holotype: A preserved specimen at the Museum and Art Gallery of the Northern Territory, Darwin, Northern Territory, Australia, Reptile Collection, specimen number R37825 collected from

20 km east of Frewena, south of the Barkly Highway, Barkly Tableland, Northern Territory, Australia, Latitude -19.74687 S., Longitude 135.45785 E.

This government-owned facility allows access to its holdings.

Diagnosis: The type form of *Micula aericeps* (Storr, 1986), type locality of 25 km south of Alice Springs, Northern Territory, Australia is a taxon found from northwest New South Wales, across northern South Australia, the Northern Territory and across a large swathe of Western Australia to the eastern edge of the Pilbara District. It is a distinctive lizard with an orange or orangish brown head, yellowish orange body, mainly yellow tail and immaculately white upper labials, that are sometimes barred or partially barred along the suture lines from the top.

Putative *M. aericeps* from the Barkly Tableland in the Northern Territory are so morphologically divergent that they appear to be of a different species. Were it not for the fact that I have seen specimens of apparently intermediate form (but more like *M. aericeps* in appearance) from Barrow Creek in the Northern Territory, I wouldn't have hesitated in naming the Barkly Tableland specimens as a separate species.

Instead, they are conservatively named herein as the subspecies *Micula aericeps howzat subsp. nov.*.

Micula aericeps howzat subsp. nov. are separated from nominate *M. aericeps* (Storr, 1986) by the following combination of characters: chocolate brown dorsum; heavy etching and markings on the upper head shields and obviously yellow upper labials, versus a dorsum that is either light brown, orangish brown or yellowish brown, with or without heavy etchings or markings on the upper head shields and with obviously white upper labials.

Nominate *Micula aericeps* and *Micula aericeps howzat subsp. nov.* are separated from all other species within the putative genus *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

A small slender skink with four fingers, four toes and a fixed eyelid in the form of a large transparent disc, with a coppery head, faintly spotted back and tail, and single preocular (rather than two). Has a yellow or yellowish tail and five supraciliaries. There is no bold upper lateral stripe although some specimens to have a faint one running along the mid and distal flank.

M. aericeps is depicted in life in Wilson and Swan on page 353 middle image.

M. aericeps howzat subsp. nov. is depicted in life online at:

https://www.flickr.com/photos/114192916@N07/54028843921/

and

https://www.flickr.com/photos/58349528@N02/51095265957/

and

https://www.flickr.com/photos/58349528@N02/51095171356/

and

https://www.inaturalist.org/observations/244648030

Distribution: *Micula aericeps howzat subsp. nov.* appears to be endemic to the Barkly Tableland in the Northern Territory, Australia.

Etymology: "*Howzat*" is a famous Australian slang and sporting term, primarily used in the game of cricket, that essentially translates to "How is that?", exclaimed when one does an incredible feat, which in this case includes grabbing one of these skinks as it tries to escape in the substrate when the conditions are hot.

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MICULIA PUSEYI SP. NOV.

LSIDurn:lsid:zoobank.org:act:D4DF2D2E-8750-41B0-9A2B-217109052ACD

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R127465 collected from 1.4 km east of Zanthus, Western Australia, Australia, Latitude -31.033333 S., Longitude 123.733333 E.

This facility allows access to its holdings.

Diagnosis: Until now, *Miculia puseyi sp. nov*. has been treated as a southern population of putative *Miculia taeniata* (Storr, 1986), a species that was originally placed by Storr in the genus *Lerista* Bell, 1833 and with a type locality of 20 km east of The Granites, Northern Territory, Australia, Latitude -20.34 S., Longitude 130.38 E.

Singhal *et al.* (2025) showed a molecular basis to separate the two morphologically divergent forms and so the unnamed one has been named herein.

Miculia taeniata (Storr, 1986) occupies much of central and southern Australia north of the Great Australian Bight and well away from the coast. At the southern extremity of this range in southern Australia, *Miculia puseyi sp. nov*. occurs.

Miculia puseyi sp. nov. is readily separated from *M. taeniata* by having yellowish upper labials with obvious dark triangles at the upper edge and a relatively thin dark upper lateral stripe, versus white upper labials without obvious dark triangles on the upper edge and a relatively thick dark upper lateral stripe.

Miculia puseyi sp. nov. and *M. taeniata* are separated from all other species within the putative genus *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

A small slender elongate skink with four fingers, four toes and a fixed eyelid in the form of a large transparent disc. A distinctively coppery coloured head, faintly spotted back and tail, which is broadly whitish grey or yellowish in colour. Single preocular. Yellow tail, five supraciliaries. A dark upper lateral stripe. A large preocular, 20 midbody rows, and whitish or yellowish tail.

Distribution: *Miculia taeniata* (Storr, 1986) occupies much of central and southern Australia north of the Great Australian Bight and well away from the coast. At the southern extremity of this range in southern Australia, *Miculia puseyi sp. nov*. occurs.

Etymology: *M. puseyi sp. nov*. is named in recognition of a formerly successful Mortgage Broker, named Richard Pusey who on 22 April 2020 lived in the inner Melbourne, Victoria, Australia suburb of Fitzroy and had a multi-million-dollar property portfolio.

By all accounts, the then 41-year-old was having a highly successful life.

Life as he knew it ended suddenly on 20 April 2020 when envious Victorian Police pulled him over while he was driving his Porche motorcar westbound on the Eastern Freeway near the Bulleen Road exit.

Rather than issuing him with an infringement notice (if he had in fact actually committed a crime) and then letting him drive off, the police decided to hold him up on the side of the road while they did "checks" on him.

This caused traffic chaos as drivers were forced to slow down in the normally 100 kmh zone when they saw the police lights flashing on the side of the road.

This was also on a blind corner!

About 45 minutes later and while the four Victoria police officers were simply wasting time by standing around in the traffic as they strutted around looking tough and intimidating, Pusey asked if he could urinate on the side of the road and out of public view.

While Pusey was urinating in the bushes, the Karma Bus in the form of a big truck driven by Mohinder

Singh ploughed through the police and the Porche killing all four Victorian Police.

Their names: Kevin King, Glen Humphris, Lynette Taylor and Joshua Prestney.

They should not be honoured in any way as they died as a direct result of their foolish and dangerous actions.

Significantly, Pusey had absolutely nothing to do with the truck driver Mohinder Singh running over the four foolish police playing in the traffic in the middle of a busy Melbourne freeway.

Rather than admit fault, the narcissistic Victorian Police blamed Pusey for the entire event and hit him with a bundle of criminal charges including "flee the scene of an accident".

Now as Pusey was not driving any vehicle at the time, the charge was ridiculous in the extreme.

That charge fell over, but the ever-vindictive police ultimately charged Pusey with countless falsified offences and thoroughly destroyed his life.

The saga was played out in the legacy media where he was vilified and demonized and a wholly false narrative about him was peddled as "fact".

Later on, in one of his numerous court appearances, Pusey quite correctly said that he'd have more hope of getting a fair trial in North Korea than in Victoria.

As for my interest in the matter, I should explain herein.

9 months earlier I was pulled up by two police at the same exact spot on the Eastern Freeway. The two police held me and my car on the side of the road for about an hour and I repeatedly asked them for me and/or them to move to a safer spot to which one of them replied "*don't tell us what to do*".

They spent most of that time yelling and screaming at me for publishing a book on police corruption in 1999 and how "*there is no corruption in the Victorian Police*" and "*you are a piece of shit*".

Luckily, I survived the encounter (fine free) as did the police, but the message is simple.

Victorian Police may be above the law and untouchable in a legal sense, but the laws of physics and Karma will eventually get those who transgress.

As a final PS in this story, the police who pulled me over posted me a fine for using a mobile phone while driving a month after the interception.

I had recorded everything of relevance and 1/ I had not used a mobile phone 2/ I could prove it 3/ They had never even alleged that I had used a mobile phone and 4/ At the time I was pulled over one of the police (the one who was doing most of the talking) actually said "*We are not alleging you have committed any offence*" in response to my opening comment "*what have I done?*".

I challenged the fine in court and after the police had lied about all the relevant facts I produced a recording that proved my case.

The corrupt female magistrate ignored the fact that I had proved my innocence and instead took umbrage at the fact that I had covertly taped the police.

This fact made me wholly dishonest and needing to be punished. That was her narrative anyway!

The outcome was that she convicted me of the offence, not because I had actually done it, but as punishment for covertly recording the police.

I had my driving license cancelled for a year.

I appealed the matter to the County Court, the next court up and this time around the female judge stuck to the case and agreed that I was innocent of the false claim of using a mobile phone while driving.

Charge dismissed.

Did I get any compensation for losing my driving license for what ended up being 6 months?

Of course not!

MICULIA VACOPOULOSAE SP. NOV.

LSIDurn:lsid:zoobank.org:act:BC7BA523-BC7F-4DA6-ACD6-014778B7955A

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R31012 collected from 7 km south of Prescott Point, Lake Eyre, South Australia, Australia, Latitude -29.03 S., Longitude 137.7 E. This government-owned facility allows access to its holdings.

Diagnosis: Until now *M. vacopoulosae sp. nov.* has been variously treated as a southeastern population of either *Miculia xanthura* (Storr, 1976), type locality of 7 km southwest of the Pollock Hills, eastern Western Australia, Australia, Latitude -22.54 S., Longitude 127.30 E. or *Miculia aericeps* (Storr, 1986) with a type locality of 25 km south of Alice Springs, Northern Territory, Australia.

Specimens of *M. vacopoulosae sp. nov.* lodged in Australian museums and posted online have been lodged and posted as both other species in an obvious case of misidentification of a hitherto unnamed species.

M. vacopoulosae sp. nov. is readily separated from the other two species *M. xanthura* or *M. aericeps* by the following combination of characters: Tail is usually not yellow, yellowish or with a yellow tinge, upper surfaces of head, body and anterior tail are of similar colour (usually brown) versus starkly contrasting head and tail as compared to the body in the other two species; upper labials are white, versus yellow or brown in the other two species; along the upper flank in particular a row of black spots on each scale forms a semi-distinct stripe (sometimes also seen in *M. aericeps* but never in *M. xanthura*); shields on top of the head are brown with dark spots or markings on them and rarely with any etching, versus etched in *M. aericeps* and usually neither in

M. xanthura.

M. vacopoulosae sp. nov. is also a slightly stockier animal than the other two species, which are more elongate and slender, especially in the case of *M. xanthura.*

The morphologically similar *Miculia taeniata* (Storr, 1986), *M. puseyi sp. nov*. and other species in the genus *Miculia* Gray, 1845 are separated from the preceding species (*M. vacopoulosae sp. nov.*, *Miculia xanthura* (Storr, 1976) and *Miculia aericeps* (Storr, 1986)) by presence of an obvious dark upper lateral stripe, large preocular and 20 midbody scale rows, versus 18 in the other species.

M. vacopoulosae sp. nov. is depicted in life online at: https://www.flickr.com/photos/120497805@N06/51203368781/

M. xanthura is depicted in life online at: https://www.flickr.com/photos/133201236@N03/18125934361/

M. aericeps is depicted in life in Wilson and Swan (2021) on page 353 middle image and online at:

https://www.flickr.com/photos/julesfarquhar/37330091550/

and

https://www.flickr.com/photos/zimny_anders/50384667207/

Distribution: *M. vacopoulosae sp. nov.* occurs in central east South Australia and nearby far west New South Wales.

Etymology: *M. vacopoulosae sp. nov*. is named in honour of Calista Vacopoulos of Marsden Park (Sydney), New South Wales, of "*Medusa Snake Catcher*" for her services for public safety in keeping people safe from venomous snakes by safely relocating them.

Yes, she lost a finger about a month after being chomped by a Red-bellied Black Snake *Pseudechis porphyriacus* (Shaw, 1794) in early 2026. The relatively small bite wound failed to heal properly after the bite and went seriously necrotic sometime after the original bite, requiring the need for amputation to stem the expanding bone infection.

PHANEROPIS ABSCONDEETUS SP. NOV.

LSIDurn:lsid:zoobank.org:act:7962EA33-69F4-43D5-ABCC-036CDBEF71A8

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R130800 collected from soil under a log, 6 km northeast of Sandfire Roadhouse, Western Australia, Australia, Latitude -19.733333 S., Longitude 121.116667 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia specimen number R106256 collected from Mandora Homestead, Western Australia, Australia, Latitude -19.733333 S., Longitude 120.85 E., 2/ Three preserved specimens at the Australian Museum, Sydney, New South Wales, Australia, specimen numbers R.172472, R.172477 and R.172478 all collected from the Eighty Mile Beach Caravan Park access road, 8.4 km North of the Great Northern Highway junction, Western Australia, Australia, Latitude -19.733333 S., Longitude 120.85 E.

Diagnosis: Since 2007, the putative species "*Lerista clara* Smith and Adams, 2007", herein placed in the genus *Phaneropis* Fischer, 1881 (type species *Phaneropis muelleri* Fischer, 1881) has been treated as a taxon occupying a distribution encompassing the coastal west Pilbara in a zone from just north of Carnarvon in the south to just north of Sandfire Flat in the north. However recent phylogenies published by Singhal *et al*. (2025) and Torkkola *et al.* (2026) showed two very divergent species level lineages within this population.

As they are morphologically divergent as well, the unnamed northern population is herein formally named as *P. abscondeetus sp. nov.*.

Phaneropis clara Smith and Adams 2007 with a type locality of Urala Station, Western Australia, Australia, Latitude -21.4722 S., Longitude 114.4204 E. appears to occupy the entire known range for the species save for a relatively small strip running from north of Port Hedland in the Pilbara District to just north of Sandfire Flat, all in Western Australia, Australia.

P. abscondeetus sp. nov. is readily separated from *P. clara* by the following characters:

The two paravertebral rows of black spots running down the dorsum are of significantly greater size and intensity than the other two rows of dots on the dorsum, versus all four rows of dots being of similar size and intensity in *P. clara.* The upper labials are white or whitish in *P. abscondeetus sp. nov.* versus yellowish in *P. clara.*

The usually irregularly shaped black spots on the upper surface of the head are large in *P. abscondeetus sp. nov.* versus medium or small and often quite scattered in *P. clara.*

The upper lateral stripe that runs from the body and along the sides of the tail does not extend to the end of the tail in *P. abscondeetus sp. nov.*, versus doing so or nearly doing so in *P. clara.*

P. abscondeetus sp. nov. and *P. clara* are separated from all other species in the genus *Phaneropis* Fischer, 1881 (type species *Phaneropis muelleri* Fischer, 1881) as defined by Hoser (2024c) by the following unique combination of characters:

Three fingers and toes; paired frontoparietals; five supraciliaries. Upper lateral stripe is broad, usually solid and straight edged; belly is pearly white and never pigmented (adapted from Smith and Adams

2007).

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal. Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger. One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line. Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

P. abscondeetus sp. nov. is depicted in life online at:

https://www.flickr.com/photos/julesfarquhar/52348668352/

and

https://www.flickr.com/photos/58349528@N02/52298287243/

P. clara is depicted in life in Wilson and Swan (2021) on page 361, left, middle and online at:

https://www.flickr.com/photos/euprepiosaur/54568550091/

and

https://www.flickr.com/photos/127392361@N04/54826866900/

Distribution: *P. abscondeetus sp. nov.* appears to confined to a coastal strip from north of Port Hedland in the Pilbara District to just north of Sandfire Flat, all in Western Australia, Australia.

Etymology: The Latin word "*absconditus*" means "*hidden*", and as this species had been hidden from science until now, the name is appropriate. However, the species name as spelt here "*abscondeetus*" is a deliberate choice and action taken to prevent any potential homonym with other names in "greater *Lerista*" including for example *Nodorha absconditus* Hoser, 2023.

Therefore, the spelling of "*abscondeetus*" in this species is intentional and it should not be changed. I note that the two relevant species are very divergent from one another and found on opposite sides of the Australian continent.

PHANEROPIS DORSEI SP. NOV.

LSIDurn:lsid:zoobank.org:act:6269D3B6-B3E3-4005-9947-252CE10009C1

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia specimen number R145071 collected from the Officer Basin area, Western Australia, Australia,

Australia, Latitude -27.052778 S., Longitude 125.155 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia specimen number R145076 collected from the Officer Basin area, Western Australia, Australia, Australia, Latitude -27.052778 S., Longitude 125.155 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Two other taxa, "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia have been generally treated as synonyms of "*Lerista timida*" by publishing herpetologists in recent years (postdating Cogger *et al.* 1983 or Cogger 2014).

Contrary to this position, Hoser (2024c), placed the relevant species into the genus *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 and recognised the east Australian species with the priority name *Phaneropis rhodonoides* (Lucas and Frost, 1896).

Molecular results of both Singhal *et al*. (2025) and Torkkola *et al.* (2026) have shown the Hoser (2024c) taxonomy and nomenclature to be robust and correct.

However, their results confirmed that the previously synonymised "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935" was divergent at the species level from all other named species-level taxa.

In addition to that, the phylogeny of Torkkola *et al.* (2026), also showed three other unnamed species within their putative "*Lerista timida*" including populations from south of Lake Eyre in South Australia, Northwest of Kalgoorlie, Western Australia, both as part of a larger mainly east Australian clade as well as another population from south-east of the Pilbara district in Western Australia.

A further sweep by myself of the various known populations of putative "*Lerista timida*" yielded five further unnamed populations.

Each are isolated by biogeographical barriers, usually in the form of extensive areas of periodically flooding black soil plains, or alternatively extensive mobile or stationary sand dunes.

The isolation of these populations is reflected in morphological divergence.

To make it clear, while species in this complex occupy a range of habitat types, they appear to be excluded from large areas of sand dunes and also periodically flooded black soil areas. This means for example that none occur in the lower Cooper's Creek drainage system (of eastern inland Australia) or the Simpson Desert proper.

Therefore, I have no hesitation in naming all 8 relevant populations as new species and caution also that there may yet be further unnamed species within this complex.

These 8 newly named species are as follows:

Phaneropis dorsei sp. nov. is the taxon of south-west Western Australia, generally west of the Nullarbor.

P. faaarkinelle sp. nov. is confined to a region north of the Nullarbor in South Australia in the south, bound by red dune country to the north, still in South Australia, extending south-east to the eastern edge of the Nullarbor.

P. wholeeecow sp. nov. is a species apparently confined to the Tomkinson Ranges and adjacent elevated and hilly areas in north-west South Australia and immediately adjacent parts of the Northern Territory and Western Australia. This area is in turn surrounded by mainly red dunes, flooding black or red soil areas or in the alternative salt pans.

P. overlookedit sp. nov. is confined to the Selwyn Ranges and Mount Isa area in northwest Queensland. This region is surrounded by black soil plains on three sides and the Gulf of Carpentaria on the north.

P. lopezi sp. nov. is a distinctive dark coloured form found in the area of the Northern Territory and South Australian border, south of the Amadeus basin and its south-eastern extension, and north of the low-lying plains between Coober Pedy in South Australia and the Northern Territory border. To the east and west, this species is constrained by significant areas of dunes.

P. greatfind sp. nov. is a relict form confined to the elevated parts of the Sturt National Park and environs in far northwest New South Wales, separated on all sides by flooding black or cracking soil lowlands.

P. laylawangae sp. nov. is a taxon found in the region occupying most of central South Australia, excluding the far south-east, far west, and far north.

P. gottabekidding sp. nov. is found in north-central Western Australia, east of the Pilbara district and west of the main Gibson Desert. It is closely related to the Pilbara species within the genus *Phaneropis*,

rather than the east Australian lineage, which includes the other species named herein.

Of relevance also is that *P. timida* of the type form is herein confined to most of New South Wales, west of the Great Dividing Range, excluding the far south-west and far northwest, as well as being found in most of south Queensland west of the Great Divide, bound in the west by the lowland flooding black soil areas of the Lake Eyre drainage system.

P. rhodonoides is found in the region of north-west Victoria, adjacent south-west New South Wales and adjacent parts of South Australia to the east of the Flinders Ranges.

P. goerlingi is a distinctive lighter coloured form found in central west Western Australia, generally near the coast, south of the Pilbara district.

The eleven preceding species are each separated from one another by the following unique combinations of characters:

P. timida of the type form has the following unique combination of characters:

Overall, it is a dark chocolate brown lizard on the top and sides. the top of the head is a similar colour to the body, although the anterior part of the top of the snout becomes a lighter greyish in colour. Most large head shields have dark brown spots on them, being either in the centre or to one side. The upper labials are white, with thick bold dark etching or bars over the sutures. Two paravertebral lines of dots on the scales are slightly more intense than those two rows to the sides of the dorsum. The dots forming these lines are slightly elongate (anterio-posteriorly). The dark upper lateral line is well defined at the top and poorly defined below. Below the lateral line, the scales are whitish with thick greyish brown edges, giving an overall impression of being a brownish white on the lower flanks. The lateral stripe extends most of the way along the (original) tail, although along most of the tail it is jagged edged and broken, being interfused with whitish areas.

See also for *P. rhodonoides* and *P. overlookedit sp. nov*. below.

P. timida is depicted in life online at:
https://www.inaturalist.org/observations/57992055
and
https://www.inaturalist.org/observations/259382344
and
https://www.inaturalist.org/observations/58824719

P. rhodonoides is similar in most respects to *P. timida* as just described, but separated from that taxon (and where relevant other species) by the following additional characters:

The spots on the dorsum are obviously elongate and also laterally expanded at the anterior edges. Dark on the upper labials is in the form of large dark spots and not barring or etching. The lateral line breaks rapidly as it enters the tail but still extends most of the way along the (original) tail.

The four rows of dots running down the dorsum of the body run onto and along about half the length of the original tail.

P. rhodonoides is depicted in life online at:
https://www.inaturalist.org/observations/165107494
and
https://www.inaturalist.org/observations/362997354

P. goerlingi is medium or even light brown on top. It has upper labials that have irregularly shaped dark triangles occupying the posterior, top corner of each, in front of which is an area of light yellow-orange in the form of patches and not a smudge or peppering of any sort, in front and below of which is some white (in the anterior lower corner).

The black scales of the upper lateral line have black anteriorly and orange posteriorly, giving a distinctive look. This carries onto the tail where the line becomes a broken blackish line with orange in between. On the mid and lower flank the scales are white, each with a reasonably sharp edged medium sized purplish brown spot at the centre of the posterior edge.

Each black spot forming rows down the dorsum are mainly squarish in shape.

On top of the head there is mainly marbling dark or peppering instead of immaculate brown with dark spots.

P. dorsei sp. nov. is light brown on top, with the posterior edges of each scale orange in colour. Rows of dark spots on the back are triangular in shape with the base on the anterior edge of each scale. Between the dark of the scales of the upper lateral band is orange. For the first row of scales below the lateral band the scales are white, each with a dark purple patch occupying the central lower edge of each scale, in total occupying about half of each scale. In lower rows this spot is reduced in size and intensity so that it does not occupy much of each scale at all and in some cases the scales are entirely white.

The upper labials anterior to the eye are barred blackish brown over the sutures, these joining at the upper edge of the scales, meaning the upper lateral line is well formed from snout to the eye and beyond there to tail.

On the head the scales are of similar colour to the body, with scattered small dark spots, flecks or peppering in an irregular manner. The lateral line remains reasonably well-formed for the entire length of the tail.

P. dorsei sp. nov. is depicted in life online at:
https://www.flickr.com/photos/wildlifeexperiences/19629134255/

P. gottabekidding sp. nov. is light brown on top and

with an upper lateral line that is poorly defined top and bottom. Along most of the flank of the body the dark on the upper lateral flank is reduced. That is, each scale is light brown anteriorly and dark posteriorly, meaning that the line is in fact broken on each scale. The scales in the two rows below this are white with mid-sized purple spots at the middle posterior edge. These spots are of the same size and intensity in each row.

The top of the snout is the same colour as the dorsum, including to tip and each scale has irregularly shaped dark spots, dashes or markings. The spots on the scales of the back (which are placed against the anterior suture) forming the lines running down the dorsum are small, rectangular, do not merge and are slightly wider at the distal end of each spot. The four rows of spots running down the back are all of similar size, shape and intensity.

All upper labials (including those posterior to the eye) are moderately barred purple at the posterior edge. The rear ones sometimes have purplish spots inside the white parts.

P. gottabekidding sp. nov. is depicted in life online at:

https://www.flickr.com/photos/toddburrows/6435824445/

and

https://www.flickr.com/photos/toddburrows/6435823241/

and

https://www.flickr.com/photos/brian_busho/16939952448/

P. faaarkinelle sp. nov. is readily separated from all the other species in the complex by the fact that below the lateral line on the flanks, the next two rows of scales are white, but with dark on each of the upper and lower edges, in effect forming a colour scheme of white on the mid and lower flank and with three distinct black lines overlaying this.

In addition, this species is a medium brown on top, the black dots on the dorsum nearly merge in forming lines, these lines commencing behind the eye, on the neck and running along the top of the body and anterior tail. The top of the head is greyish rather than brown and with scattered patches of orange on the scale edges and irregular small darkish spots in some but not all scales. The anterior upper labials have irregularly shaped bars on the sutures. The posterior to the eye upper labials are white with one or more dark spots of small size and irregular shape and not necessarily associated with the suture lines. The upper lateral line dissipates rapidly at the anterior (original) tail, instead becoming a semi distinct edge separating a brownish upper surface and yellowish sides.

P. wholeeecow sp. nov. is light copper brown on the upper surfaces of the head, the larger scales of which have small irregularly shaped dark markings or patches, slightly orangish on the dorsum of the body and yellowish on the upper and side parts of the tail.

This species is readily separated from all the others in the complex by the fact that the upper lateral stripe is well-defined blackish, edged light yellow on top and with a bold thin orange line as a boundary on the lower edge. Below this is thin purplish edge formed by an over thick etching of the upper edge of the adjoining scale rows.

The two scale rows below are etched purplish, but this etching is not as thick as the top boundary of the top row of these otherwise white scales. Anterior and posterior upper labials have thin bars formed by elongate blobs crossing the suture lines. In addition to this, the posterior labials also have small irregularly shaped purple spots in the white of each scale.

P. overlookedit sp. nov. is similar in most respects to *P. timida* as described above but differs in that the anterior upper labials are white and with thin etching along the suture lines only. Rear upper labials have none. The dorsum is noticeably lighter (yellowish) instead of brown at the outer (lateral) edge, versus only slightly lighter in *P. timida.* Upper surfaces and flanks of the tail are yellowish brown rather than a dark or medium brown as seen in *P. timida.*

P. lopezi sp. nov. is significantly different in appearance to all other species in the complex.

Dorsally and laterally the colour appears overwhelmingly to be a dark chocolate brown. The upper surface of the head is slightly coppery, with all or almost all scales having small dark irregularly shaped spots. This copper colour fades gradually on the neck and anterior body. The tiny black spots forming lines down the body are reduced in size and intensity and are barely noticeable along the length of most of the body. There is only the slightest of lightening on the outer edges of the dorsum and the black of the upper lateral stripe is reduced in amount and intensity, making the line itself barely noticeable in contrast to all the other species. There is no continuation of any lateral stripe onto the tail. The (original) tail rapidly becomes mainly a light yellowish in colour both above and on the sides. This is usually with the main colour becoming light and with a limited amount of brown spotting or peppering.

Anterior upper labials are mainly dark, save for small white crescents or patches at the central base of each. Posterior upper labials have dark patches, usually on the suture lines and are otherwise a combination of pink and white.

P. lopezi sp. nov. is depicted in life online at:

https://www.flickr.com/photos/58349528@N02/51102562606/

P. greatfind sp. nov. is another species that is of quite different appearance to the other species in the complex.

Overall, it is a darkish-brown lizard with poorly defined markings of any kind.

The anterior upper labials are yellowish white with only thin brown etching on the upper parts of each suture. Those below the eye and posterior to it are immaculate. The upper surface of the head and snout is a dull reddish purple in colour. The scales on the upper parts of the head have semi-distinct dark patches of irregular shape. The brownish dorsum has reduced spotting as compared to the other species in the complex. The dark spots on the dorsal scales are tiny, placed at the central anterior edge and only reach about a third of the way into each scale, making them relatively tiny as compared to other species in the group, where the spots may go half or sometimes the full length of each scale. In addition, in this species, where the dark lateral line is found each scale is simply brown and some, but not all have a small dark spot on the anterior edge.

The row of scales below this line are occupied by white scales that have medium sized black spots on the anterior edge, this being a barely distinct black line (over white) in this lizard.

The upper surfaces of the limbs in this lizard are unusual in that they are lightish brown as a base, but heavily superimposed with densely spaced uniform-coloured dark brown blotches or spots. The upper surfaces of the limbs in most other species in the complex are usually dark edged scales with orange centres.

P. greatfind sp. nov. is depicted in life online at: https://www.flickr.com/photos/120497805@N06/51203368781/

P. laylawangae sp. nov. is greyish rather than brownish on top, this alone making it an unusual and unique species in this group. The black spotting of the scales on the dorsum is bold, the spots are large, irregularly shaped, but often more-or-less rectangular and unlike in most if not all of the other species where they run from the anterior edge of each scale, in this species the spots are typically in the centre of each scale. The upper scales of the head are heavily etched black, but otherwise a light brownish grey at the centres. The dark patches forming the upper lateral stripe are more-or-less triangular in shape and join in the upper parts, forming a slightly jagged upper edge and very jagged lower edge, with a row of triangular intrusions of light at the lower edge.

The scales of the mid and lower flank are whitish and etched dark evenly on all sides. The (original) tail is pinkish grey in colour, the pink being faded and dull, but not bright or brilliant.

P. laylawangae sp. nov. is depicted in life online at: https://www.inaturalist.org/observations/229887615

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov*., *P. gottabekidding sp. nov*., *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length

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25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal. Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger. One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line. Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *Phaneropis dorsei sp. nov.* occurs in south-west Western Australia, generally west of the Nullarbor.

Etymology: Named in honour of Toowoomba, Queensland, Australia, wildlife demonstrator, Marc Dorse, of the business "Deadly Australians" for his services to wildlife conservation and education spanning some decades. He has also bred a number of rare and endangered species for the first time ever, including the Manning River Turtle *Wollumbinia purvisi* (Wells and Wellington, 1985), which is a unique endangered species that only exists in the wild state in the mid and upper reaches of the Manning River, New South Wales.

The genus name *Myuchelys* Thomson and Georges, 2009 is an illegally coined invalid junior objective synonym of *Wollumbinia* Wells, 2007, which has date priority over the latter name.

Thomson and Georges have engaged in narcissistic taxonomic vandalism by lying to others and promoting their name as the only available and correct name for the genus, while at all times being fully aware that *Wollumbinia* Wells, 2007 has date priority over their name.

Egregious taxonomic vandalism, with all the lies the action entails appears to be a good diagnostic trait for the Narcissistic Personality Disorder.

Thomson's NPD, shows in other traits he has in line with the disorder. This includes pathological lying, hijacking and rewriting narratives, blame shifting, gaslighting, deflection and theft. He also manipulates others to be his "flying monkeys".

Did I mention the other trait of NPD which is cheating in marriage?

Thomson abandoned his wife and disabled child in Australia to shack up with a very young girl in Brazil.

PHANEROPIS FAAARKINELLE SP. NOV.

LSIDurn:lsid:zoobank.org:act:BE09686E-6749-4938-B427-88F6E29BF213

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R60366 collected from 132.7 km west of Oak Valley, South Australia, Australia, Latitude -29.4992 S., Longitude 129.3319 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R70641 collected from 55.6 km northwest of Midgening Native Dam, South Australia, Australia, Latitude -29.0933 S., Longitude 129.6233 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. faaarkinelle sp. nov. is confined to a region north of the Nullarbor in South Australia in the south, bound by red dune country to the north, still in South Australia, extending south-east to the eastern edge of the Nullarbor.

P. faaarkinelle sp. nov. is separated from all the preceding species and also the closely related species "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

P. faaarkinelle sp. nov. is readily separated from all the other ten species in the complex by the fact that below the lateral line on the flanks, the next two

rows of scales are white, but with dark on each of the upper and lower edges, in effect forming a colour scheme of white on the mid and lower flank and with three distinct black lines overlaying this.

In addition, this species is a medium brown on top, the black dots on the dorsum nearly merge in forming lines, these lines commencing behind the eye, on the neck and running along the top of the body and anterior tail. The top of the head is greyish rather than brown and with scattered patches of orange on the scale edges and irregular small darkish spots in some but not all scales. The anterior upper labials have irregularly shaped bars on the sutures. The posterior to the eye upper labials are white with one or more dark spots of small size and irregular shape and not necessarily associated with the suture lines. The upper lateral line dissipates rapidly at the anterior (original) tail, instead becoming a semi distinct edge separating a brownish upper surface and yellowish sides.

Diagnostic characters used to separate the other ten preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov.*, *P. gottabekidding sp. nov.*, *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal.

Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger.

One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line.

Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *Phaneropis faaarkinelle sp. nov.* is confined to a region north of the Nullarbor in South Australia in the south, bound by red dune country to the north, still in South Australia, extending south-east to the eastern edge of the Nullarbor. On the west side, a particularly arid zone just over the border in Western Australia separates this species from *P. dorsei sp. nov.* which is the species taxon of south-west Western Australia, generally west of the Nullarbor.

Etymology: I asked an Aboriginal Pindiini native of the relevant region what was their tribe's local name for this species, or even if they had one, and he exclaimed "*faaarkinelle*". So, there you have it!

PHANEROPIS GOTTABEKIDDING SP. NOV.

LSIDurn:lsid:zoobank.org:act:68889788-46F4-4948-A59B-A9CFE0F87B42

Holotype: A preserved specimen at the Museum of Zoology, University of Michigan, Ann Arbor, Michigan, USA., specimen number UMMZ 244363 collected from Lorna Glen Station, Wiluna, Western Australia, Australia, Latitude -26.22637 S., Longitude 121.51882 E.

This facility allows access to its specimens.

Paratypes: Two preserved specimens at the Australian Museum, Sydney, New South Wales, Australia, specimen numbers R.100416 and R.100417 both collected from Wiluna Rubbish Tip, Wiluna, Western Australia, Australia, Latitude -26.6 S., Longitude 120.23333 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. gottabekidding sp. nov. is found in north-central Western Australia, east and south of the Pilbara district and west of the main Gibson Desert. It is closely related to the Pilbara species within the genus *Phaneropis*, rather than the east Australian lineage, which includes the other species named herein.

P. gottabekidding sp. nov. is separated from all the preceding species and also the closely related species "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

P. gottabekidding sp. nov. is light brown on top and with an upper lateral line that is poorly defined top and bottom. Along most of the flank of the body the dark on the upper lateral flank is reduced. That is, each scale is light brown anteriorly and dark posteriorly, meaning that the line is in fact broken on each scale. The scales in the two rows below this are white with mid-sized purple spots at the middle posterior edge. These spots are of the same size and intensity in each row.

The top of the snout is the same colour as the dorsum, including to tip and each scale has irregularly shaped dark spots, dashes or markings. The spots on the scales of the back (which are placed against the anterior suture) forming the lines running down the dorsum are small, rectangular, do not merge and are slightly wider at the distal end of each spot. The four rows of spots running down the back are all of similar size, shape and intensity.

All upper labials (including those posterior to the eye) are moderately barred purple at the posterior edge. The rear ones sometimes have purplish spots inside the white parts.

P. gottabekidding sp. nov. is depicted in life online at:

https://www.flickr.com/photos/toddburrows/6435824445/

and

https://www.flickr.com/photos/toddburrows/6435823241/

and

https://www.flickr.com/photos/brian_busho/16939952448/

Diagnostic characters used to separate the other ten preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov*., *P. gottabekidding sp. nov*., *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881

(as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal. Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger. One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line. Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *P. gottabekidding sp. nov*. is found in north-central Western Australia, east and south of the Pilbara district and west of the main Gibson Desert. It is closely related to the Pilbara species within the genus *Phaneropis*, rather than the east Australian lineage, which includes the other species named herein.

Etymology: When discussing this species as new with west Australian herpetologist Kaj-Erik Bulliard, he said to me “Gotta be kidding!”.

PHANEROPIS GREATFIND SP. NOV.

LSIDurn:lsid:zoobank.org:act:DF62BF24-8845-4EAE-85EE-0A65F40F7996

Holotype: A preserved specimen at the Australian Museum, Sydney, New South Wales, Australia, specimen number R.155035 collected from the Mount Wood Homestead, Sturt National Park, northwest New South Wales, Australia, Latitude -29.480278 S., Longitude 142.23361 E.

This government-owned facility allows access to its holdings.

Paratypes: 1/ A preserved specimen at the Australian Museum, Sydney, New South Wales, Australia, specimen number R.158872 collected from the Mount Wood Homestead, Sturt National Park, northwest New South Wales, Australia, Latitude -29.480278 S., Longitude 142.23361 E., 2/ Seven preserved specimens at the Australian Museum, Sydney, New South Wales, Australia, specimen numbers R.127080, R.127081, R.127082, R.127083, R.127084, R.127085 and R.127086 all collected from about 2.5 km east of Tibooburra, New South Wales, Australia, Latitude -29.433332 S., Longitude 142.05 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the

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widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. greatfind sp. nov. is a relict and divergent form confined to the elevated parts of the Sturt National Park and environs in far northwest New South Wales, separated on all sides by flooding black or cracking soil lowlands.

P. greatfind sp. nov. is separated from all the preceding species and also the closely related species "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

P. greatfind sp. nov. is another species that is of quite different appearance to the other species in the complex.

Overall, it is a darkish-brown lizard with poorly defined markings of any kind.

The anterior upper labials are yellowish white with only thin brown etching on the upper parts of each suture. Those below the eye and posterior to it are immaculate. The upper surface of the head and snout is a dull reddish purple in colour. The scales on the upper parts of the head have semi-distinct dark patches of irregular shape. The brownish dorsum has reduced spotting as compared to the other species in the complex. The dark spots on the dorsal scales are tiny, placed at the central anterior edge and only reach about a third of the way into each scale, making them relatively tiny as compared to other species in the group, where the spots may go half or sometimes the full length of each scale. In addition, in this species, where the dark lateral line is found each scale is simply brown and some, but not all have a small dark spot on the anterior edge.

The row of scales below this line are occupied by white scales that have medium sized black spots on the anterior edge, this being a barely distinct black line (over white) in this lizard.

The upper surfaces of the limbs in this lizard are unusual in that they are lightish brown as a base, but heavily superimposed with densely spaced uniform-coloured dark brown blotches or spots. The upper surfaces of the limbs in most other species in the complex are usually dark edged scales with orange centres.

P. greatfind sp. nov. is depicted in life online at: https://www.flickr.com/photos/120497805@N06/51203368781/

Diagnostic characters used to separate the other ten preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov*., *P. gottabekidding sp. nov*., *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected

by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal.

Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger.

One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line.

Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *P. greatfind sp. nov.* is a relict and divergent form confined to the elevated parts of the Sturt National Park and environs in far northwest New South Wales, separated on all sides by flooding black or cracking soil lowlands.

Etymology: When I first set eyes on a specimen of this species from the Sturt National Park in far northwest New South Wales I could see immediately that in terms of appearance, it was nothing like congeners to the east (as in *P. timida*), and so in terms of it being a new species, it was “a great find”.

PHANEROPIS LAYLAWANGAE SP. NOV.

LSIDurn:lsid:zoobank.org:act:61FB489D-0746-4DD9-BC55-7FFC33AE3C5E

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R51928 collected from 1.9 km south of Reedy Hole Springs Camp, South Australia, Australia, Latitude -30.2644 S., Longitude 138.8247 E. This government-owned facility allows access to its holdings.

Paratypes: Three preserved specimens at the South Australian Museum, Adelaide, South Australia, Australia, being specimen number R46479 a female collected from 2 km southeast of Beresford Rail Siding, South Australia, Australia, Latitude -29.2542 S., Longitude 136.6658 E., specimen number R28171 a male collected from Beresford Rail Siding. South Australia, Australia, Latitude -29.23 S., Longitude 136.6583 E., and specimen number R17059 collected from Beresford Hill, South Australia, Australia, Latitude -29.28 S., Longitude 136.68 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon “*Lerista timida* (De Vis, 1888)”, with a type locality of Charleville, Queensland, Australia or “*Lerista muelleri* (Fischer, 1881)”, with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. laylawangae sp. nov. is a taxon found in the region occupying most of central South Australia, excluding the far south-east, far west, and far north.

P. laylawangae sp. nov. is separated from all the preceding species and also the closely related species “*Ablepharus rhodonoides* Lucas and Frost, 1896” with a type locality of Mildura, Victoria and “*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935”, with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

P. laylawangae sp. nov. is greyish rather than brownish on top, this alone making it an unusual and unique species in this group. The black spotting of the scales on the dorsum is bold, the spots are large, irregularly shaped, but often more-or-less rectangular and unlike in most if not all of the other species where they run from the anterior edge of each scale, in this species the spots are typically in the centre of each scale. The upper scales of the head are heavily etched black, but otherwise a light brownish grey at the centres. The dark patches forming the upper lateral stripe are more-or-less triangular in shape and join in the upper parts, forming a slightly jagged upper edge and very jagged lower edge, with a row of triangular intrusions of light at the lower edge.

The scales of the mid and lower flank are whitish and

etched dark evenly on all sides. The (original) tail is pinkish grey in colour, the pink being faded and dull, but not bright or brilliant.

P. laylawangae sp. nov. is depicted in life online at: https://www.inaturalist.org/observations/229887615

Diagnostic characters used to separate the other ten preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov.*, *P. gottabekidding sp. nov.*, *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov.* are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal.

Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger.

One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line.

Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *P. laylawangae sp. nov*. is a taxon found in the region occupying most of central South Australia, excluding the far south-east, far west, and far north.

Etymology: *P. laylawangae sp. nov*. is named in honour of Layla Wang of Nanjing, China, spending a few years in Melbourne, Victoria, Australia working with Snakebusters: Australia's best reptiles, educating people about wildlife and conservation and also with our world-leading "Snake Avoidance" dog training business.

Details at http://www.dogsnakeavoidancetraining.com.au

PHANEROPIS LOPEZI SP. NOV.

LSIDurn:lsid:zoobank.org:act:3FB81E13-75C2-4246-AAC9-6D7619200449

Holotype: A preserved adult male specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R50193 collected from 14 km southeast of Sentinel Hill, South Australia, Australia, Latitude -26.1717 S., Longitude 132.5478 E.

This government-owned facility allows access to its holdings.

Paratypes: Two preserved specimens at the South Australian Museum, Adelaide, South Australia, Australia, specimen numbers R50188 and R50192 both collected from 14 km southeast of Sentinel Hill, South Australia, Australia, Latitude -26.1717 S., Longitude 132.5478 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. lopezi sp. nov. is a distinctive dark coloured form found in the area of the Northern Territory and South Australian border, south of the Amadeus basin and its south-eastern extension, and north of the low-lying plains between Coober Pedy in South Australia and the Northern Territory border. To the east and west, this species is constrained by significant areas of dunes.

P. lopezi sp. nov. is separated from all the preceding species and also the closely related species "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

P. lopezi sp. nov. is significantly different in appearance to all other species in the complex.

Dorsally and laterally the colour appears overwhelmingly to be a dark chocolate brown (only potentially matched in far southeastern Queensland specimens of *P. timida*). The upper surface of the head is slightly coppery (not seen in far southeastern Queensland specimens of *P. timida*), with all or almost all scales having small dark irregularly shaped spots. This copper colour fades gradually on the neck and anterior body. The tiny black spots forming lines down the body are reduced in size and intensity and are barely noticeable along the length of most of the body. There is only the slightest of lightening on the outer edges of the dorsum and the black of the upper lateral stripe is reduced in amount and intensity, making the line itself barely noticeable in contrast to all the other species. There is no continuation of any lateral stripe onto the tail. The (original) tail rapidly becomes mainly a light yellowish in colour both above and on the sides. This is usually with the main colour becoming light and with a limited amount of brown spotting or peppering.

Anterior upper labials are mainly dark, save for small white crescents or patches at the central base of each. Posterior upper labials have dark patches, usually on the suture lines and are otherwise a combination of pink and white.

Diagnostic characters used to separate the other ten preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov*., *P. gottabekidding sp. nov*., *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species,

Hoser 2026 - Australasian Journal of Herpetology 81-82:1-128.

Lerista vermicularis Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal. Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger. One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line. Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *Phaneropis lopezi sp. nov.* is a distinctive dark coloured form found in the area of the Northern Territory and South Australian border, south of the Amadeus basin and its south-eastern extension, and north of the low-lying plains between Coober Pedy in South Australia and the Northern Territory border. To the east and west, this species is constrained by significant areas of dunes.

Etymology: *Phaneropis lopezi sp. nov.* is named in honour of Oscar Lopez of the Ivanhoe area, Victoria, Australia, originally from Colombia (South America) for services to the community through his landscaping business and in government / politics.

PHANEROPIS OVERLOOKEDIT SP. NOV.

LSIDurn:lsid:zoobank.org:act:D592092E-97F0-4A70-A0F4-1765DF1BC475

Holotype: A preserved specimen at the Queensland Museum, Brisbane, Queensland, Australia, specimen number J32305 collected from Mount Isa, Queensland, Australia, Latitude -20.733333 S., Longitude 139.48333 E.

This government-owned facility allows access to its holdings.

Paratypes: Two preserved specimens at the Australian Museum, Sydney, New South Wales, Australia, specimen numbers R.15695 and R.26075 both collected from Mount Isa, Queensland, Australia, Latitude -20.733333 S., Longitude 139.48333 E.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. overlookedit sp. nov. is a species apparently confined to Selwyn Ranges area of north-west Queensland, Australia and is apparently not even common there.

P. timida of the type form is separated from all the preceding species and also the closely related species "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

P. timida of the type form has the following unique

combination of characters:

Overall, it is a dark chocolate brown lizard on the top and sides. the top of the head is a similar colour to the body, although the anterior part of the top of the snout becomes a lighter greyish in colour. Most large head shields have dark brown spots on them, being either in the centre or to one side. The upper labials are white, with thick bold dark etching or bars over the sutures. Two paravertebral lines of dots on the scales are slightly more intense than those two rows to the sides of the dorsum. The dots forming these lines are slightly elongate (anterio-posteriorly). The dark upper lateral line is well defined at the top and poorly defined below. Below the lateral line, the scales are whitish with thick greyish brown edges, giving an overall impression of being a brownish white on the lower flanks. The lateral stripe extends most of the way along the (original) tail, although along most of the tail it is jagged edged and broken, being interfused with whitish areas.

See also for *P. overlookedit sp. nov*. in the relevant description of that species in this paper.

P. timida is depicted in life online at:
https://www.inaturalist.org/observations/57992055
and
https://www.inaturalist.org/observations/259382344
and
https://www.inaturalist.org/observations/58824719

P. overlookedit sp. nov. is similar in most respects to *P. timida* as described above but differs in that the anterior upper labials are white and with thin etching along the suture lines only. Rear upper labials have none. The dorsum is noticeably lighter (yellowish) instead of brown at the outer (lateral) edge, versus only slightly lighter in *P. timida.* Upper surfaces and flanks of the tail are yellowish brown rather than a dark or medium brown as seen in *P. timida.*

Diagnostic characters used to separate the other nine preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding (full) description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov.*, *P. gottabekidding sp. nov.*, *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nicholsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal.

Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger.

One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line.

Hoser 2026 - Australasian Journal of Herpetology 81-82:1-128.

Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *P. overlookedit sp. nov*. is confined to the Selwyn Ranges / Mount Isa area in northwest Queensland. This region is surrounded by black soil plains on three sides and the Gulf of Carpentaria on the north.

Etymology: When discussing this unnamed species with Walloon (Ipswich) Queensland herpetologist Paul Woolf in 2025, he said to me "*over looked it*!" and hence the etymology.

PHANEROPIS OXYI SP. NOV.

LSIDurn:lsid:zoobank.org:act:A22A09B7-5A96-434B-8276-7F331BCF171C

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R162184 collected from 33 km west of Rhodes Ridge, Western Australia, Australia, Latitude -23.103056 S., Longitude 119.038889 E.

This government-owned facility allows access to its holdings.

Paratypes: Three preserved specimens at the Western Australian Museum, specimen numbers R117272, R117275 and R117340 all collected from Hope Downs, Western Australia, Australia, Latitude -22.973611 S., Longitude 119.118056 E.

Diagnosis: The concept of *Phaneropis muelleri* Fischer, 1881 (type species for that genus named at the same time) has changed significantly in recent years.

From 1983 to 2026, almost all authors have followed on from Cogger *et al*. (1983) by placing the relevant species within the better-known genus *Lerista* bell, 1833.

Wells and Wellington, 1985, shifted the relevant species to *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, but Wells (2012) shifted the relevant species to *Phaneropis* Fischer, 1881 based in particular on a recent molecular study published by Skinner *et al.* (2008).

Hoser (2024c) and this paper follow on from that position.

P. muelleri, was originally viewed as a pan-Australian species found across most of the continent, except for the extremities, especially in the north and south and far east.

Various studies and papers have basically cut the putative species up so that in most recent years, since the publication of Smith and Adams (2007) *P. muelleri* has been essentially confined to the Pilbara district of Western Australia.

However, the phylogeny of Torkkola *et al.* (2026) showed two very divergent species level lineages within this population. These populations were divided across the Fortescue River, a well-known biogeographical barrier of known antiquity. Furthermore, they are consistently morphologically divergent and so I have no hesitation in naming the unnamed southern form as a new species herein.

Phaneropis muelleri Fischer, 1881 has a type locality of Nichol Bay, Western Australia, Australia, (lectotype designation by Smith and Adams (2007)), being the part of the Pilbara District that is north of the Fortescue River.

P. oxy sp. nov. is the species from the main Pilbara District, south of the Fortescue River.

P. oxy sp. nov. is readily separated from *P. muelleri* by the following combination of characters:

P. oxy sp. nov. is mainly light under the eye (versus mainly black in *P. muelleri*); has yellowish at the distal end of tail (versus not so in *P. muelleri*). It is light brown on top versus dark chocolate brown, with a slight reddish tinge on top (in *P. muelleri*). The dark upper lateral stripe is well defined with clear borders in *P. oxy sp. nov.* versus not so in *P. muelleri*.

Notwithstanding sexual dimorphism in that females have proportionately longer bodies than males in both species, between the species, *P. oxy sp. nov*. is moderately elongated, versus significantly elongated in *P. muelleri*.

P. oxy sp. nov. and *P. muelleri* are separated from all other species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) by the following unique combination of characters:

Fingers and toes 3, Frontoparietals paired, Supraciliaries 1+2 (second supraciliary fused to first supraocular) (adopted from Smith and Adams 2007).

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species

Rhodona punctata Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal.

Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger.

One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line.

Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish-brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

P. oxy sp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/334339800

and

https://www.flickr.com/photos/114192916@N07/53770961363/

and

https://www.flickr.com/photos/zimny_anders/53676857712/

and

https://www.flickr.com/photos/reptileshots/53174347455/

and

https://www.flickr.com/photos/reptileshots/53174398298/

P. muelleri of the nominate form is depicted in life in Wilson and Swan (2021) on page 375 bottom left and online at:

https://www.flickr.com/photos/127392361@N04/53766186378/

and

https://www.flickr.com/photos/127392361@N04/53695676913/

and

https://www.flickr.com/photos/jaricornelis/51300503144/

Distribution: *Phaneropis oxy isp. nov.* is essentially a Pilbara endemic, from Western Australia, Australia. It is believed to occur in the region south and west of the Fortescue River in the north and the Ashburton River in the south, obviously bound by the Indian Ocean in the west.

The related and morphologically similar species *P. muelleri* appears to be restricted to the North Pilbara only, including nearby outlier hill areas on the southern edge of the Great Sandy Desert.

Etymology: *Phaneropis oxyi sp. nov.* is named in honour of the author's now deceased Great Dane dog, named "*Oxyuranus*" or "Oxy" for short, in recognition of his roughly 8 and a half years of protecting the author's facility from thieves from end 2003 to mid 2012.

PHANEROPIS WHOLEEECOW SP. NOV.

LSIDurn:lsid:zoobank.org:act:25860030-797E-4B68-9855-C2FFE73BC4A5

Holotype: A preserved specimen at the South Australian Museum, Adelaide, South Australia, Australia, specimen number R46144 collected 8 km northwest of Kunytjanu, South Australia, Australia, Latitude -26.615 S., Longitude 129.3139 E.

This government-owned facility allows access to its holdings.

Diagnosis: Until now, each of the following species, *Phaneropis dorsei sp. nov*., *P. faarkinelle sp. nov*. *P. greatfind sp. nov*., *P. gottabekidding sp. nov*., *P. laylawangae sp. nov*., *P. lopezi sp. nov*., *P. overlooked it sp. nov*., and *P. wholeeecowe sp. nov*. have all been treated as populations of the widespread putative taxon "*Lerista timida* (De Vis, 1888)", with a type locality of Charleville, Queensland, Australia or "*Lerista muelleri* (Fischer, 1881)", with a type locality of Nickol Bay, Western Australia, herein restricted to the Pilbara district, Western Australia, *sensu* Smith and Adams (2007) or perhaps more properly *sensu* Cogger (2014).

Refer to the preceding description of *Phaneropis dorsei sp. nov*. for further relevant information.

P. wholeeecow sp. nov. is a species apparently confined to the Tomkinson Ranges and adjacent elevated and hilly areas in north-west South Australia and immediately adjacent parts of the Northern Territory and Western Australia. This area is in turn surrounded by mainly red dunes or flooding black or red soil areas.

P. wholeeecow sp. nov. is separated from all the preceding species and also the closely related species "*Ablepharus rhodonoides* Lucas and Frost, 1896" with a type locality of Mildura, Victoria and "*Lygosoma* (*Rhodona*) *goerlingi* Ahl, 1935", with a type locality of Wurarga, Marloo Station (western) Western Australia, by the following unique combination of characters:

This species is readily separated from all the other ten species in the complex by the fact that the upper lateral stripe is well-defined blackish, edged light yellow on top and with a bold thin orange line as a boundary on the lower edge. Below this is thin purplish edge formed by an over thick etching of the upper edge of the adjoining scale rows.

The two scale rows below are etched purplish, but this etching is not as thick as the top boundary of the top row of these otherwise white scales. Anterior and posterior upper labials have thin bars formed by elongate blobs crossing the suture lines. In addition to this, the posterior labials also have small irregularly shaped purple spots in the white of each scale.

P. wholeeecow sp. nov. is light copper brown on the upper surfaces of the head, the larger scales of which have small irregularly shaped dark markings or patches, slightly orangish on the dorsum of the body and yellowish on the upper and side parts of the tail.

Diagnostic characters used to separate the other ten preceding listed species (all herein placed within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881) can be found in the preceding description of *P. dorsei sp. nov.* used herein explicitly as part of this scientific description for this taxon.

The previous eleven species being *P. timida*, *P. rhodonoides*, *P. goerlingi*, *P. dorsei sp. nov*., *P. gottabekidding sp. nov*., *P. faaarkinelle sp. nov.*, *P. wholeeecow sp. nov.*, *P. overlookedit sp. nov.*, *P. lopezi sp. nov.*, *P. greatfind sp. nov.* and *P. laylawangae sp. nov*. are separated from all other species within *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881 by the following unique combination of characters:

3 fingers and 3 toes; the frontoparietals are fused into a single shield; supraciliaries 1+2 (second supraciliary fused to the first supraocular); 20 midbody rows; margins of dorsals rows one and two are not dark grey.

Species within the genus *Phaneropis* Fischer, 1881 (as defined by Hoser 2024c and adopted herein) are separated from all other members of *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, (in this case excluding *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881), *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991,

Tychismia Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Very small to large semifossorial skinks (*P. verhmens* up to 50 mm) with elongate to moderately elongate bodies. Mean forelimb length to 13.9% (*P. occulta* and *P. verhmens)* of SVL and mean hindlimb length 25%-30% of SVL. Eyelid immovable (a spectacle). Digits 4+4,3+3,2+3. Midbody scale rows 18-22. Nasals large, rarely narrowly separated, usually in short to moderate contact. Prefrontals widely separated. Frontoparietals in long contact or fused. Supraoculars three, first two in contact with frontal.

Supraciliaries three, four or five in a complete or interrupted series. Loreals two, anterior the larger.

One or two presuboculars. Upper labials six. Nuchals zero to five on either side of vertebral line.

Primary temporals one; secondary temporals two, upper much the larger and overlapped by the lower.

Dorsum usually olive brown, brown or creamy white. Usually, pattern on the first two rows of dorsal scales, absent or reduced to a dot, dash or oblong on each scale. Pale whitish dorsolateral stripe moderately strong, weak or absent. Blackish brown upper lateral stripe completely solid, solid at its upper edge, fragmented at lower edge. Lower flanks and venter spotted with blackish brown of various intensity to immaculate pearly white (adopted and modified from Smith and Adams 2007).

The morphologically similar genus *Spectrascincus* Wells, 2012 are separated from the genus *Phaneropis* Fischer, 1881 by having a mean hindlimb length of 32-35% versus a mean hindlimb length 25%-30% of SVL in *Phaneropis* Fischer, 1881.

Distribution: *Phaneropis wholeeecow sp. nov.* is a species apparently confined to the Tomkinson Ranges and adjacent elevated and hilly areas in north-west South Australia and immediately adjacent parts of the Northern Territory and Western Australia. This area is in turn surrounded by mainly red dunes, flooding black or red soil areas and salt pans.

Etymology: In early 1983, I was discussing with my travel partner Charles Acheson the idea of driving from Marla Bore on the NT/SA border for about a day to get to Tomkinson Ranges to collect this species and other obscure reptile taxa. Charles gave the following response to my idea: "*wholeeecow*".

RHODONA CONNIVENS NANDA SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:2800BC29-F3FF-45C1-8698-EC7CDF26BAAE

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R22897 collected from Kalbarri, Western Australia, Australia, Latitude -27.72 S., Longitude 114.17 E. This government-owned facility allows access to its holdings.

Paratypes: Five preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R22902, R34317, R40286, R41731 and R43857 all collected from Kalbarri, Western Australia, Australia, Latitude -27.72 S., Longitude 114.17 E.

Diagnosis: Until now, no subspecies of *Rhodona connivens* (Storr, 1971) have been recognised.

Singhal *et al*. (2025) in their supplementary data recognised three so-called "Operational species" within putative "*Lerista connivens*" of west Western Australia, Australia.

These were the nominate subspecies from the immediate vicinity of Shark Bay, including islands and the type locality of Salutation Island, Western Australia, Latitude 26.32 S., Longitude 113.46 E. (Holotype WAM R25777), a form from the coast further north in the area north of Carnarvon and generally south of the Cape Range, and a population to the south found in a zone extending from Billabong Roadhouse to Kalbarri in Western Australia.

As these forms are morphologically separable, they are herein formally named as new subspecies.

They are separated from one another by the following character combinations:

The type form of *Rhodona connivens* has two very dark brown paravertebral lines running from the back of the head onto the tail. These lines are formed by large dots that tend to merge with one another on the upper and lower edge. Inside these lines is a ragged edged vertebral line, usually brown in colour. On the other edge of the dorsum the colour is light brown or yellow. There is dark marbling on the upper surfaces of the head. On the top of the original tail of this subspecies the colour of the vertebral line is obviously much darker (usually grey or greyish) versus that outside the paravertebral lines (yellow or yellowish).

Type *R. connivens* is depicted in life in Wilson and Swan (2021) on page 361 bottom left, Cogger (2014) on page 600 at top right and online at:

https://www.flickr.com/photos/colonel_007/52793982981/

and

https://www.flickr.com/photos/colonel_007/49112257816/

R. connivens yinggarda subsp. nov. is readily

separated from the other two subspecies by the fact that the vertebral area between the paravertebral lines is the same colour as the lines on the outer edge of the dorsum (versus contrasting in the other two subspecies).

In this subspecies it is light brown or yellowish for both. The size of the paravertebral spots is also reduced as compared to the other two subspecies, these merely being tiny dots in this subspecies. This reduction is such that on the tail, one merely sees very fine lines, versus distinct and thick in the other two subspecies, meaning that to all intents and purposes the upper part of the tail is unicolour save for the fine longitudinal lines.

R. connivens nanda subsp. nov. is a much darker coloured lizard than the other two subspecies. The vertebral line is dark, invariably with a slight reddish tinge (not seen in the other two subspecies), sometimes being nearly black in colour. On the top of the original tail of this subspecies the colour of the vertebral line is the same as that outside the paravertebral lines.

R. connivens nanda subsp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/341633026

and

https://www.flickr.com/photos/114192916@N07/54101386792/

The three preceding subspecies are otherwise similar in form to species within the genus *Telchinoscincus* Wells and Wellington, 1984 (also common in this part of the world) as defined by Hoser (2024c) save for their movable eyelid, vertebral line that is ragged edged as opposed to smooth and larger size.

The three preceding species are separated from other species within *Lerista sensu lato sensu* Hoser (2024c) or Cogger (2014) by the following combination of characters:

Usually no trace of a forelimb, rarely a tubercle. Maximum snout-vent length is usually about 72 mm.

Length of appendages (as % SVL): hind-leg 12.6-15.2; snout to forelimb groove 21.5-23.4. Nasals in moderately long contact. Prefrontals widely separated. Frontoparietals and interparietal fused into a single shield. 0-2 nuchals. 3 supraoculars 3, first two in contact with the frontal. 3-4 supraciliaries 6 upper labials. Upper secondary temporal is much the largest, lower secondary is much the smallest. 20 or 22 midbody scale-rows. 9-12 lamellae under the longer toe 9-12. Dorsally white or greyish-white, sometimes, yellow, brown or beige, with a wide ragged edged vertebral stripe in most specimens in most locations (modified from Storr, 1971).

Distribution: The subspecies *R. connivens nanda subsp. nov.* is found in a zone from Billabong Roadhouse to Kalbarri in Western Australia. *R. connivens yinggarda subsp. nov*. is the taxon found from Carnarvon in the south to Cape Range in the north, of Western Australia. Around and in Shark Bay, Western Australia one finds nominate *R. connivens*.

Etymology: The subspecies name comes from the Nanda Aboriginal people who were the original inhabitants of the area this taxon occurs. Most were either exterminated by the British invaders in the 1800's, died from European diseases, or fled to remote places. For most of the past 200 years the survivors have scavenged for scraps and waste on the outskirts of towns, while living under sheets of tin on land with zero economic value.

Not much has changed in recent years save for a "Woke" Aboriginal Industry that has emerged.

This business is largely controlled by so called "hobby coons", this being non-Aboriginal people who "identify" as Aboriginals.

They are then faking it as charities and scamming money from governments and gullible private donors to ostensibly spend on Aboriginal welfare.

Invariably the cash is siphoned off and the Aboriginals themselves get little if any money or benefits.

The spelling of the name "nanda" is intentional and should not be amended to include "ourum" or similar as would normally be the case in correct Latinisation.

RHODONA CONNIVENS YINGGARDA SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:92BECAD8-E38D-4DED-B191-C212CFC9E599

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R76713 collected 9 km south-east of Gnaraloo Homestead, Western Australia, Australia, Latitude -23.9 S., Longitude 113.55 E.

This government-owned facility allows access to its holdings.

Paratypes: Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R76714 and R76715 collected 9 km south-east of Gnaraloo Homestead, Western Australia, Australia, Latitude -23.9 S., Longitude 113.55 E.

Diagnosis: Until now, no subspecies of *Rhodona connivens* (Storr, 1971) have been recognised.

Singhal *et al*. (2025) in their supplementary data recognised three so-called "Operational species" within putative "*Lerista connivens*" of west Western Australia, Australia.

These were the nominate subspecies from the immediate vicinity of Shark Bay, including islands and the type locality of Salutation Island, Western Australia, Latitude 26.32 S., Longitude 113.46 E. (Holotype WAM R25777), a form from the coast

further north in the area north of Carnarvon and generally south of the Cape Range, and a population to the south found in a zone extending from Billabong Roadhouse to Kalbarri in Western Australia.

As these forms are morphologically separable, they are herein formally named as new subspecies.

They are separated from one another by the following character combinations:

The type form of *Rhodona connivens* has two very dark brown paravertebral lines running from the back of the head onto the tail. These lines are formed by large dots that tend to merge with one another on the upper and lower edge. Inside these lines is a ragged edged vertebral line, usually brown in colour. On the other edge of the dorsum the colour is light brown or yellow. There is dark marbling on the upper surfaces of the head. On the top of the original tail of this subspecies the colour of the vertebral line is obviously much darker (usually grey or greyish) versus that outside the paravertebral lines (yellow or yellowish).

Type *R. connivens* is depicted in life in Wilson and Swan (2021) on page 361 bottom left, Cogger (2014) on page 600 at top right and online at:

https://www.flickr.com/photos/colonel_007/52793982981/

and

https://www.flickr.com/photos/colonel_007/49112257816/

R. connivens yinggarda subsp. nov. is readily separated from the other two subspecies by the fact that the vertebral area between the paravertebral lines is the same colour as the lines on the outer edge of the dorsum (versus contrasting in the other two subspecies).

In this subspecies it is light brown or yellowish for both. The size of the paravertebral spots is also reduced as compared to the other two subspecies, these merely being tiny dots in this subspecies. This reduction is such that on the tail, one merely sees very fine lines, versus distinct and thick in the other two subspecies, meaning that to all intents and purposes the upper part of the tail is unicolour save for the fine longitudinal lines.

R. connivens nanda subsp. nov. is a much darker coloured lizard than the other two subspecies. The vertebral line is dark, invariably with a slight reddish tinge (not seen in the other two subspecies), sometimes being nearly black in colour. On the top of the original tail of this subspecies the colour of the vertebral line is the same as that outside the paravertebral lines.

R. connivens nanda subsp. nov. is depicted in life online at:

https://www.inaturalist.org/observations/341633026

and

https://www.flickr.com/photos/114192916@N07/54101386792/

The three preceding subspecies are otherwise similar in form to species within the genus *Telchinoscincus* Wells and Wellington, 1984 (also common in this part of the world) as defined by Hoser (2024c) save for their movable eyelid, vertebral line that is ragged edged as opposed to smooth and larger size.

The three preceding species are separated from other species within *Lerista sensu lato sensu* Hoser (2024c) or Cogger (2014) by the following combination of characters:

Usually no trace of a forelimb, rarely a tubercle. Maximum snout-vent length is usually about 72 mm.

Length of appendages (as % SVL): hind-leg 12.6-15.2; snout to forelimb groove 21.5-23.4. Nasals in moderately long contact. Prefrontals widely separated. Frontoparietals and interparietal fused into a single shield. 0-2 nuchals. 3 supraoculars 3, first two in contact with the frontal. 3-4 supraciliaries 6 upper labials. Upper secondary temporal is much the largest, lower secondary is much the smallest. 20 or 22 midbody scale-rows. 9-12 lamellae under the longer toe 9-12. Dorsally white or greyish-white, sometimes, yellow, brown or beige, with a wide ragged edged vertebral stripe in most specimens in most locations (modified from Storr, 1971).

Distribution: *R. connivens yinggarda subsp. nov*. is the taxon found from Carnarvon in the south to Cape Range in the north, of Western Australia. Around and in Shark Bay, Western Australia one finds nominate *R. connivens*. The subspecies *R. connivens nanda subsp. nov.* is found in a zone from Billabong Roadhouse to Kalbarri in Western Australia.

Etymology: The subspecies name comes from the Yinggarda Aboriginal people who were the original inhabitants of the area this taxon occurs. Most were either exterminated by the British invaders in the 1800's, died from European diseases, or fled to remote places. For most of the past 200 years the survivors have scavenged for scraps and waste on the outskirts of towns, while living under sheets of tin on land with zero economic value.

The spelling of the name is intentional and should not be amended to include "ourum" or similar as would normally be the case in correct Latinisation.

TELCHINOSCINCUS CALISTACLEMENTSAE SP. NOV.

LSIDurn:lsid:zoobank.org:act:B0A54CFC-2215-480D-8FE7-43F4A93337B5

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R54208 collected from Tallering, Western Australia, Australia, Latitude -26.27 S., Longitude 115.25 E. This government-owned facility allows access to its holdings.

Paratypes: Six preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R87845, R87846, R87847, R87848, R87849 and R87850 all collected from Nilung Pool, Western Australia, Australia, Latitude -25.7 S., Longitude 116.0 E.

Diagnosis: Until now, *Telchinoscincus calistaclementsae sp. nov.* has been treated as a divergent northern population of the better known *Telchinoscincus nichollsi* (Loveridge, 1933) *sensu* Hoser (2024c) and previously placed by most earlier authors in the greater genus *Lerista* Bell, 1833, but notably excluding Wells and Wellington (1984) who first erected the genus *Telchinoscincus*.

T. calistaclementsae sp. nov. is separated from *T. nichollsi* by the presence of a brownish vertebral line that is thicker than the bold and distinct yellow lines on either side of this line on the lateral parts of the dorsum. Furthermore, the colour intensity of the vertebral line in *T. calistaclementsae sp. nov.* is somewhat faded, versus bold in *T. nichollsi.*

In addition the outer edges of the brownish vertebral line in *T. calistaclementsae sp. nov.* is noticeably darker than the central parts, versus not so in *T. nichollsi.*

Both *T. calistaclementsae sp. nov.* and *T. nichollsi* are readily separated from the closely related and morphologically similar species *T. petersoni* (Storr, 1976) by having one loreal scale (as opposed to two), a smaller adult size (average 60 mm, vs 70 mm), and a vertebral stripe well separated from the dark brown lines of the lateral edge and flank, versus not so in *T. petersoni.*

The related species *T. gascoynensis* (Storr, 1986), is separated from *T. calistaclementsae sp. nov.* and *T. nichollsi* by its greater size (average snout-vent 70 mm, vs 60 mm) and having 3 not 4 supraciliaries.

T. kendricki (Storr, 1991) is separated from *T. calistaclementsae sp. nov.* and *T. nichollsi* by its lesser size (not more than 50 mm snout-vent), relatively shorter tail, larger second loreal, larger presuboculars, darker head, wider vertebral stripe and narrower upper lateral stripe.

T. kennedyensis (Kendrick, 1989) is separated from the preceding species by having a pale yellowish-brown dorsum with two rows of dark dots down either side of the vertebral line, narrow brown band on the upper flank, dark brown stripes down the top and sides of the tail on an otherwise light grey background (original tail).

T. onsloviana (Storr, 1984) is similar in appearance to the preceding species, but instead with a thick dark brown band on the upper flank, the dots running either side of the vertebral line forming a barrier and boundary to a darker brown vertebral line, which is sometimes split over, extending on to a yellowish (original) tail with brown stripes on either side.

It is further defined by the following characters:

Usually 2 supraciliaries (the first and second being fused to first supraocular), second loreal fused to prefrontal, and relatively narrow vertebral stripe (enclosing 2 series of dark spots) (modified from Storr 1984).

T. uniduo (Storr, 1974) is separated from the preceding taxa by having a greyish-brown vertebral line, with ill-defined outer edges, with the outer edges of the dorsum being beige, a bold thick, blackish brown stripe along the top half of the flank, bound by bold yellow on the lower half, four supraciliaries and a snout vent in adults of 55 mm.

T. yuna (Storr, 1991) is the only member of the genus *Telchinoscincus* with four lines of dots running the length of the dorsum on the body. Brownish on top, except for the outer edges of the dorsum which is greyish. Most of the upper flank is black, with only the lower part yellow and bound by a well-defined edge. Tail (original) is spotted, tending to form stripes and brownish grey overall on top, brown on the sides and yellow underneath. Heavily barred upper labials.

The preceding species, forming the entirety of the genus *Telchinoscincus* (Wells and Wellington, 1984) are separated from their closest related genus *Rhodona* Gray, 1839, by having a fixed, fused unmovable lower eyelid, versus a moveable one in *Rhodona.*

T. calistaclementsae sp. nov. is depicted in life online at:

https://www.flickr.com/photos/reptileshots/14930185189/

from Woodleigh, Western Australia, Latitude 26.2081 S., Longitude 114.8771 E.

T nichollsi is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/54019853019/

from Yalgoo, Western Australia, Australia, Latitude 28.3400 S., Longitude 116.6829 E.

Distribution: *Telchinoscincus calistaclementsae sp. nov.* is a relatively restricted taxon found within 200 km of the Western Australian coastline in the arid area mainly to the south of the Wooramei River and north of the Murchison River. South of the Murchison River one finds the closely related taxon *T. nichollsi* (Loveridge, 1933) and in the region bounded by the Wooramei River and Gascoyne River to the north one finds the closely related taxon *T. gascoynensis* (Storr, 1986). All other species in the genus *Telchinoscincus* are concentrated within the same general region of Western Australia.

Etymology: *T. calistaclementsae sp. nov.* is named in honour of Calista Clements who has made a name for herself exposing deep seated corruption in Victoria, Australia by creating videos and posting them on publishing platforms like “Facebook”.

TELCHINOSCINCUS KENDRICKI KALBARRIENSIS SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:E92EE6FF-F95F-464D-92F1-B1D7A262DF9E

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R37640 collected from Kalbarri, Western Australia, Australia, Latitude -27.71 S., Longitude 114.17 E.

This government-owned facility allows access to its holdings.

Paratypes: Two preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R37641 and R22900 collected from Kalbarri, Western Australia, Australia, Latitude -27.71 S., Longitude 114.17 E.

Diagnosis: *Telchinoscincus kendricki kalbarriensis subsp. nov.* is separated from the nominate form of *Telchinoscincus kendricki* (Storr, 1991) by having a grey-brown, versus chocolate brown vertebral line, and barely yellow on the lower flank, versus strongly yellow on the lower flank (adult specimens). Speckling on the head of *T. kendricki kalbarriensis subsp. nov.* is well defined, versus poorly defined in nominate *T. kendricki kendricki.*

Both the preceding taxa are separated from all other species in the genus *Telchinoscincus* Wells and Wellington, 1984 by the following unique combination of characters:

A moderately small, strongly patterned member of the genus *Telchinoscincus* (growing up to 118 mm long total length) with immovable eyelids (diagnostic for genus), 4 supraciliaries, wide vertebral stripe, two toes and forelimb represented by a small pit or groove (occasionally with a minute stump up to 0.2 mm long).

Distinguishable from the *T. nicholsi* and closely associated taxa by lesser size, relatively shorter tail, larger second loreal, larger presuboculars, darker head, wider vertebral stripe and narrower upper lateral stripe (modified from Storr 1991).

Species of the genus *Telchinoscincus* (Wells and Wellington, 1984) are separated from their closest related genus *Rhodona* Gray, 1839, by having a fixed, fused unmovable lower eyelid, versus a moveable one in *Rhodona.*

Distribution: *Telchinoscincus kendricki kalbarriensis subsp. nov.* is restricted to the Kalbarri area of Western Australia and while not under any known threat, is a range-restricted taxon.

The nominate form of *T. kendricki* has a distribution centred on Shark Bay, some 300 km to the north.

Etymology: *T. kendricki kalbarriensis subsp. nov.* is named from where this subspecies occurs.

TELCHINOSCINCUS ONSLOVIANA AH SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:C01BC2B3-A160-457D-8B3E-D6B072FBD7A2

Holotype: A preserved adult specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R157175 collected from Yanrey Station, Western Australia, Australia, Latitude -22.329444 S., Longitude 114.440278 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved subadult specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R157175 collected from Yanrey Station, Western Australia, Australia, Latitude -22.250278 S., Longitude 114.507778 E.

Diagnosis: *Telchinoscincus onsloviana ah subsp. nov.* a taxon apparently restricted to the eastern side of the Exmouth Gulf in Western Australia is readily separated from the nominate form of *Telchinoscincus onsloviana* (Storr, 1984), a taxon found further north in the vicinity of Onslow, extending along the coast about 40 km to the south in Western Australia, by having a preponderance of peppering on the top of the head as opposed to dark markings and spots, and a body and (original) tail that has an obscured vertebral line caused by brown pigment overrunning the brownish black spotted boundary, in turn reducing the relative width of the yellowish zone on the outer edges of the dorsum before the dorsolateral edge and the dark brown line occupying the upper part of the flank.

In nominate from of *Telchinoscincus onsloviana* the brownish black spotted boundary is also the boundary for the brown pigment in the vertebral line. Brown pigment does not overspill this.

The other subspecies *Telchinoscincus onsloviana har subsp. nov.* is separated from the previous two subspecies by having very little peppering or spots on the head, the head being mainly brown on top instead. The thick dark upper labial bands or triangles seen in the previous two subspecies are in *T. onsloviana har subsp. nov.* reduced to be thin etchings or absent all together. In adult *T. onsloviana har subsp. nov.* the vertebral stripe is very faint on the anterior half of the tail, versus well defined in the other two subspecies.

T. onsloviana har subsp. nov. is known from 14 km north of Giralia, to 2 km south of Marrilla, extending about 50 km east in the adjacent sandy zone, all in west, Western Australia, Australia.

The three preceding subspecies are separated from all other species within *Telchinoscincus* Wells and Wellington, 1984 by the following unique combination of characters:

Usually 2 supraciliaries (the first and second being fused to first supraocular), second loreal fused to prefrontal, and relatively narrow vertebral stripe (enclosing 2 series of dark spots) (modified from Storr 1984).

The type form of *Telchinoscincus onsloviana* is depicted in life in Wilson and Swan (2021) on page 377 middle right and online at:

https://www.flickr.com/photos/127392361@N04/54012359945/

and

https://www.flickr.com/photos/114192916@N07/54079865338/

all from Onslow, Western Australia

Telchinoscincus onsloviana ah subsp. nov. is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/54826802693/

from the Exmouth Gulf, Western Australia.

Species of the genus *Telchinoscincus* (Wells and Wellington, 1984) are separated from their closest related genus *Rhodona* Gray, 1839, by having a fixed, fused unmovable lower eyelid, versus a moveable one in *Rhodona.*

Distribution: *Telchinoscincus onsloviana ah subsp. nov.* is confined to the eastern edge of the Exmouth Gulf in Western Australia in an area immediately opposite the Cape Range.

Etymology: When the lizard is seen escaping in the sand, a common exclamation is the word “ah”.

TELCHINOSCINCUS ONSLOVIANA HAR SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:978D7F14-C542-43B5-8569-D21C29CD8AD9

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R63701 collected from the Marrilla Homestead, Western Australia, Australia, Latitude -22.966667 S., Longitude 114.466667 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R103290 collected from 2 km south of Marrilla Homestead, Western Australia, Australia, Latitude -22.983333 S., Longitude 114.45 E.

Diagnosis: *Telchinoscincus onsloviana ah subsp. nov.* a taxon apparently restricted to the eastern side of the Exmouth Gulf in Western Australia is readily separated from the nominate form of *Telchinoscincus onsloviana* (Storr, 1984), a taxon found further north in the vicinity of Onslow, extending along the coast about 40 km to the south in Western Australia, by having a preponderance of peppering on the top of the head as opposed to dark markings and spots, and a body and (original) tail that has an obscured vertebral line caused by brown pigment overrunning the brownish black spotted boundary, in turn reducing the relative width of the yellowish zone on the outer edges of the dorsum before the dorsolateral edge and the dark brown line occupying the upper part of the flank.

In nominate from of *Telchinoscincus onsloviana* the brownish black spotted boundary is also the boundary for the brown pigment in the vertebral line. Brown pigment does not overspill this.

The other subspecies *Telchinoscincus onsloviana har subsp. nov.* is separated from the previous two subspecies by having very little peppering or spots on the head, the head being mainly brown on top instead. The thick dark upper labial bands or triangles seen in the previous two subspecies are in *T. onsloviana har subsp. nov.* reduced to be thin etchings or absent all together. In adult *T. onsloviana har subsp. nov.* the vertebral stripe is very faint on the anterior half of the tail, versus well defined in the other two subspecies.

T. onsloviana har subsp. nov. is known from 14 km north of Giralia, to 2 km south of Marrilla, extending about 50 km east in the adjacent sandy zone, all in west, Western Australia, Australia.

The three preceding subspecies are separated from all other species within *Telchinoscincus* Wells and Wellington, 1984 by the following unique combination of characters:

Usually 2 supraciliaries (the first and second being fused to first supraocular), second

loreal fused to prefrontal, and relatively narrow vertebral stripe (enclosing 2 series

of dark spots) (modified from Storr 1984).

Species of the genus *Telchinoscincus* (Wells and Wellington, 1984) are separated from their closest related genus *Rhodona* Gray, 1839, by having a fixed, fused unmovable lower eyelid, versus a moveable one in *Rhodona.*

The type form of *Telchinoscincus onsloviana* is depicted in life in Wilson and Swan (2021) on page 377 middle right and online at:

https://www.flickr.com/photos/127392361@N04/54012359945/

and

https://www.flickr.com/photos/114192916@N07/54079865338/

all from Onslow, Western Australia

Telchinoscincus onsloviana ah subsp. nov. is depicted in life online at:

https://www.flickr.com/photos/127392361@N04/54826802693/

from the Exmouth Gulf, Western Australia.

Distribution: *T. onsloviana har subsp. nov.* is known from 14 km north of Giralia, to 2 km south of Marrilla,

extending about 50 km east in the adjacent sandy zone, all in west, Western Australia, Australia.

Etymology: When the lizard is grabbed before it escapes in the sand, a common exclamation is the word “har”.

TELCHINOSCINCUS PETERSONI FAAARK SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:6BF7A7E8-6DB3-486A-801C-91CD8C4E1261

Holotype: A preserved adult female specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R120514 collected from the Kennedy Range National Park, Western Australia, Australia, Latitude -24.551389 S., Longitude 114.958889 E.

This government-owned facility allows access to its holdings.

Diagnosis: Until now, no subspecies of *Telchinoscincus petersoni* (Storr, 1976) (a west Western Australian taxon) have been recognised since Aplin *et al.* (2022) synonymised “*Lerista talpina* Storr, 1991”, type locality Bullara, Western Australia, Australia (Holotype: WAM R74947) with “*L. petersoni* Storr, 1976”, type locality Yinnietharra, Western Australia, Australia, (Holotype: WAM R46197).

Singhal *et al*. (2025) in their supplementary data recognised four so-called “Operational species” within putative “*Lerista petersoni*”. On the basis of consistent morphological divergences and the fact that each relevant population is geographically isolated and separated from one another, with each being centred on a given range of hills, it is appropriate that each be given formal taxonomic recognition.

Nominate *Telchinoscincus petersoni* occupies most of the known range for the putative species, including the Mount Augustus area of Western Australia.

T. petersoni talpina appears to be a Cape Range endemic.

T. petersoni sheeta subsp. nov. is a taxon found in the Barlee Range.

T. petersoni faaark subsp. nov. is a taxon found in the Kennedy Range.

The four subspecies are separated from one another (as normal healthy adult specimens) by the following unique combinations of characters:

Nominate *Telchinoscincus petersoni* has 75 or more paravertebral scales (versus 74 or less in *T. petersoni talpina*), beige on the dorsum, with two rows of semi distinct brown spots on either side of the vertebral line, with the para-vertebral lines becoming well defined on the anterior of the (original) tail and along most of its length. The tail is more silvery-grey than the body. Labials are heavily barred, this being thick dark etching of the scales, with the intervening whites on the scales being wider than the dark. Yellow venter.

T. petersoni of the nominate form is depicted in life in Cogger (2014) on page 618 at top left and online at: https://www.flickr.com/photos/euprepiosaur/54567677407/

and

https://www.flickr.com/photos/127392361@N04/54842858848/

and

https://www.inaturalist.org/observations/287176171

T. petersoni talpina has 74 or less paravertebral scales, (versus 75 or more in the other three subspecies), heavily barred upper labials, making the white interspaces smaller than the dark, yellowish on the outer dorsum, including on the outer edges of the upper surface of the majority of the (original) tail. Cream venter.

T. petersoni talpina is depicted in life in Wilson and Swan (2021) on page 379 at top right and online at: https://www.flickr.com/photos/colonel_007/49103740923/

and

https://www.flickr.com/photos/colonel_007/49103740923/

T. petersoni faaark subsp. nov. has 75 or more paravertebral scales (versus 74 or less in *T. petersoni talpina*), light brown on the dorsum, with two rows of large expanded, irregularly shaped distinct brown spots on either side of the vertebral line, with the para-vertebral lines being well defined along most of the length of the body. At the lower part of the body and the anterior tail, the spots wholly merge to form dark brown paravertebral lines and this in turn is infilled with dark brown of similar intensity, in effect forming a single thick vertebral stripe running down the rear of the body and onto the (original) tail. This stripe declines in intensity along the tail and completely fades by the latter part, this being light brown in colour instead. The first four upper labials are mainly brown. The white is reduced to a tiny patch in the lower centre of each, being rectangular, triangular, or otherwise irregularly shaped. These scales are not in an obviously barred configuration. Yellow venter.

T. petersoni faaark subsp. nov. is depicted in life online at: https://www.flickr.com/photos/195006528@N02/54824139117/

T. petersoni sheeta subsp. nov. has 75 or more paravertebral scales (versus 74 or less in *T. petersoni talpina*) and is similar in most respects to nominate *T. petersoni* as described above, but with few if any dark flecks or scale etchings on the top of the head, more yellowish than beige on the dorsum, including the tail and only thin dark etching of the upper labial scales. Cream venter.

The four preceding subspecies are separated from

all other species within *Telchinoscincus* Wells and Wellington, 1984 and all other species within *Lerista sensu lato sensu* Hoser (2024c) or Cogger (2014) by the following unique combination of characters:

A wider paravertebral stripe, 4 supraciliaries (versus 3 or less), Nasals in very short to moderately long contact. Supraoculars 3, first two in contact with frontal. Loreals 2, second much smaller than first. Presuboculars very small. Upper labials 5, when third and fourth fused, or more often simply 6.

1-3 nuchals and 18 to 22 (usually 20) midbody rows. 101-14 lamellae under the fourth toe.

Forelimb represented by a small groove or pit and usually a very small stump (up to 0.5 mm long).

Upper surface brownish white, beige, yellowish, to brown. Head, especially sutures, usually irregularly marked with dark brown etchings and tiny flecks (mainly at the rear of the head). Back with 4 (closely placed) (occasionally 2) longitudinal rows of small angular dark brown or blackish brown spots of irregular shape, the space between them usually pale greyish brown or yellowish, the resultant stripe extending on to tail in varying form depending on the subspecies. Wide dark brown or blackish brown upper lateral stripe from nasal to end of tail, on body usually occupying a longitudinal row of scales and lower half of row immediately above it, and continuous with vertical bars or dark on the upper labials (mostly located on the sutures between the upper labials). Upper surface of hindlegs mottled with greyish brown or blackish brown. Lower lateral surface and sometimes ventral surface is cream or yellow in life (modified from Storr, 1971).

Distribution: *T. petersoni faaark subsp. nov.* is a taxon found in the Kennedy Range, Western Australia, Australia. Although range-restricted, it is not believed to be under any imminent threat.

Etymology: In 1981, when grabbing a specimen of this species on the run from a burning spinifex bush the Malgaru child grabbed a specimen and cut off its tail. He held it in the air and yelled “Faaark”.

TELCHINOSCINCUS PETERSONI SHEETA SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:C7CF37EF-FC16-49BC-9B09-E3E7EBB16FB6

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R102455 collected from the Barlee Range Nature Reserve, Western Australia, Australia, Latitude -23.113056 S., Longitude 116.011667 E.

This government-owned facility allows access to its holdings.

Paratypes: Four preserved specimens at the Western Australian Museum, Perth, Western Australia, Australia, specimen numbers R102277, R102278, R102457 and R102509, collected from the Barlee Range Nature Reserve, Western Australia, Australia, Latitude -23.113056 S., Longitude 116.011667 E.

Diagnosis: Until now, no subspecies of *Telchinoscincus petersoni* (Storr, 1976) (a west Western Australian taxon) have been recognised since Aplin *et al.* (2022) synonymised “*Lerista talpina* Storr, 1991”, type locality Bullara, Western Australia, Australia (Holotype: WAM R74947) with “*L. petersoni* Storr, 1976”, type locality Yinnietharra, Western Australia, Australia, (Holotype: WAM R46197).

Singhal *et al*. (2025) in their supplementary data recognised four so-called “Operational species” within putative “*Lerista petersoni*”. On the basis of consistent morphological divergences and the fact that each relevant population is geographically isolated and separated from one another, with each being centred on a given range of hills, it is appropriate that each be given formal taxonomic recognition.

Nominate *Telchinoscincus petersoni* occupies most of the known range for the putative species, including the Mount Augustus area of Western Australia.

T. petersoni talpina appears to be a Cape Range endemic.

T. petersoni sheeta subsp. nov. is a taxon found in the Barlee Range.

T. petersoni faaark subsp. nov. is a taxon found in the Kennedy Range.

The four subspecies are separated from one another (as normal healthy adult specimens) by the following unique combinations of characters:

Nominate *Telchinoscincus petersoni* has 75 or more paravertebral scales (versus 74 or less in *T. petersoni talpina*), beige on the dorsum, with two rows of semi distinct brown spots on either side of the vertebral line, with the para-vertebral lines becoming well defined on the anterior of the (original) tail and along most of its length. The tail is more silvery-grey than the body. Labials are heavily barred, this being thick dark etching of the scales, with the intervening whites on the scales being wider than the dark. Yellow venter.

T. petersoni of the nominate form is depicted in life in Cogger (2014) on page 618 at top left and online at: https://www.flickr.com/photos/euprepiosaur/54567677407/

and

https://www.flickr.com/photos/127392361@N04/54842858848/

and

https://www.inaturalist.org/observations/287176171

T. petersoni talpina has 74 or less paravertebral scales, (versus 75 or more in the other three subspecies), heavily barred upper labials, making the white interspaces smaller than the dark, yellowish

on the outer dorsum, including on the outer edges of the upper surface of the majority of the (original) tail. Cream venter.

T. petersoni talpina is depicted in life in Wilson and Swan (2021) on page 379 at top right and online at: https://www.flickr.com/photos/colonel_007/49103740923/

and

https://www.flickr.com/photos/colonel_007/49103740923/

T. petersoni faaark subsp. nov. has 75 or more paravertebral scales (versus 74 or less in *T. petersoni talpina*), light brown on the dorsum, with two rows of large expanded, irregularly shaped distinct brown spots on either side of the vertebral line, with the para-vertebral lines being well defined along most of the length of the body. At the lower part of the body and the anterior tail, the spots wholly merge to form dark brown paravertebral lines and this in turn is infilled with dark brown of similar intensity, in effect forming a single thick vertebral stripe running down the rear of the body and onto the (original) tail. This stripe declines in intensity along the tail and completely fades by the latter part, this being light brown in colour instead. The first four upper labials are mainly brown. The white is reduced to a tiny patch in the lower centre of each, being rectangular, triangular, or otherwise irregularly shaped. These scales are not in an obviously barred configuration. Yellow venter.

T. petersoni faaark subsp. nov. is depicted in life online at: https://www.flickr.com/photos/195006528@N02/54824139117/

T. petersoni sheeta subsp. nov. has 75 or more paravertebral scales (versus 74 or less in *T. petersoni talpina*) and is similar in most respects to nominate *T. petersoni* as described above, but with few if any dark flecks or scale etchings on the top of the head, more yellowish than beige on the dorsum, including the tail and only thin dark etching of the upper labial scales. Cream venter.

The four preceding subspecies are separated from all other species within *Telchinoscincus* Wells and Wellington, 1984 and all other species within *Lerista sensu lato sensu* Hoser (2024c) or Cogger (2014) by the following unique combination of characters:

A wider paravertebral stripe, 4 supraciliaries (versus 3 or less), Nasals in very short to moderately long contact. Supraoculars 3, first two in contact with frontal. Loreals 2, second much smaller than first. Presuboculars very small. Upper labials 5, when third and fourth fused, or more often simply 6.

1-3 nuchals and 18 to 22 (usually 20) midbody rows. 101-14 lamellae under the fourth toe.

Forelimb represented by a small groove or pit and usually a very small stump (up to 0.5 mm long).

Upper surface brownish white, beige, yellowish, to brown. Head, especially sutures, usually irregularly marked with dark brown etchings and tiny flecks (mainly at the rear of the head). Back with 4 (closely placed) (occasionally 2) longitudinal rows of small angular dark brown or blackish brown spots of irregular shape, the space between them usually pale greyish brown or yellowish, the resultant stripe extending on to tail in varying form depending on the subspecies. Wide dark brown or blackish brown upper lateral stripe from nasal to end of tail, on body usually occupying a longitudinal row of scales and lower half of row immediately above it, and continuous with vertical bars or dark on the upper labials (mostly located on the sutures between the upper labials). Upper surface of hindlegs mottled with greyish brown or blackish brown. Lower lateral surface and sometimes ventral surface is cream or yellow in life (modified from Storr, 1971).

Distribution: *T. petersoni sheeta subsp. nov.* is a taxon found in the Barlee Range, Western Australia, Australia. Although range-restricted, it is not believed to be under any imminent threat.

Etymology: In 1981, when grabbing a specimen of this species on the run in the loose sand the Thudgari child got burs in his hand and exclaimed “Sheeta”.

TISM KALUMBURU BESTOFF SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:45139FD5-88CE-4FA9-8A26-C22DB1DE6117

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R113949 collected from the Carson’s Escarpment, Western Australia, Australia, Latitude -14.8 S., Longitude 126.8 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R113983 collected from the Carson’s Escarpment, Western Australia, Australia, Latitude -15.4 S., Longitude 126.6 E.

Diagnosis: *Tism kalumburu bestoff subsp. nov*. is known only from the Carson’s Escarpment, Western Australia, Australia.

Tism kalumburu bestoff subsp. nov. is separated from the other two subspecies of *Tism kalumburu* (Storr, 1976) by the following character combination:

Having three toes instead of two; is a relatively dark brownish grey on top and with strong a reddish-orange hue at the distal end of the tail and there are no longitudinal stripes formed by spots on the flanks.

Tism kalumburu whatareya subsp. nov. is known only from the vicinity of Theda Station in the north Kimberley District of Western Australia.

It is readily separated from all other subspecies of

Tism kalumburu (Storr, 1976) by having two toes instead of three.

In terms of dorsal pattern, adults have reduced spotting forming longitudinal lines and a strong yellowish sheen in the colour, including the length of the tail, these traits not seen in the other two subspecies.

Tism kalumburu of the nominate subspecies is from the Kalumburu area and near coastal environs of Western Australia.

Tism kalumburu of the nominate subspecies is a brownish, rather than grey colour on top, with a slight purplish tinge and reasonably well-defined longitudinal lines on the upper surfaces and flanks.

The tail is the same colour as the body, save for a greater amount of darker pigment.

The genus *Tism gen. nov.* monotypic at the species level, with a type species of *Lerista kalumburu* Storr, 1976 is separated from all other species within the putative genus *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982*, Aphroditia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012*,* type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012*,* type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012*,* type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012*,* type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012*,* type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012*,* type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012*,* type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, (obviously not including *Tism* Hoser, 2024 in this case), *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

Small obscurely spotted sliding skink with digits 1 + 3 or 1 + 2 and a movable eyelid. Average adult snout-vent length is 50 mm. Length of appendages, etc. as approximate percentage of snout-vent length: foreleg 2 percent; hindleg 13 percent; tail 73 percent; snout to foreleg 25 percent.

Nasals forming a long median suture. Prefrontals widely separated.

Frontoparietals separated, and very much smaller than the interparietal. 2-3 nuchals.

3 supraoculars, first two contacting the frontal. 5 supraciliaries; first and fourth the largest; second, third and fifth the smallest. Upper secondary temporal largest, lower secondary is far smaller. 18 midbody rows; 8-9 lamellae under the longest toe.

Dorsal surface is a ground colour, ranging from yellowish, brown-grey, and often with a purplish tinge, each dorsal scale with a central brown dark spot (often reduced in size to be specks); spots on flanks often tending to coalesce into obscure longitudinally orientated lines. There is more dark in the scales than light on the tail, making the tail appearing to be darker than the body. Lips are whitish, barred with a purplish dark brown. Belly whitish, sparsely spotted with pale brown towards tip of tail (modified and amended from Storr 1976).

The monotypic species, *Tism kalumburu* (Storr, 1976) is readily distinguishable from the morphologically similar Kimberley taxa *Marrunisauria walkeri* (Boulenger, 1891), *M. borealis* (Storr, 1971), *M. wam sp. nov*., *Gaia oomph sp. nov.*, *G. griffini* (Storr, 1982) and associated species by having the combination of smaller frontoparietals, larger primary temporal and fewer rows of midbody scales (18, versus 20 (rarely 19 or 21) in all the other species); less heavily spotted under the tail, and the supraciliaries are less disparate in size from front to back.

Distribution: *Tism kalumburu bestoff subsp. nov*. is known only from the Carson's Escarpment, Western Australia, Australia.

Tism kalumburu of the nominate form is from the Kalumburu area and near coastal environs of Western Australia.

Tism kalumburu whatareya subsp. nov. is known only from the vicinity of Theda Station in the north Kimberley District of Western Australia.

Etymology: "Best off" was a popular TISM album that was released in 2002. TISM was an iconic Australian band playing around the turn of the century.

TISM KALUMBURU WHATAREYA SUBSP. NOV.

LSIDurn:lsid:zoobank.org:act:108410A5-DEC8-4E7D-AC51-E1F8D0DB374C

Holotype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R172358 collected from Theda Station in Western Australia, Australia, Latitude -14.8 S., Longitude 126.5 E.

This government-owned facility allows access to its holdings.

Paratype: A preserved specimen at the Western Australian Museum, Perth, Western Australia, Australia, specimen number R172346 collected from Theda Station in Western Australia, Australia, Latitude -14.8 S., Longitude 126.5 E.

Diagnosis: *Tism kalumburu whatareya subsp. nov*. is known only from the vicinity of Theda Station in the north Kimberley District of Western Australia.

It is readily separated from all other subspecies of *Tism kalumburu* (Storr, 1976) by having two toes instead of three.

In terms of dorsal pattern, adults have reduced spotting forming longitudinal lines and a strong yellowish sheen in the colour, including the length of the tail, these traits not seen in the other two subspecies.

Tism kalumburu bestoff subsp. nov. is known only from the Carson's Escarpment, Western Australia, Australia.

Tism kalumburu bestoff subsp. nov. in turn is a relatively dark brownish grey on top and with strong a reddish-orange hue at the distal end of the tail. There are no longitudinal stripes formed by spots on the flanks.

Tism kalumburu of the nominate subspecies is from the Kalumburu area and near coastal environs of Western Australia.

Tism kalumburu of the nominate subspecies is a brownish, rather than grey colour on top, with a slight purplish tinge and reasonably well-defined longitudinal lines on the upper surfaces and flanks.

The tail is the same colour as the body, save for a greater amount of darker pigment.

The genus *Tism gen. nov.* monotypic at the species level, with a type species of *Lerista kalumburu* Storr, 1976 is separated from all other species within the putative genus *Lerista sensu lato* this including all other species within the putative genera *Lerista* Bell, 1833, type species *Lerista lineata* Bell, 1833, *Rhodona* Gray, 1839, type species *Rhodona punctata* Gray, 1839, *Soridia* Gray, 1839, type species *Soridia lineata* Gray, 1839, *Miculia* Gray, 1845, type species *Miculia elegans* Gray, 1845, *Phaneropis* Fischer, 1881, type species *Phaneropis muelleri* Fischer, 1881, *Nodorha* Mittleman, 1952, type species *Riopa boulengeri* Duméril and Bibron ,1839, *Gavisus* Wells and Wellington, 1984, type species: *Lygosoma* (*Rhodona*) *wilkinsi* Parker, 1926, *Telchinoscincus* Wells and Wellington, 1984, type species *Rhodona nichollsi* Loveridge, 1933, *Xynoscincus* Wells and Wellington, 1988, type species *Lerista stictopleura* Storr, 1985, *Alcisius* Wells, 2012, type species, *Lerista vermicularis* Storr 1982, *Aphroditia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *macropisthopus* Werner, 1903, *Cybelia* Wells, 2012, type species *Lygosoma* (*Rhodona*) *terdigitatum* Parker, 1926, *Gaia* Wells, 2012, type species, *Rhodona bipes* Fischer, 1882, *Goldneyia* Wells, 2012, type species, *Rhodona planiventralis* Lucas and Frost, 1902, *Krishna* Wells, 2012, type species *Rhodona fragilis* Günther, 1876, *Lokisaurus* Wells, 2012, type species: *Ablepharus timidus* De Vis, 1888, *Marrunisauria* Wells, 2012, type species, *Lerista borealis* Storr, 1971, *Spectrascincus* Wells, 2012, type species, *Lerista ingrami* Storr, 1991, *Tychismia* Wells, 2012, type species *Lerista chordae* Amey, Kutt and Hutchinson 2005 and *Wondjinia* Wells, 2012, type species, *Lerista apoda* Storr, 1976, all as defined by Wells (2012) (or alternatively including those in the eight additional genera erected by Hoser 2024c, being *Aaah* Hoser, 2024, *Ah* Hoser, 2024, *Oh* Hoser, 2024, *Tism* Hoser, 2024, *Acdc* Hoser, 2024, *Labi* Hoser, 2024, *Go* Hoser, 2024 and *Get* Hoser, 2024) and assuming placement of all relevant species within these genera, by the following unique combination of characters:

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Nasals forming a long median suture. Prefrontals widely separated.

Frontoparietals separated, and very much smaller than the interparietal. 2-3 nuchals.

3 supraoculars, first two contacting the frontal. 5 supraciliaries; first and fourth the largest; second, third and fifth the smallest. Upper secondary temporal largest, lower secondary is far smaller. 18 midbody rows; 8-9 lamellae under the longest toe.

Dorsal surface is a ground colour, ranging from yellowish, brown-grey, and often with a purplish tinge, each dorsal scale with a central brown dark spot (often reduced in size to be specks); spots on flanks often tending to coalesce into obscure longitudinally orientated lines. There is more dark in the scales than light on the tail, making the tail appearing to be darker than the body. Lips are whitish, barred with a purplish dark brown. Belly whitish, sparsely spotted with pale brown towards tip of tail (modified and amended from Storr 1976).

The monotypic species, *Tism kalumburu* (Storr, 1976) is readily distinguishable from the morphologically similar Kimberley taxa *Marrunisauria walker* (Boulenger, 1891), *M. borealis* (Storr, 1971), *M. wam sp. nov.*, *Gaia oomph sp. nov.*, *G. griffini* (Storr, 1982) and associated species by having the combination of smaller frontoparietals, larger primary temporal and fewer rows of midbody scales (18, versus 20 (rarely 19 or 21) in all the other species); less heavily spotted under the tail, and the supraciliaries are less disparate in size from front to back.

Distribution: *Tism kalumburu whatareya subsp. nov*. is known only from the vicinity of Theda Station in the north Kimberley District of Western Australia.

Tism kalumburu bestoff subsp. nov. is known only from the Carson's Escarpment, Western Australia, Australia.

Tism kalumburu of the nominate form is from the Kalumburu area and near coastal environs in Western Australia.

Etymology: "Whatareya?" Was a single released by the iconic Australian rock band, TISM.

It was a hilarious and biting take on Australian social identities and working-class culture.

"Whatareya?" was released in 1998. Specifically, it was issued on July 6, 1998, as the third single from TISM's fourth studio album, www.tism.wanker.com. The track went on to reach number 66 on the ARIA Charts and ranked at number 36 in the Australian Federal Government-owned ABC Radio Triple J's Hottest 100 for that year.

CONSERVATION

For all species referred to in this paper, the comments in Hoser (2024c) apply herein.

Also relevant to the ongoing conservation of the preceding species are the nomenclatural comments in line with the choices of new names for the relevant taxa as detailed in Hoser (2024c) and sources cited therein.

Those comments are adopted and apply herein.

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CONFLICT OF INTEREST

None.

CURRENT GENUS AND SPECIES LIST FOR *LERISTA* BELL, 1833 *SENSU LATO*

Genera are listed in alphabetical order. Genera are same as listed in (Hoser, 2024c). Type species for each genus, using the ICZN compliant priority species name is listed first, then rest are alphabetical. This list does not include the dozens of known and recognised subspecies or available synonyms for the relevant species.

AAAH GEN. NOV.
Aaah edwardsae (Storr, 1982)
Aaah baynesi (Storr, 1972)
Aaah picturata (Fry, 1914)
ACDC GEN. NOV.
Acdc zietzi (Wells and Wellington, 1985)
Acdc flammicauda (Storr, 1985)
Acdc thunderstruck sp. nov.
Acdc tnt sp. nov.
AH GEN. NOV.
Ah ha Hoser, 2024
Ah christinae (Storr, 1979).
Ah yess sp. nov.
Ah pfuk sp. nov.
Ah sheat sp. nov.
***APHRODITIA* WELLS, 2012**
Aphroditia macropisthopus (Werner, 1903)
Aphroditia axillaris (Storr, 1991)
Aphroditia bunglebungle (Storr, 1991)
Aphroditia desertorum (Sternfeld, 1919)
Aphroditia eupoda (Smith, 1996)
Aphroditia fusciceps (Storr, 1991)
Aphroditia galea (Storr, 1991)
Aphroditia gerrardii (Gray, 1864)
Aphroditia harriganorum sp. nov.
Aphroditia neander (Storr, 1971)
Aphroditia puncticauda (Storr, 1991)
Aphroditia remota (Storr, 1991)
***CYBELIA* WELLS, 2012**
Cybelia terdigitatum (Parker, 1926)
Cybelia elongata (Storr, 1990)
Cybelia speciosa (Storr, 1990
Cybelia tridactyla (Storr, 1990)
***GAIA* WELLS, 2012**
Gaia bipes (Fischer, 1882)
Gaia amazing sp. nov.
Gaia anangu sp. nov.
Gaia arrernte Hoser, 2024
Gaia aviyemenii sp. nov.
Gaia dezifreemani sp. nov.
Gaia greeri (Storr, 1982)
Gaia griffini (Storr, 1982
Gaia ips (Storr, 1980)
Gaia kunja Hoser, 2024
Gaia labialis (Storr, 1971)
Gaia liesarenotscience sp. nov.
Gaia narcissismawareness sp. nov.
Gaia nomenclaturematters sp. nov.
Gaia nottobemissed sp. nov.
Gaia oomph Hoser, 2024
Gaia owe sp. nov.
Gaia otit sp. nov.
Gaia pitjantjatjara Hoser, 2024
Gaia praefrontalis (Greer, 1986)
Gaia questionthenarrative sp. nov.
Gaia rate sp. nov.
Gaia robusta (Storr, 1990)
Gaia robwatsoni sp. nov.
Gaia rolloi (Wells and Wellington, 1985)
Gaia ruleofpriority sp. nov.
Gaia simillima (Storr, 1984)
Gaia taxonomymatters sp. nov.
Gaia truthisscience sp. nov.
Gaia vellai sp. nov.
Gaia vermicularis (Storr, 1982)
Gaia wow sp. nov.
***GAVISUS* WELLS AND WELLINGTON, 1984**
Gavisus wilkinsi (Parker, 1926)
*Gavisus alia (*Amey, Couper and Worthington-Wilmer, 2019)
Gavisus allanae (Longman, 1937)
Gavisus ameles (Greer, 1979)
Gavisus anyara (Amey, Couper and Worthington-Wilmer, 2019)
Gavisus cinereus (Greer, McDonald and Lawrie, 1983)
Gavisus colliveri (Couper and Ingram, 1992)
Gavisus hobsoni (Couper, Amey and Worthington-Wilmer, 2016)
Gavisus karichigara (Zozaya, Vanderduys, Macor, Read and Amey, 2025)
Gavisus karlschmidti (Marx and Hosmer, 1959)
*Gavisus munuwajarlu (*Zimny, Vanderduys, Kemp and Zozoya, 2025)
Gavisus parameles (Amey, Couper and Worthington-Wilmer, 2019)
Gavisus rochfordensis (Amey and Couper, 2009)
Gavisus storri (Greer, McDonald and Lawrie, 1983)
Gavisus vanderduysi (Couper, Amey and Worthington-Wilmer, 2016)
Gavisus vittatus (Greer, McDonald and Lawrie, 1983)
GET GEN. NOV.
Get it Hoser, 2024
Get carpentariae (Greer, 1983)
Get intoit Hoser, 2024
Get stylis (Mitchell, 1955)
GO GEN. NOV.
Go punctatovittata (Günther, 1867)
Go colliveri (Couper and Ingram, 1992)
Go emmotti (Ingram, Couper and Donnellan, 1993)
Go forit sp. nov.

***GOLDNEYIA* WELLS, 2012**
Goldneyia planiventralis (Lucas and Frost, 1902)
Goldneyia decora (Storr, 1978)
Goldneyia maryani (Storr, 1991)
LABI GEN. NOV.
Labi microtis (Gray, 1845)
Labi arenicola (Storr, 1971)
Labi schwaneri (Storr, 1991)
***LERISTA* BELL, 1833**
Lerista lineata Bell, 1833.
***MARRUNISAURIA* WELLS, 2012**
Marrunisauria borealis (Storr, 1971)
Marrunisauria walkeri (Boulenger, 1891)
Marrunisauria wam Hoser, 2024
***MICULIA* GRAY, 1845**
Miculia elegans Gray, 1845
Miculia aericeps (Storr, 1986)
Miculia chrismartini sp. nov.
Miculia christinae (Storr, 1979)
Miculia coldplayorum sp. nov.
Miculia distinguenda (Werner, 1910)
Miculia puseyi sp. nov.
Miculia ruficauda Hoser, 2024
Miculia separanda (Storr, 1976)
Miculia taeniata (Storr, 1986)
Miculia vacopoulosae sp. nov.
Miculia xanthura (Storr, 1976)
***NODORHA* MITTLEMAN, 1952**
Nodorha bougainvillii (Duméril and Bibron, 1839)
Nodorha absconditus Hoser, 2023
Nodorha cassandrae Wells and Wellington, 1984
Nodorha divergans Hoser, 2023
Nodorha hoserae Hoser, 2023
Nodorha insularis Hoser, 2023
Nodorha martinekae Hoser, 2023
Nodorha tasmaniensis Hoser, 2023
Nodorha viduata (Storr, 1991)
OH GEN. NOV.
Oh kay Hoser, 2024
Oh dorsalis (Storr, 1985)
Oh frosti (Zietz, 1895)
Oh know Hoser, 2024
Oh phuk Hoser, 2024
Oh sheet Hoser, 2024
Oh yes Hoser, 2024
***PHANEROPIS* FISCHER, 1881**
Phaneropis muelleri Fischer, 1881
Phaneropis abscondeetus sp. nov.
Phaneropis amicorum (Smith and Adams, 2007)
Phaneropis allochirus (Kendrick, 1989)
Phaneropis clarus (Smith and Adams, 2007)
Phaneropis dorsei sp. nov.
Phaneropis faaarkinelle sp. nov.
Phaneropis gottabekidding sp. nov.
Phaneropis greatfind sp. nov.
Phaneropis haroldi (Storr, 1983)
Phaneropis jacksoni (Smith and Adams, 2007)
Phaneropis kingi (Smith and Adams, 2007)
Phaneropis laylawangae sp. nov.
Phaneropis lopezi sp. nov.
Phaneropis micra (Smith and Adams, 2007)
Phaneropis nevinae (Smith and Adams, 2007)
Phaneropis occultus (Smith and Adams, 2007)
Phaneropis overlookedit sp. nov.
Phaneropis oxyi sp. nov.
Phaneropis rhodonoides (Lucas and Frost, 1896)
Phaneropis rolfei (Smith and Adams, 2007)
Phaneropis timidus (De Vis, 1888)
Phaneropis verhmens (Smith and Adams, 2007)
Phaneropis wholeeecowe sp. nov.
***RHODONA* GRAY, 1839**
Rhodona lineopunctulata (Dumeril and Bibron, 1839)
Rhodona connivens (Storr, 1971)
Rhodona varia (Storr, 1986)
***SORIDIA* GRAY, 1839**
Soridia praepedita (Boulenger, 1887)
Soridia humphriesi (Storr, 1971)
Soridia luxflavo Hoser, 2024
***SPECTRASCINCUS* WELLS, 2012**
Spectrascincus ingrami (Storr, 1991)
Spectrascincus hit Hoser, 2024
Spectrascincus orientalis (De Vis, 1889)
Spectrascincus thingi Hoser, 2024
Spectrascincus zonulatus (Storr, 1991)
***TELCHINOSCINCUS* WELLS AND WELLINGTON, 1984**
Telchinoscincus nichollsi (Loveridge, 1933)
Telchinoscincus calistaclementsae sp. nov.
Telchinoscincus gascoynensis (Storr, 1986)
Telchinoscincus kendricki (Storr, 1991)
Telchinoscincus kennedyensis (Kendrick, 1989)
Telchinoscincus onsloviana (Storr, 1984)
Telchinoscincus petersoni (Storr, 1976)
Telchinoscincus uniduo (Storr, 1984)
Telchinoscincus yuna (Storr, 1991)
TISM GEN. NOV.
Tism kalumburu (Storr, 1976)
***TYCHISMIA* WELLS, 2012**
Tychismia chordae (Amey, Kutt and Hutchinson 2005)
Tychismia concolor (Storr, 1990)
Tychismia fragilis (Gunther, 1876)
Tychismia valentici Hoser, 2024
Tychismia wellsei Hoser, 2024
***WONDJINIA* WELLS, 2012**
Wondjinia apoda (Storr, 1976)
***XYNOSCINCUS* WELLS AND WELLINGTON, 1988**
Xynoscincus stictopleurus (Storr, 1985)

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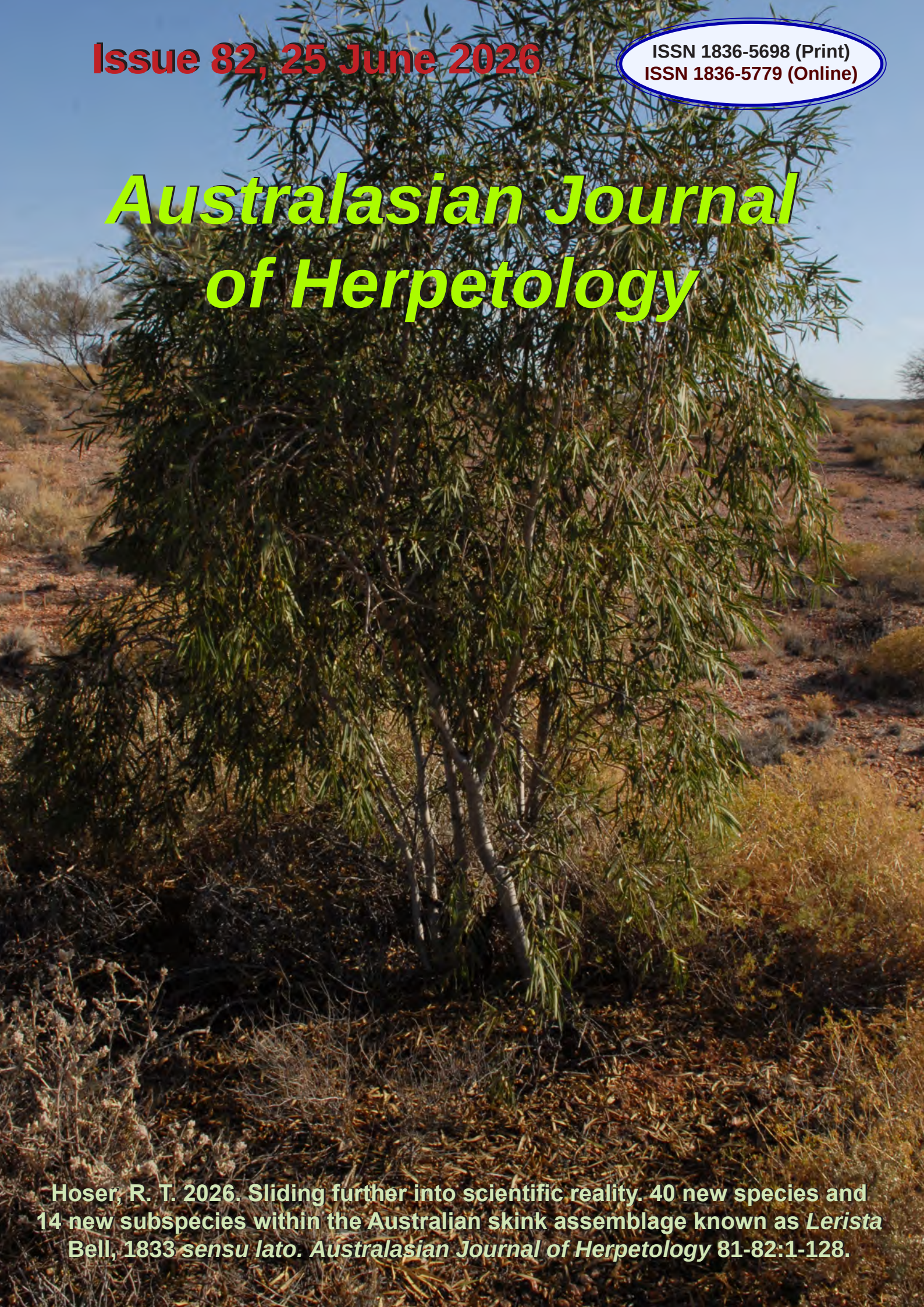

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